

Instrumentation, Automation and Standards in Structural Biology

Gergely PAPP

Interim Head – EMBL Grenoble Instrumentation Team



EMBL Grenoble Instrumentation Team - Focus



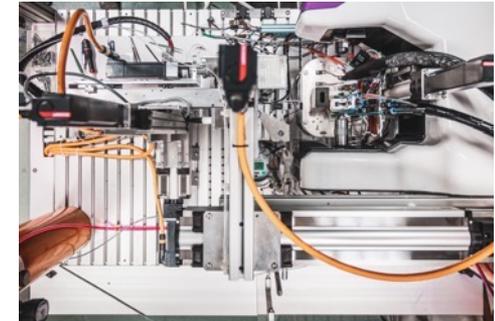
- Sample Environment of MX and SAXS beamlines
 - Diffractometers (20 years)
 - Automated sample changers (15 years)
 - Automated crystal harvesting CrystalDirect™ (10 years)
 - Sample holder standards & tracking solutions (15 years)
- Preparation of Cryo-EM sample grids (2017)

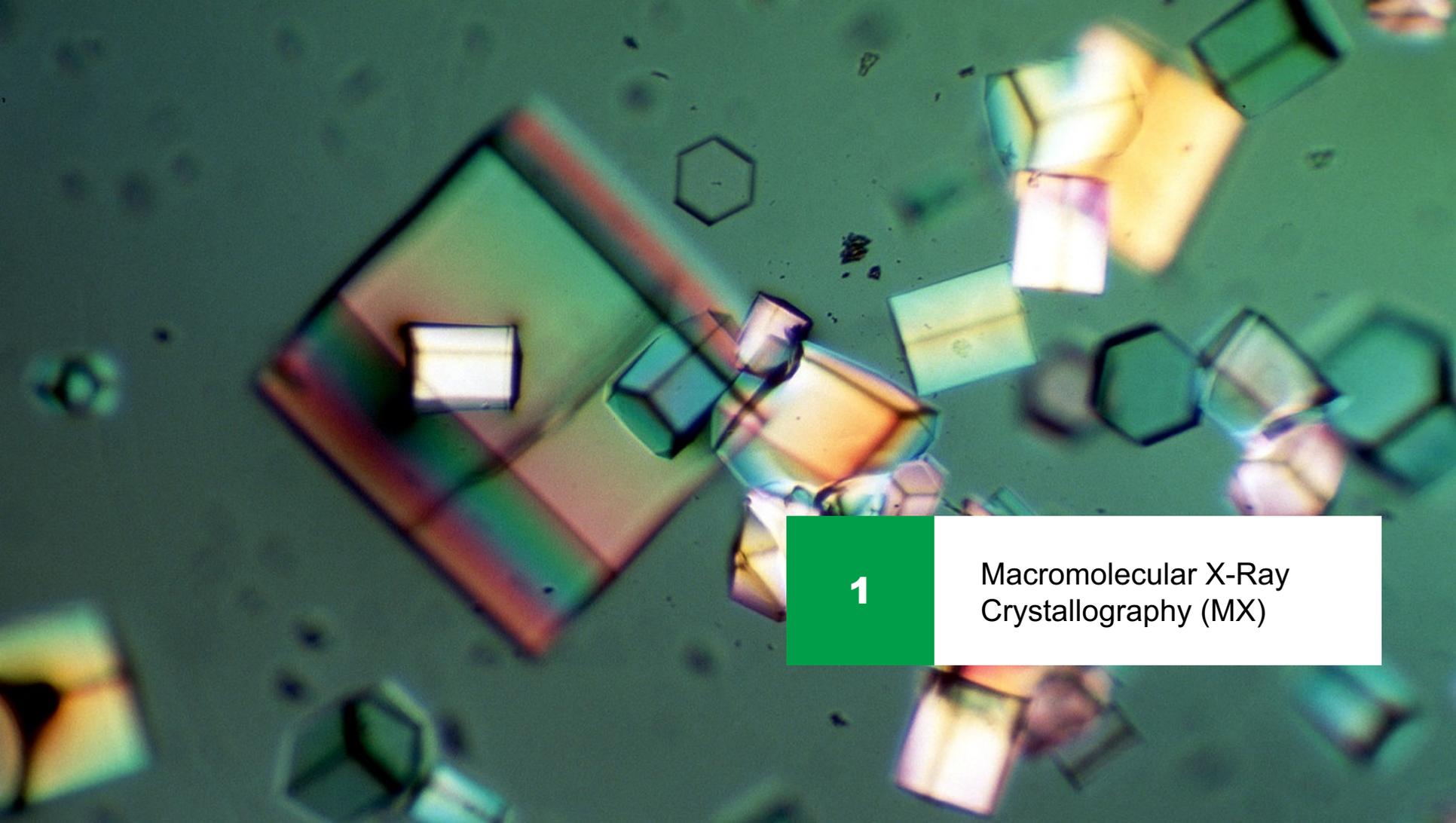
Our objectives

- Develop tools **from scientific demand** that are not available on the market
- Make them available for the entire scientific community by the bias of industrial partners
- Scientific collaborations are essential for our group:
 - EMBL-GR Crystallographers
 - ESRF Structural Biology Group
 - EMBL – Hamburg Crystallographers
 - EMBL – Heidelberg Cryo Electron microscopists
- As well as collaboration with industry
 - ARINAX
 - MiTeGen
 - MolecularDimensions
 - ...

Expertise – System Engineering

- Precision mechanics
- Cryogenics
- Analog/Digital Electronics
- Motion control
- Software engineering

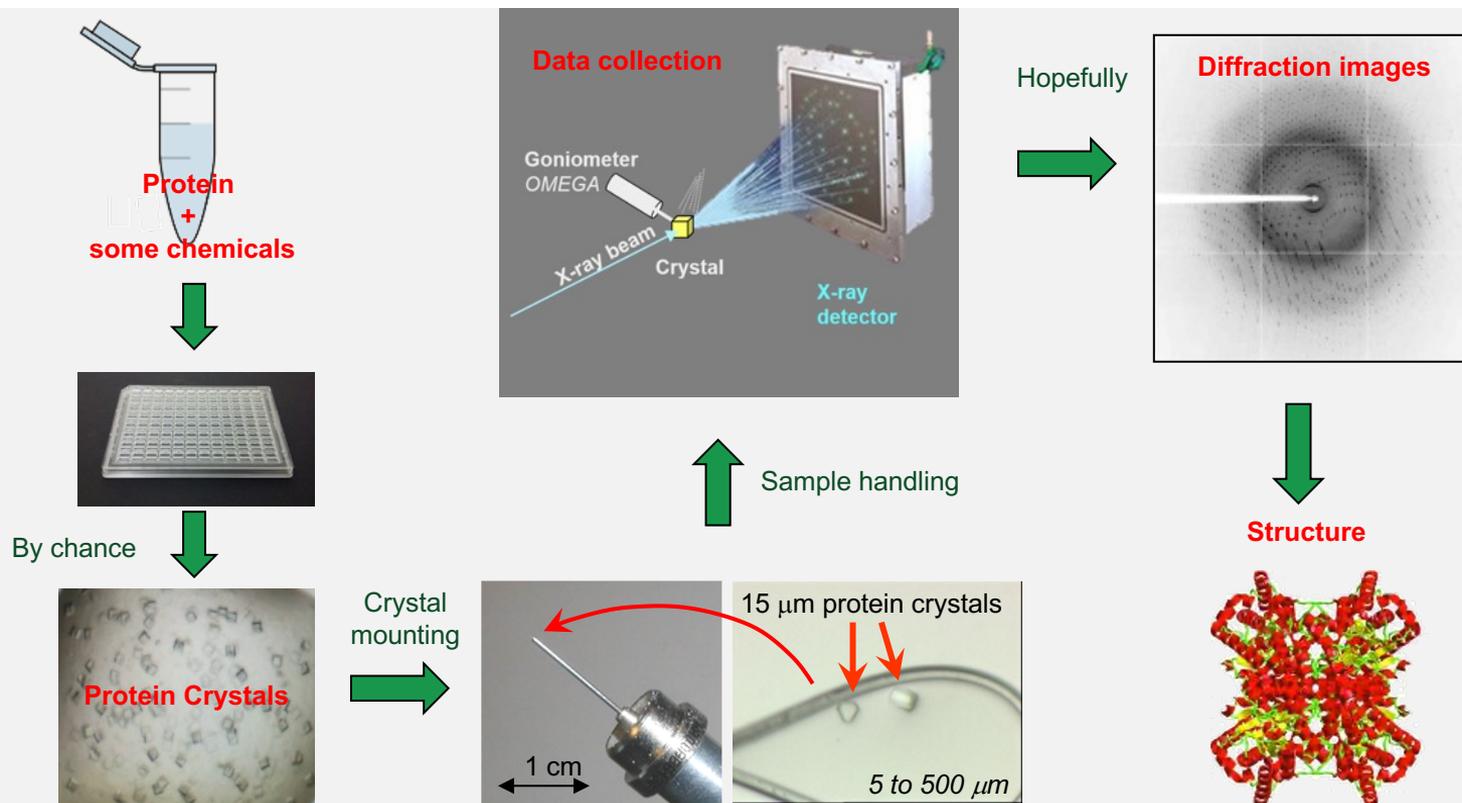




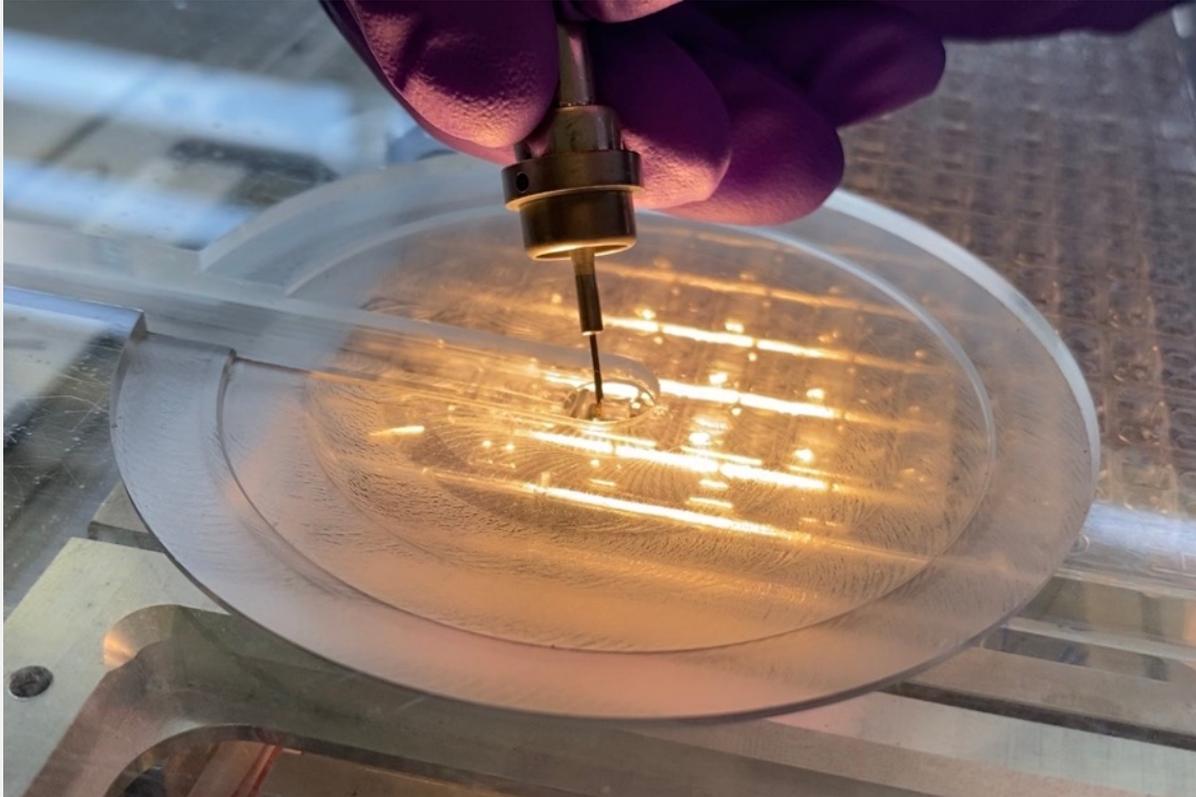
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Macromolecular X-Ray
Crystallography (MX)

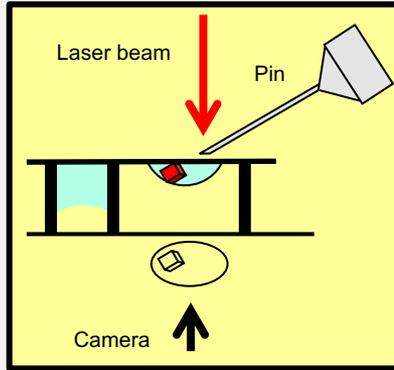
MX in brief



Crystal Harvesting – The traditional way

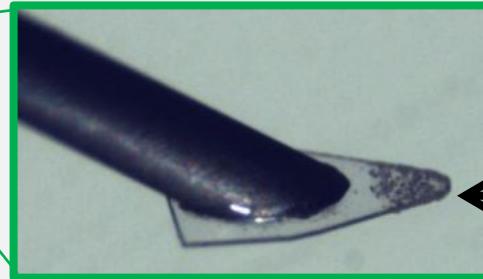


CrystalDirect: Harvesting by laser photoablation (2010)



Film with crystal **cut** with a **femtosecond** laser beam

Automatically mounted on a **SPINE** sample holder



Crystal(s)

CrystalDirect : Harvesting video

The screenshot displays the CrystalDirect software interface. The main window shows a microscope view of a crystal with a red dashed outline. The interface includes a menu bar (File, Devices, Help), a toolbar with buttons for Restart, Focus, Zoom, and Abort, and a bottom panel with various control panels.

Monitoring Panel:

- General Hardware Status: OK
- Laser: Ready
- Cryo: 99.9

Processing Plan Panel:

Crystal ID	Position	State
69749843...	A8-3	Needs repository...
71178261...	B2-3	Needs repository...
21066382...	B3-3	Needs repository...
17214176...	E5-3	Needs repository...
16295764...	F4-2	Needs repository...
40380573...	F1-3	Needs repository...
15850797...	F5-3	Needs repository...
87797964...	F6-3	Needs repository...

Harvesting Panel:

- Harvest commands for Xtal-467... in F9-3
- Force Film Focusing
- Cryo Protection
- Harvest
- Buttons: Start, Stop, Pause, Reposition

Crystal Direct Plate Panel:

- Row: A, Column: 1, Shelf: 1
- Current: E7-2, Inverted:
- Buttons: Move to position
- Table grid (A-H, 1-12)

Pin Storage Panel:

- Pin: CutShape-asym.lib.svg
- Cryo Protection: gate1.lms
- Depressurization: cross1.lms
- Buttons: Pin Barcode, Puck Barcode, New Puck

Pin Robot Positions Panel:

- Buttons: Park, Pin Mount, Cryo Drift, Fishing Drift
- Aspiration: Pin Venturi (Off), Pin Venturi Soft (Off)

Ready Admin 13:52:51

CrystalDirect machine family

CD1 Prototype – 2011

- Automated harvesting



CD2 – 2016

- Pin can be oriented
- Coupled to a sample storing robot
- Commercial version available

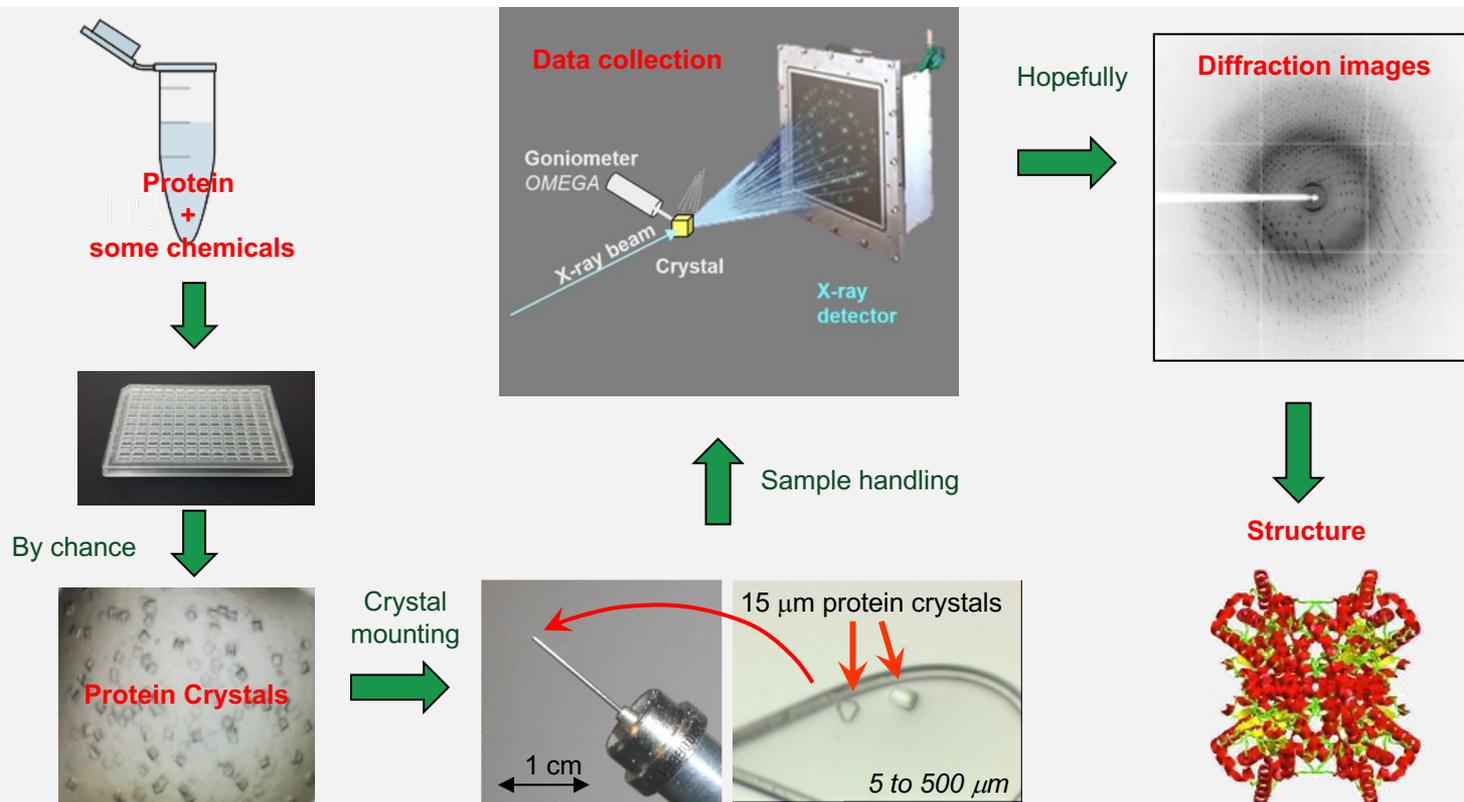


CD3 – 2021

- Temperature regulation (4 – 20 degrees)
- Fast harvesting (< 1 min/crystal)

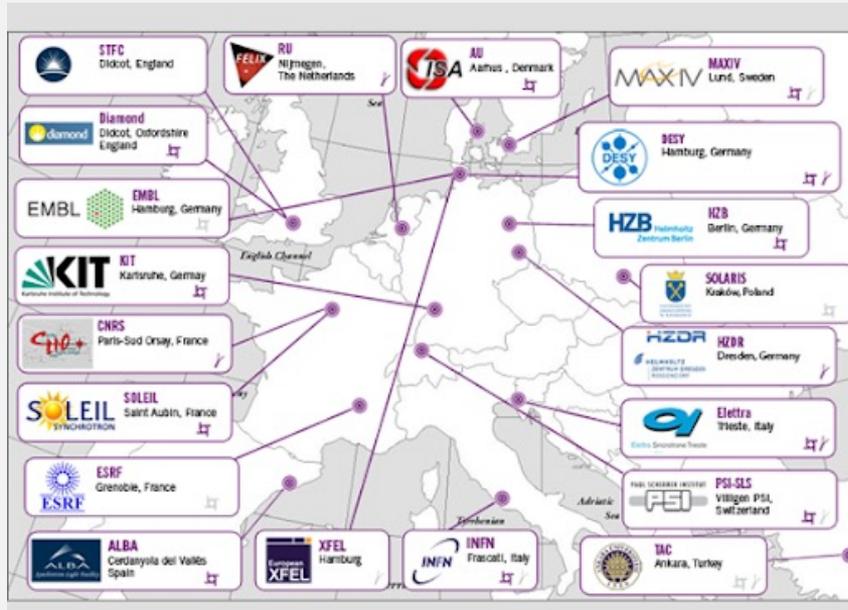


MX in brief



Sample holders

- Tens of synchrotron radiation facilities across europe
 - Samples can be collected at different facilities



Need for

STANDARD (S)

Sample holder standards

SPINE puck and pin standard – 2004

- Up to 10 samples in standard pucks
- 16 samples in unipucks



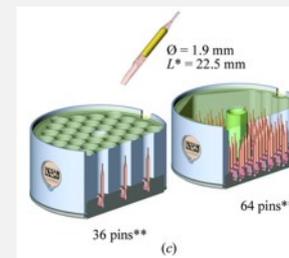
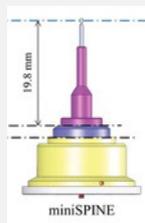
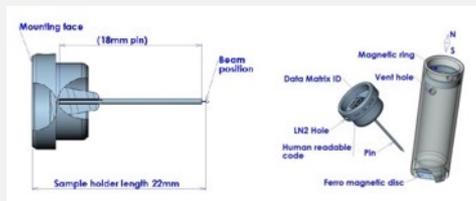
miniSPINE sample holder and puck – 2016

- Up to 36 samples



NewPin sample holder and puck – 2016

- High density & high repositioning precision
- Up to 64 samples in a puck

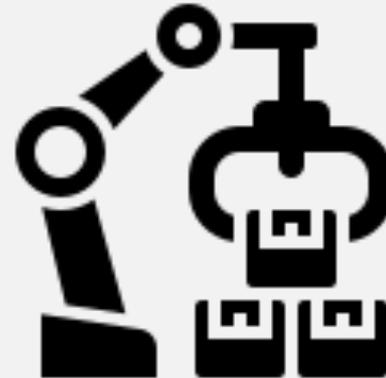


Sample handling on beamlines

- Synchrotrons compatibility
- How to handle samples on beamlines



Automation needed



Sample changer robots

SC3 sample changer – 2006

- EMBL / ESRF
- Capacity: 50 samples



FlexED8 – 2013

- EMBL
- Capacity: 128 samples
- Up to 288 with miniSPINEs



FlexHCD – 2016

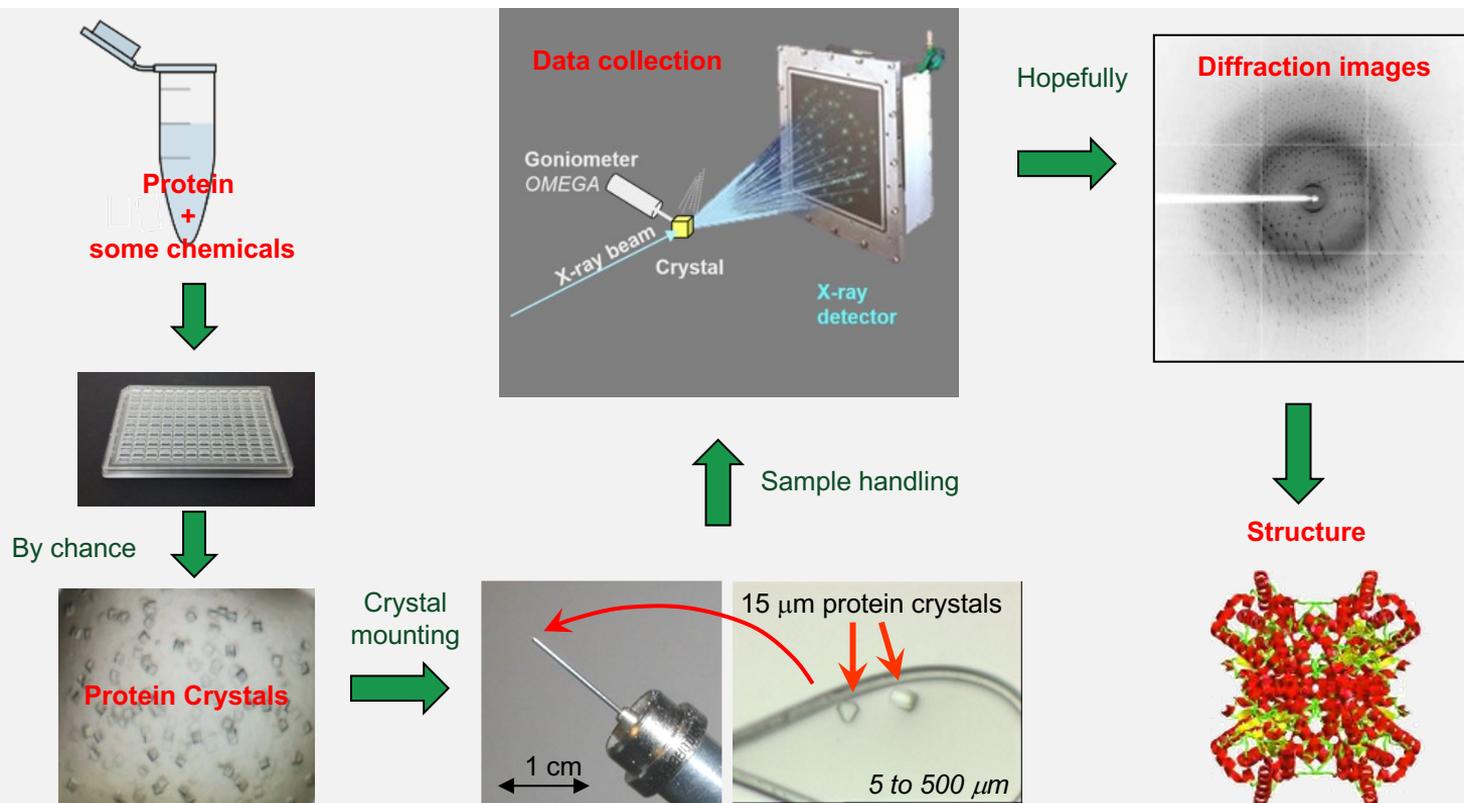
- EMBL / ESRF
- Capacity: 312 samples
- Up to 864 with miniSPINEs



Sample loading video

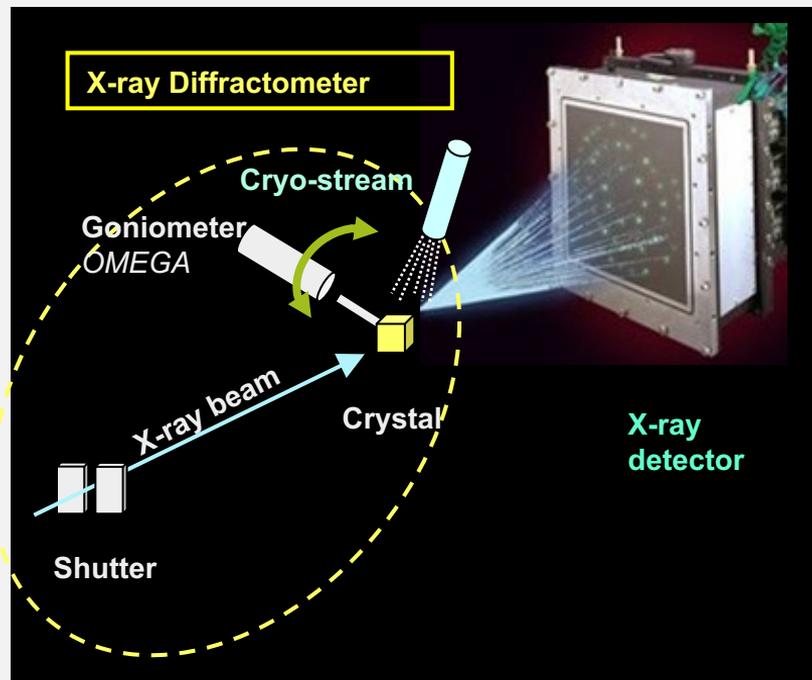


MX in brief

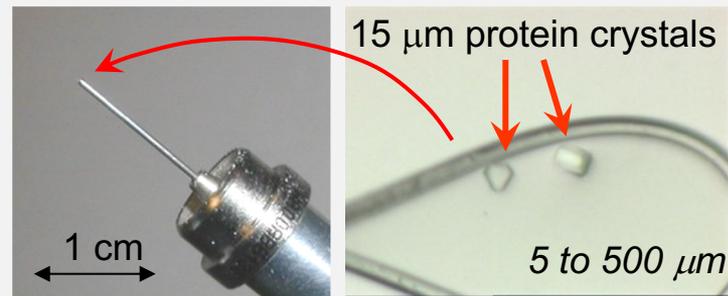


X-ray data collection at synchrotron

Typical experimental setup



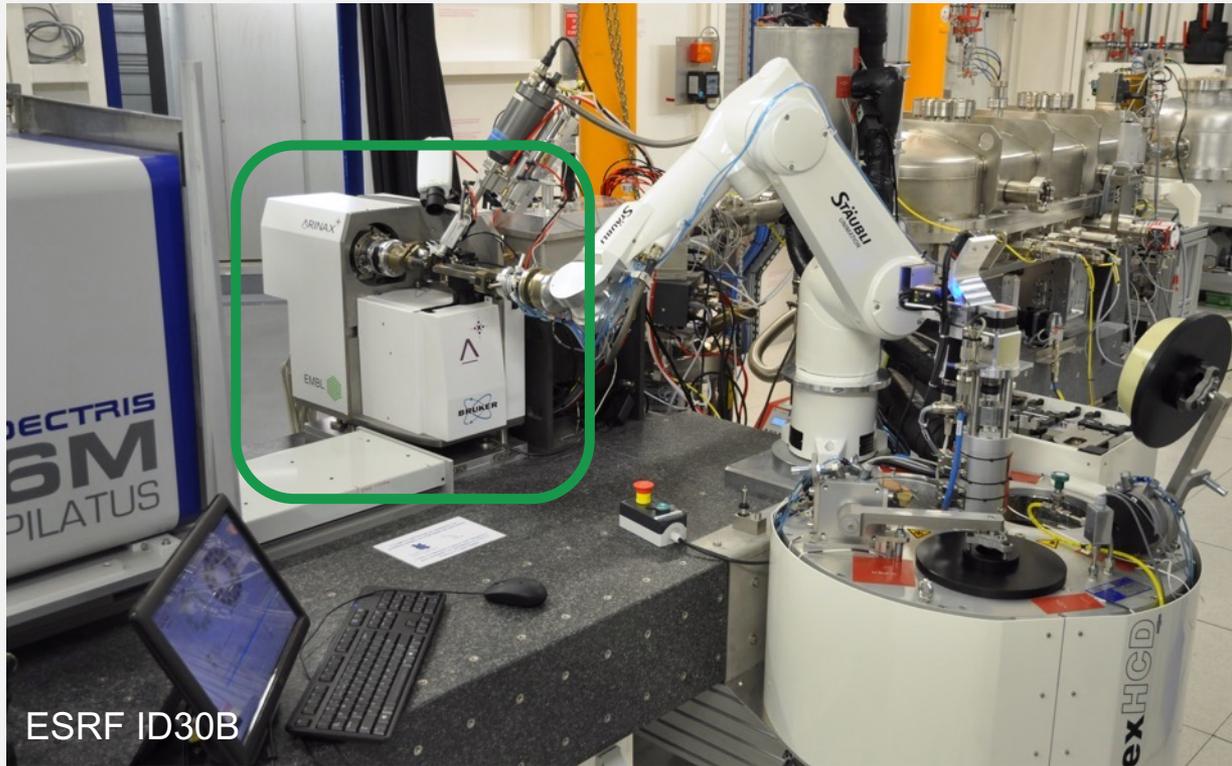
Crystals mounted in a “cryo-loop”



Collecting a data set

recording hundreds of diffraction images during **angular Scans**
In minutes, seconds for the most intense beamlines

MX beamline: Diffractometers



MD goniometer family

MD1 Prototype - 1998
EMBL GR / ESRF
Process 10 μm crystals



MD2 industrial version - 2002
EMBL GR



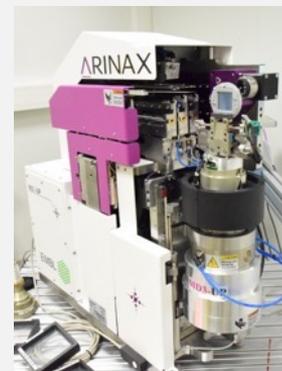
MD3 - 2012
EMBL GR / HH / Arinax
Process routinely 1 μm
crystals



MD3up - 2016
EMBL GR / Arinax
Easy sample changer
access

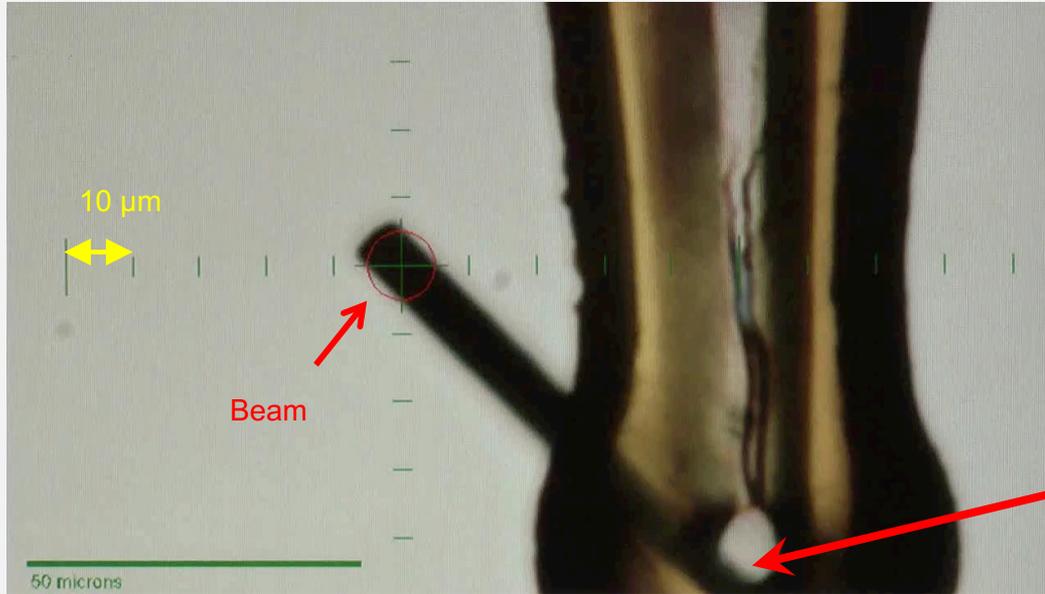


MD3ssx - 2020
EMBL GR / ESRF / Arinax
Serial data-collection on
SSX chips



MD3 – High dynamic precision

- Video



4D Scan along a 8 μm Ø needle - 180 deg., 10 sec



Synchronous move

- OMEGA (rotation)
- Centring table (XY)
- Alignment table (YZ)

MD goniometer family

MD1 Prototype - 1998
EMBL GR / ESRF
Process 10 μm crystals



MD2 industrial version - 2002
EMBL GR



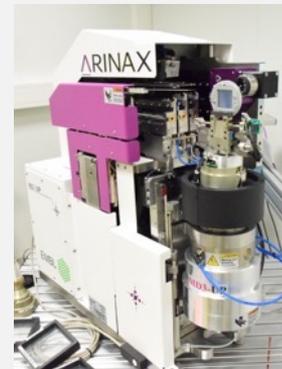
MD3 - 2012
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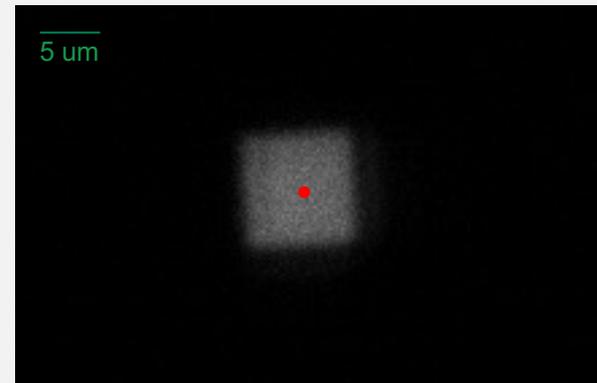
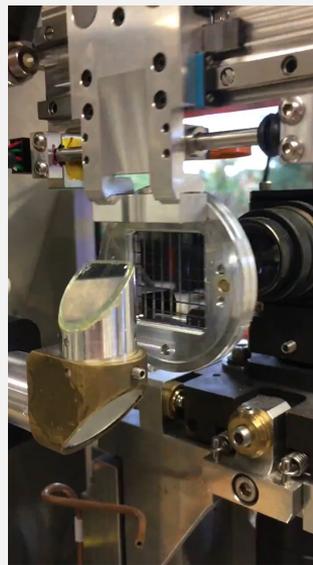
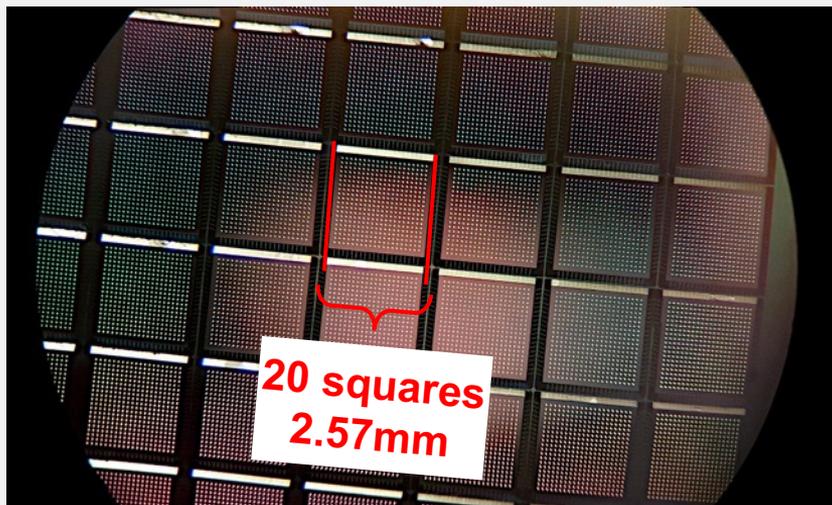
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Process routinely 1 μm
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MD3ssx - 2020
EMBL GR / ESRF / Arinax
Serial data-collection
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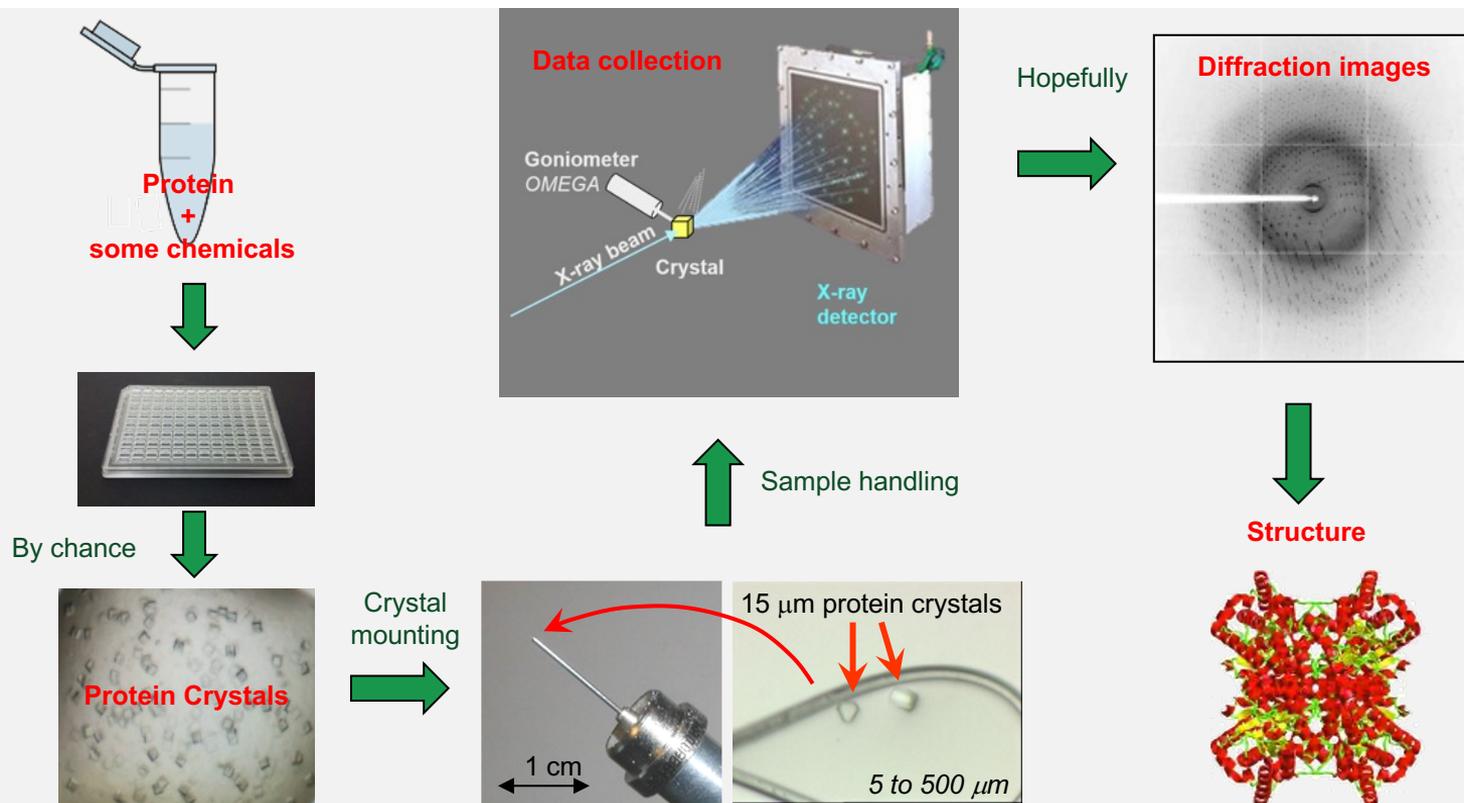


MD3 SSX – Chopper synchronized data collection on silicon chips



Captured at **20 mm/s**
Synchronized on **1kHz beam chopper**
Dynamic **positioning precision** $< \pm 1\mu\text{m}$

MX in brief

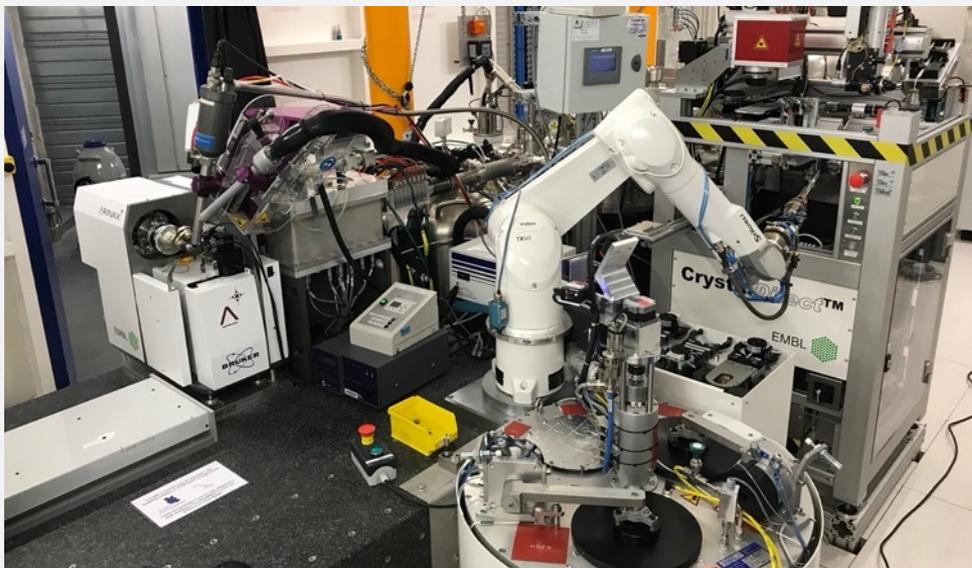


CrystalDirect to Beam

Fully automated Harvest & Collect pipeline

CD1 Prototype - 2017

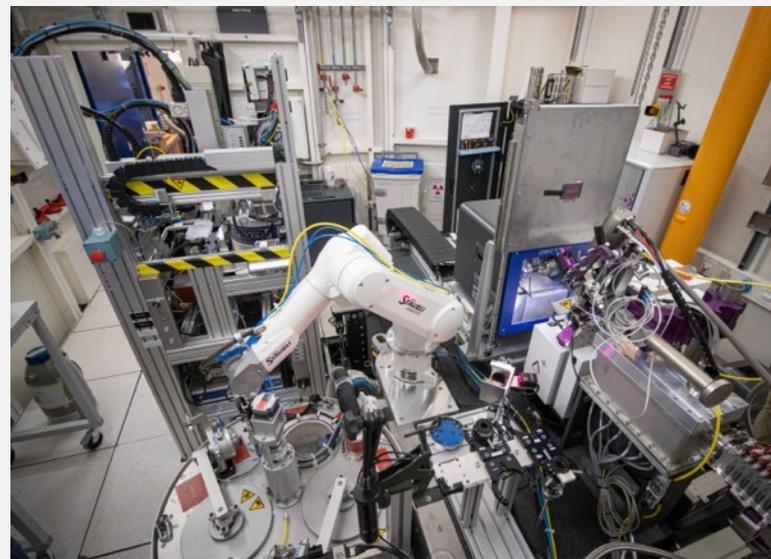
Proof of principle at ID30B - 2017

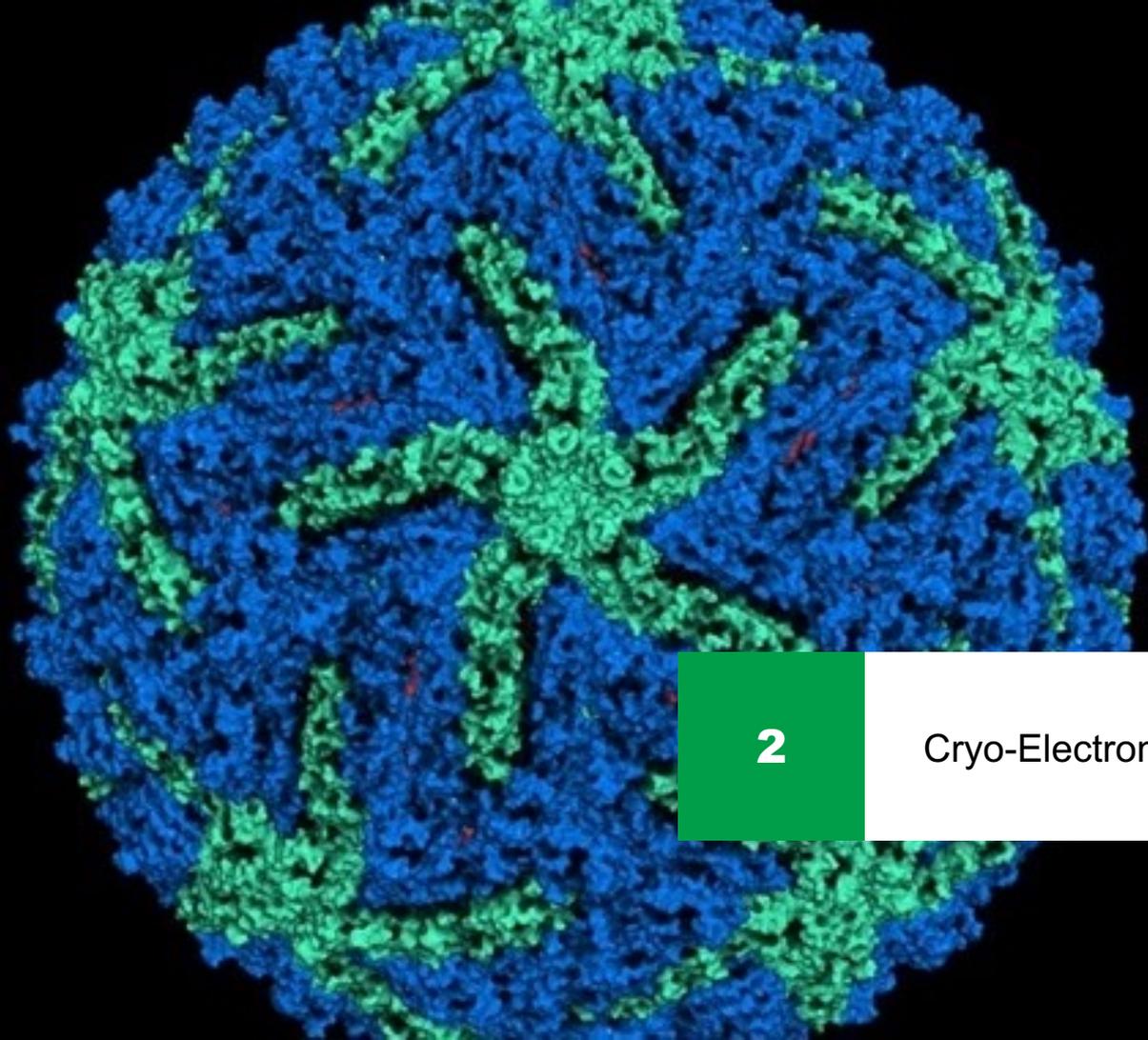


CD2 – 2021 (ongoing)

EMBL / ESRF

Permanent installation on Massif1

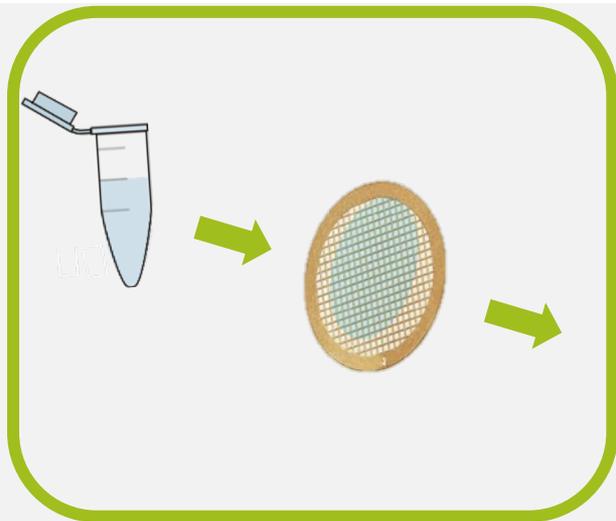




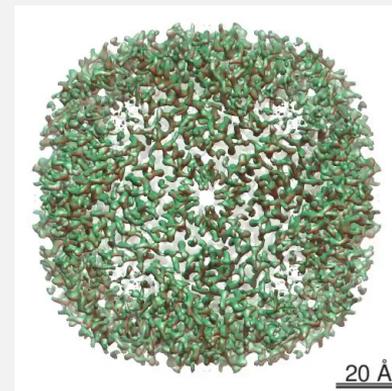
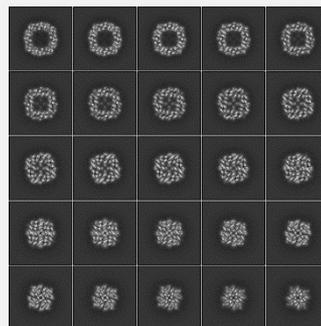
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Cryo-Electron Microscopy

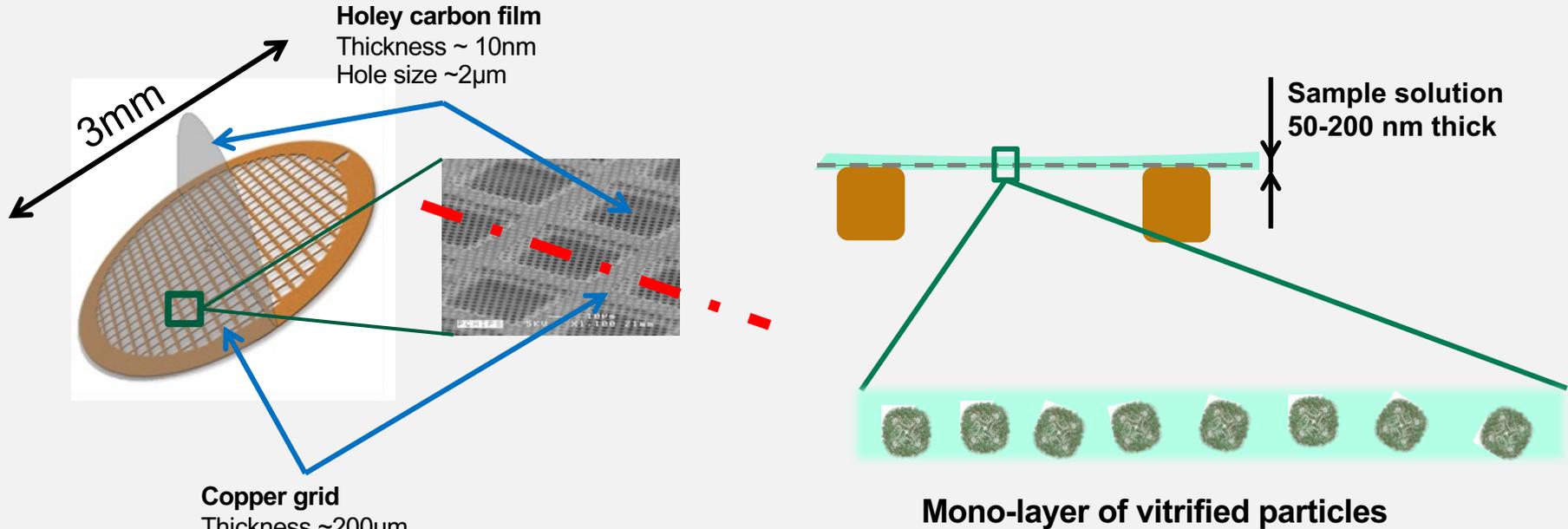
Cryo-EM in short



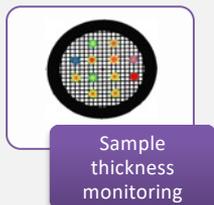
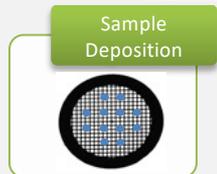
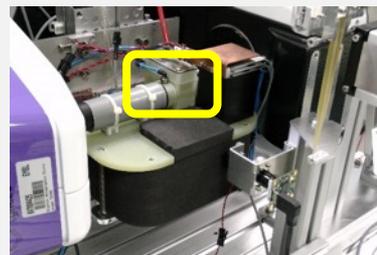
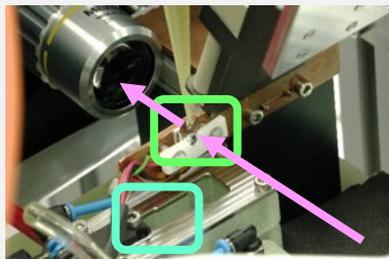
Sample preparation



Cryo-EM : **sample grid** and **optimal sample layout**



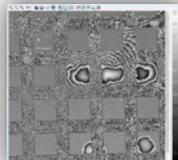
EasyGrid: automated Cryo-EM sample preparation



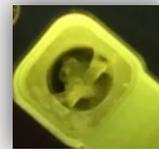
Plasma treatment



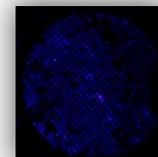
Picoliter drop dispenser



Interferometric method

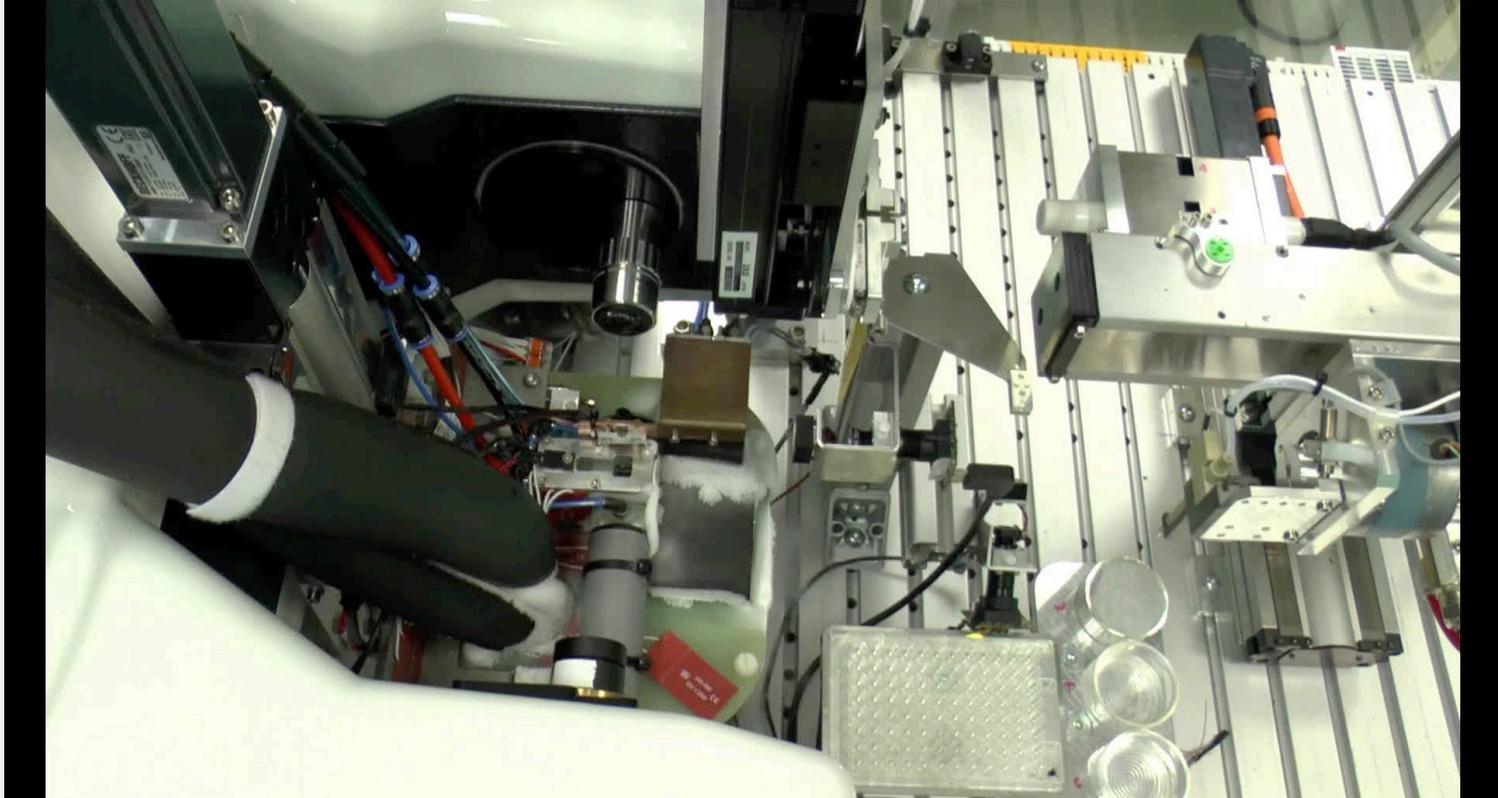


Liquid Propane-Ethane jet



Interferometric method

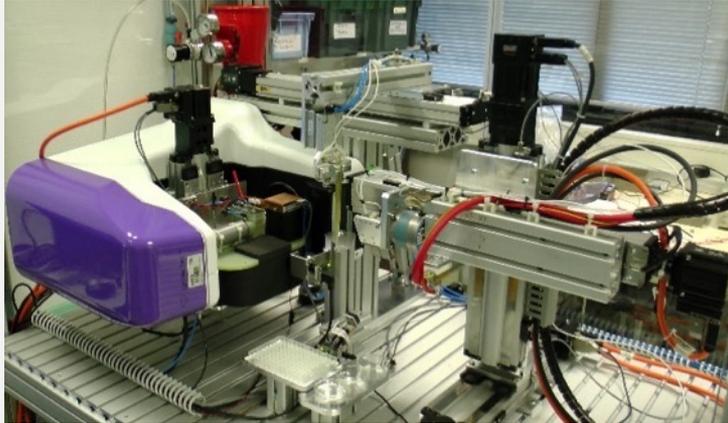
EasyGrid video



EasyGrid machine family

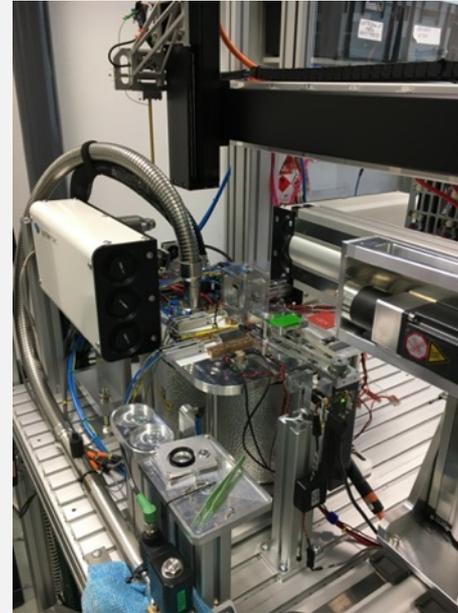
EG1 Prototype – 2018

- *Preparation of 1 grid*
- *Manual grid transfer to boxes*



EG2 – 2021 (ongoing)

- *Automated preparation and storing up to 40 grids*





3

Cryogenic Sample tracking

More and more samples at platforms

- Traditional tracking methods:
 - Datamatrix codes
 - Frost makes reading difficult
 - Damaged barcodes due to handling
 - Hand written codes
 - Difficult to read
 - Possible human errors
 - But
 - Samples represent months of work
 - Loss is hardly tolerated

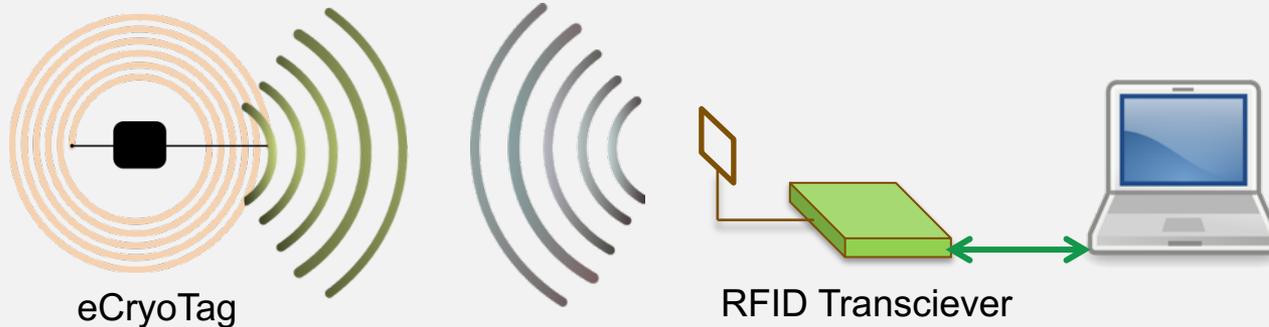
Need for reliable



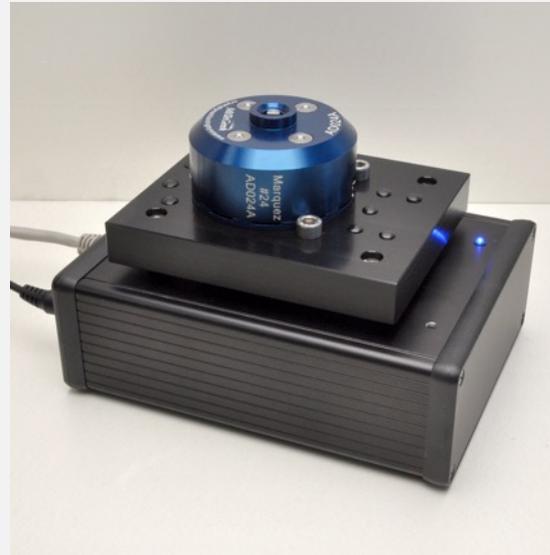
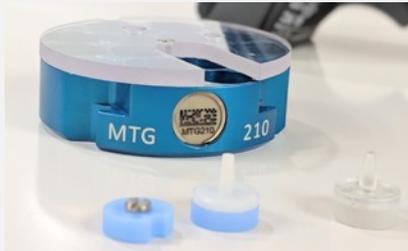
How to effectively track samples at cryogenic temperature?

- **eCryoTag**

- RFID (Radio Frequency Identification) technology
- Contactless
- Fast & reliable reading



A new standard for MX and Cryo-EM sample tracking



eCryoTag usecase at HTXlab crystal harvesting platform



ARISE Fellowship opportunities



<https://www.embl.org/training/technology-developers-programme/arise/>

EMBL Explore Search

Training and development Related: [EMBL Heidelberg](#)

ARISE [Programme overview](#) [Applying](#) [Research focus](#) [Contacts](#)

EMBL Technology Developers Programme

ARISE – Career Accelerator for Research Infrastructure Scientists

The ARISE fellowship programme offers fellowships for **experienced professionals** with a background in science, technology, engineering, or mathematics (STEM) who wish to advance **technology development** in the life sciences and receive training to operate **life science research infrastructures**, which provide novel technologies as a service for life scientists.

The ARISE programme

- ARISE is the first fellowship programme of its kind that is training technology developers to become highly educated research infrastructure scientists. After successfully finishing their training, we expect fellows to take positions as senior scientists or leaders in core facilities, research infrastructures, or technology development groups in academia, industry, healthcare, or other sectors.

Technology development, especially with the purpose of enriching service provision in the life sciences, is currently one of EMBL's main strategic focuses. EMBL groups are developing technologies in **imaging, chemical biology, computational modelling, microfluidics, robotics, X-ray optics, high-precision mechanics, data acquisition, automation, bioinformatics and software development, and integrated structural biology.**

The ARISE programme is co-funded by EMBL and the [Marie-Sklodowska-Curie Action \(MSCA\)](#) of the European Commission's Horizon 2020 programme – co-funding of regional, national and international programmes (COFUND).

[Get a full overview of the ARISE Programme](#)

Jump to

- Training of ARISE Fellows
- Career guidance
- Employment aspects
- Open Science
- Ethical issues

**2nd Call:
1st September – 31 October 2021**



Acknowledgements

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