

Ab initio modelling: recovering 3D shapes from 1D data

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How can one obtain a 3d model from the 1D SAXS data?

- SAS curve contain information about distances.
- How to use this information to build model based on the SAXS data.

Outline

- Computing form factor from geometrical shape
- Bead modelling:
 - Principle
 - Target function and minimization
 - Bead modeling
 - Dummy residue modeling
- Words of caution

Form factor of simple geometrical shape

Computing form factor from simple geometrical shape

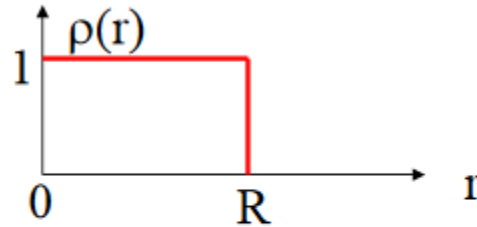
$$I(\mathbf{s}) = A(\mathbf{s}) \cdot A^*(\mathbf{s})$$

$$A(\mathbf{s}) = \int_{V_r} \rho(\mathbf{r}) \cdot e^{-i\mathbf{r} \cdot \mathbf{s}} dV_r$$

For simple geometrical shapes, the form factor can be computed from the electron density

Example: form factor of a solid sphere

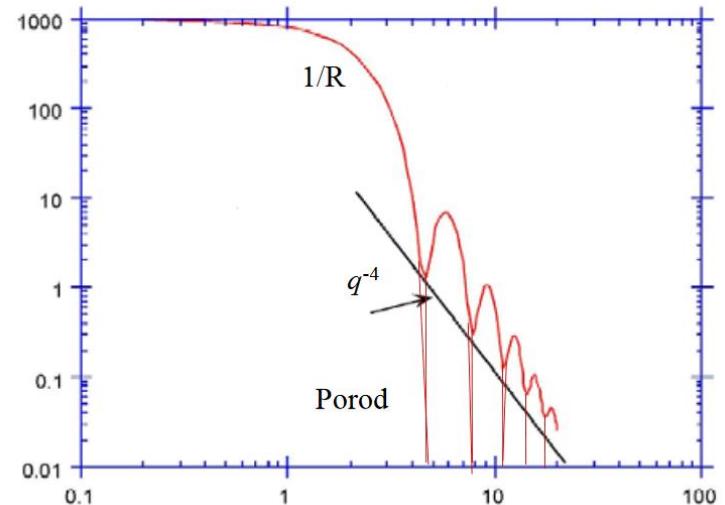
Form factor of a solid sphere



$$A(q) = 4\pi \int_0^{\infty} \rho(r) \frac{\sin(qr)}{qr} r^2 dr = 4\pi \int_0^R \frac{\sin(qr)}{qr} r^2 dr$$
$$= \frac{4\pi}{q} \int_0^R \sin(qr) r dr =$$

Form factor of sphere

$$P(q) = A(q)^2/V^2$$



The sphere case is trivial,
It quickly become complicated

Table 3.4. Equations for Scattering Intensities of Simple Bodies

| | |
|---|--|
| Uniform sphere of radius R | $9 \left(\frac{\sin t - t \cos t}{t^3} \right)^2 = \phi^2(t), \quad t = sR$ |
| Spherical layer with radii $R_1 > R_2$ | $(R_1^3 - R_2^3)^{-2} [R_1^3 \phi(sR_1) - R_2^3 \phi(sR_2)]^2$ |
| Triaxial ellipsoid (semiaxes a, b, c) | $\int_0^1 \int_0^1 \phi^2 \{ s [a^2 \cos^2(\frac{1}{2}\pi x) + b^2 \sin^2(\frac{1}{2}\pi x)(1 - y^2) + c^2 y^2]^{1/2} \} dx dy$ |
| Ellipsoid of rotation $a: a: va$ | $\int_0^1 \phi^2 [sa(1 + x^2(v^2 - 1))^{1/2}] dx$ |
| Parallelepiped (edges A, B, C) | $\int_0^1 \Psi_p[s, B(1 - x^2)^{1/2}, A] S^2(sBCx/2) dx; S(t) = \sin(t)/t$ $\Psi_p(s, B, A) = \frac{2}{\pi} \int_0^{\pi/2} S^2[sA \sin(y/2)] S^2[sB \cos(y/2)] dy$ |
| Right elliptical cylinder with height H , semiaxes of ellipse a, va | $\int_0^1 \Psi_{ec}[s, a(1 - x^2)^{1/2}] S^2(sHx/2) dx$ $\Psi_{ec}(s, a) = \frac{1}{\pi} \int_0^\pi A_1^2 \left[sa \left(\frac{1 + v^2}{2} + \frac{1 - v^2}{2} \cos y \right)^{1/2} \right] dy$ $A_1(t) = 2J_1(t)/t$ |
| Right hollow cylinder with height H , outer radius R_1 , inner radius R_2 | $\int_0^1 \Psi_{hc}[s, R_1(1 - x^2)^{1/2}, R_2(1 - x^2)^{1/2}] S^2(sHx/2) dx$ $\Psi_{hc}(s, R_1, R_2) = \frac{1}{1 - \gamma^2} [A_1(sR_1) - \gamma^2 A_1(sR_2)]$ $\gamma = R_2/R_1$ |
| Right circular cylinder of radius R , height H | $4 \int_0^1 \frac{J_1^2[sR(1 - x^2)^{1/2}]}{[sR(1 - x^2)^{1/2}]^2} S^2(sHx/2) dx$ |
| (a) $R = 0$ (infinitely thin rod, height H) | $2 \text{Si}(sH)/sH - S^2(sH/2), \quad \text{Si}(t) = \int_0^t S(x) dx$ |
| (b) $H = 0$ (infinitely thin disk, radius R) | $[2 - A_1(2sR)]/s^2 R^2$ |

DETERMINATION OF THE INTEGRAL PARAMETERS OF PARTICLES

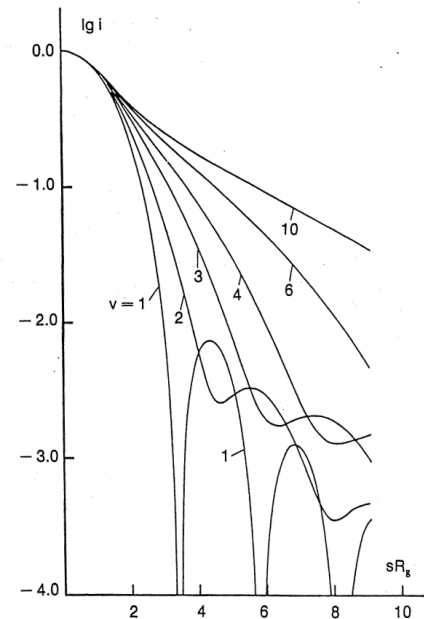
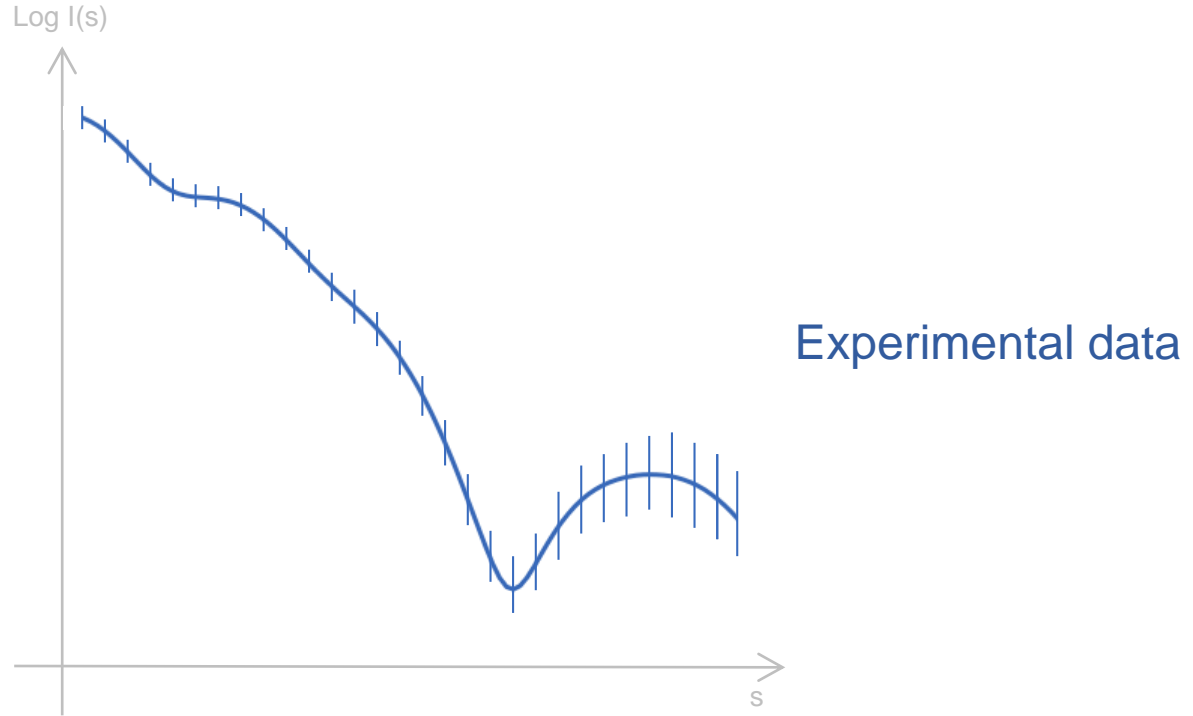


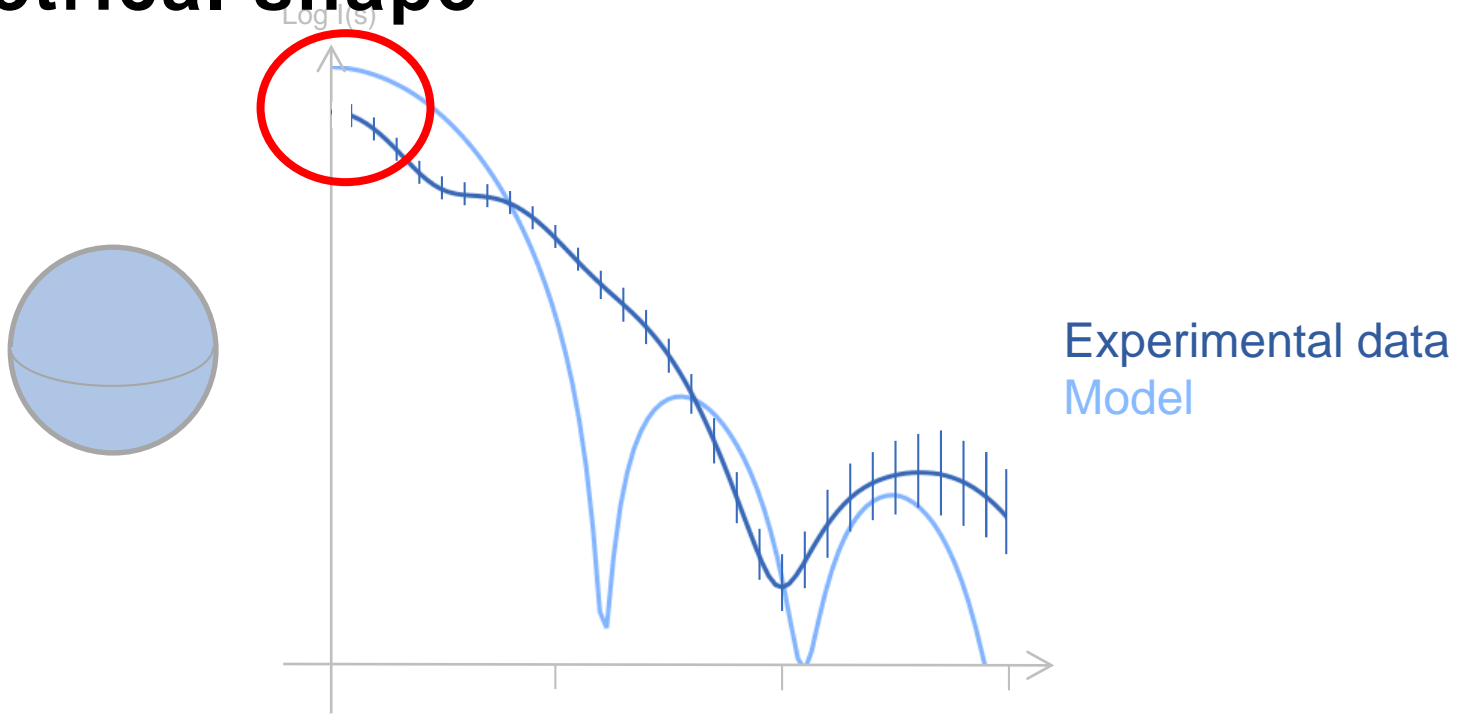
Figure 3.12. Scattering curves for prolate ellipsoids of rotation with $v = c/a$ (after Kratky and Pilz, 1972).

$I(s) = I(s, \mathbf{X})$ [e.g., for an ellipsoid $\mathbf{X} = (a, b, c)$], one can apply the algorithm described in Section 3.2. From the approximate classes of bodies we can choose that providing the best fit with experiment (namely, with the scattering curve and the invariants). It should be noted, however, that the scatterer must be sufficiently large s , even in a region of homogeneity, cannot be represented by the scattering curve from a simple body;

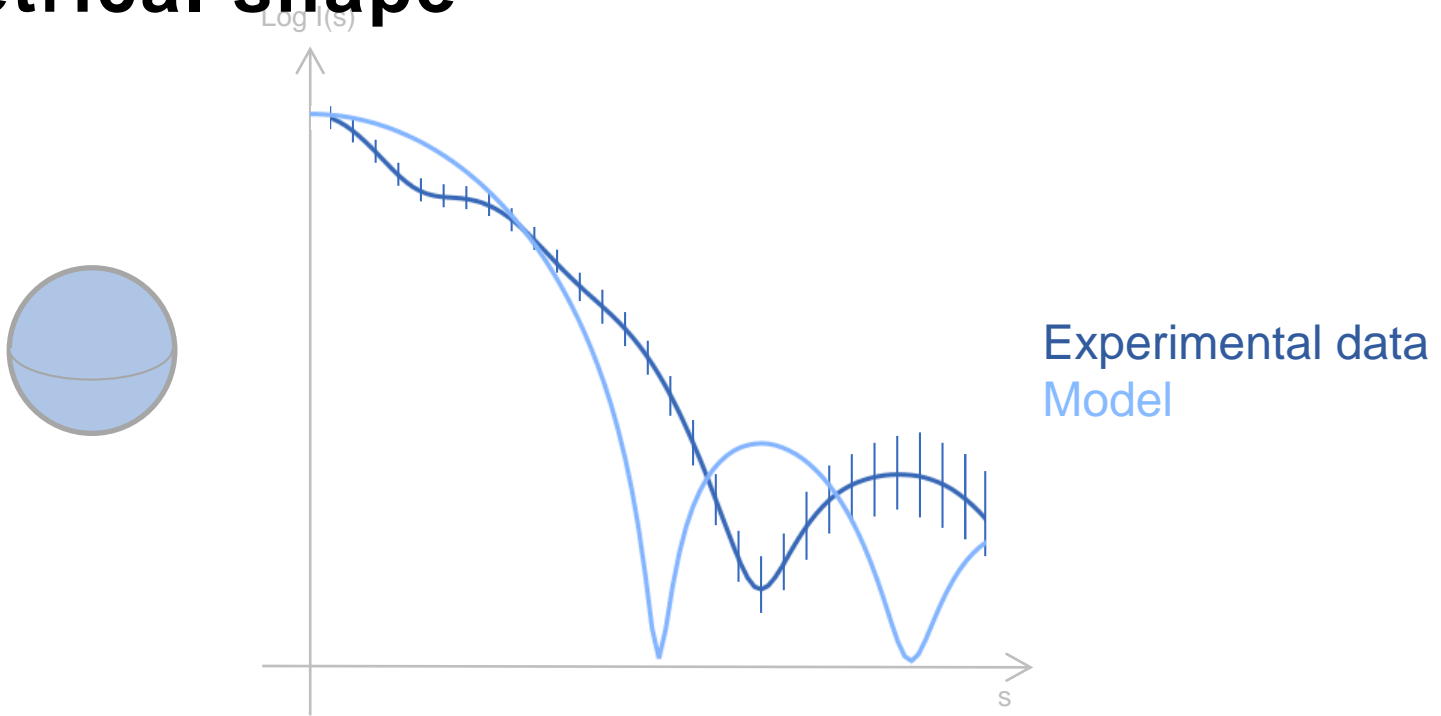
SAXS curve



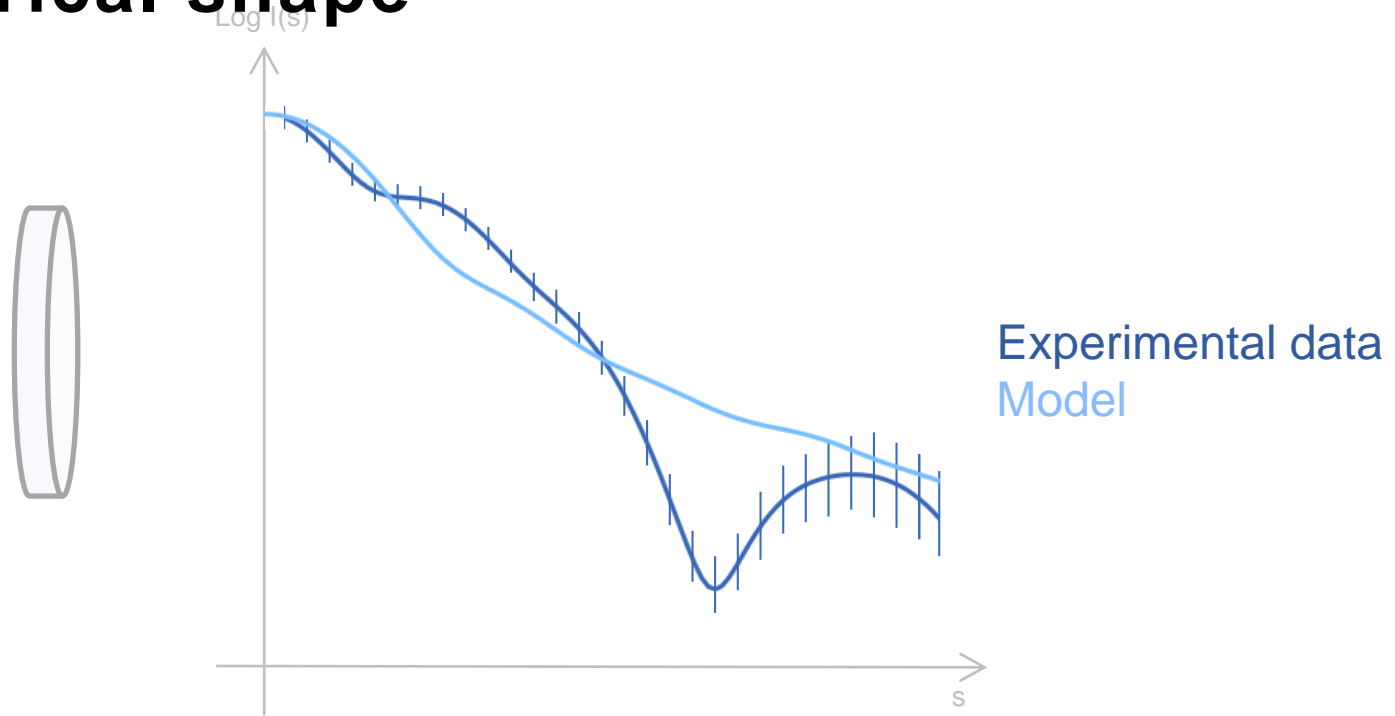
Form factor computed from simple geometrical shape



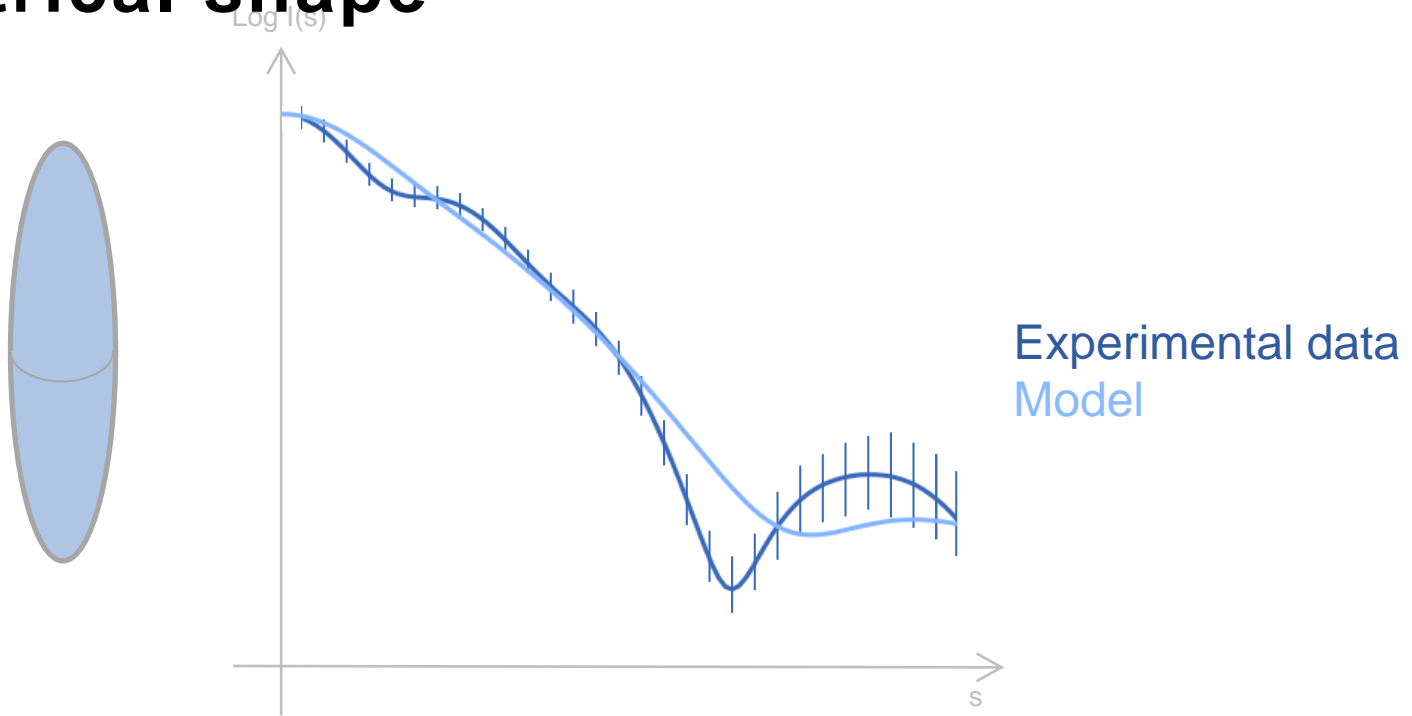
Form factor computed from simple geometrical shape



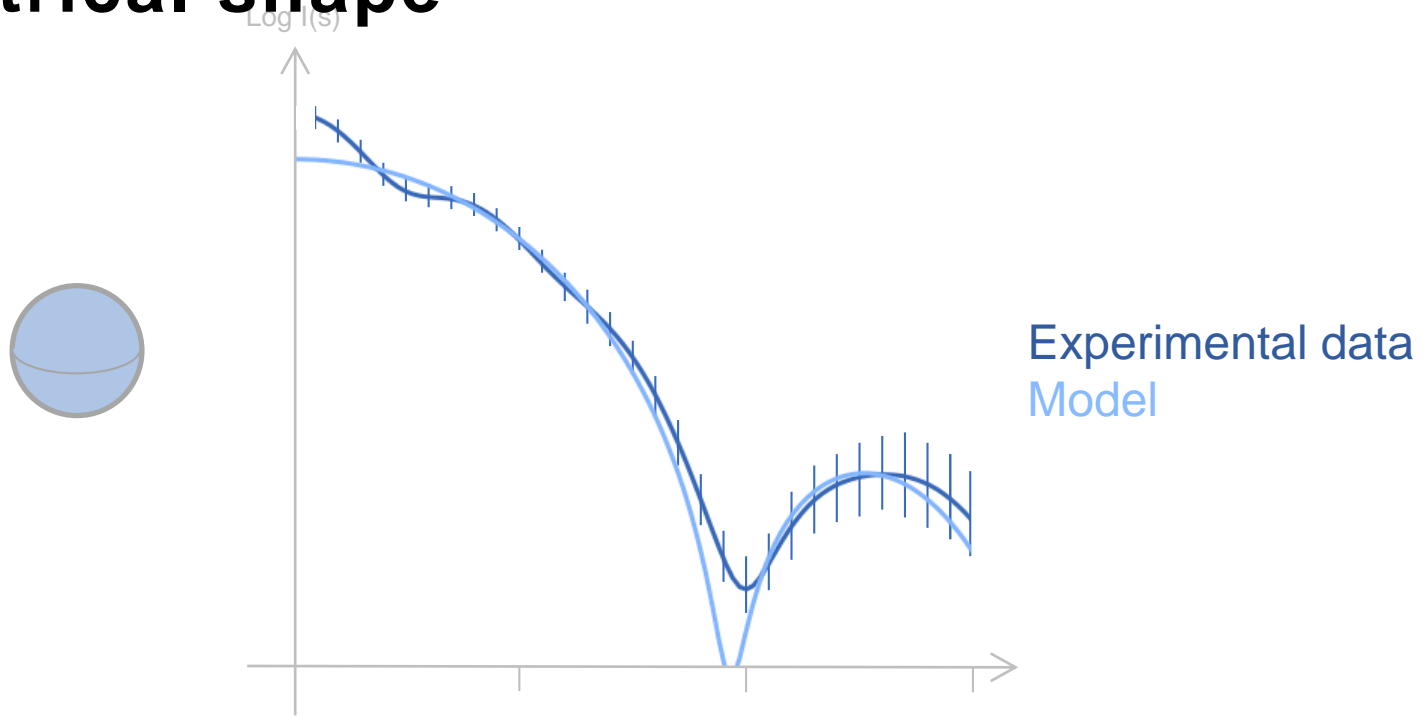
Form factor computed from simple geometrical shape



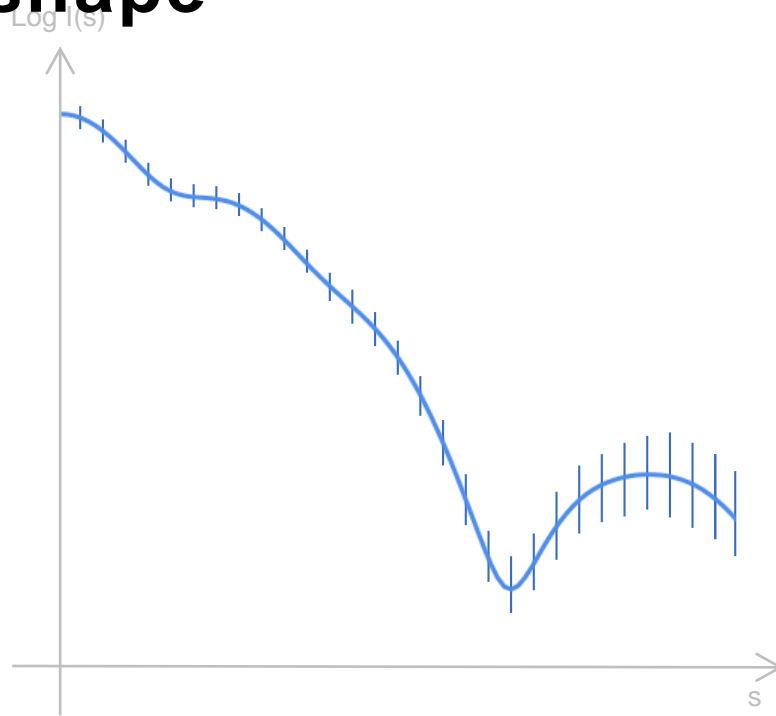
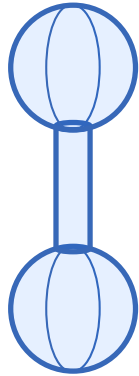
Form factor computed from simple geometrical shape



Form factor computed from simple geometrical shape

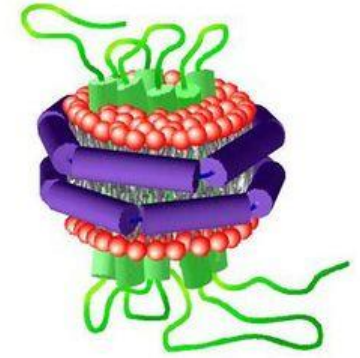
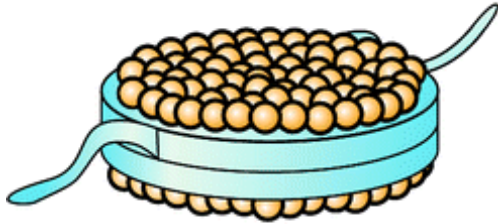


Form factor computed from simple geometrical shape



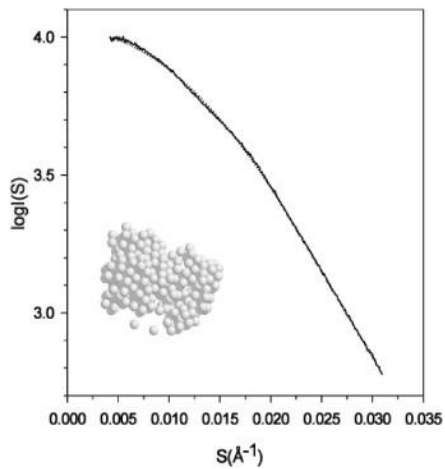
Experimental data
Model

Example: modelling nano-disc



$$A_{tags} + A_{cap} + A_{tails} + A_{meth} + A_{belt} = A_{disc}$$

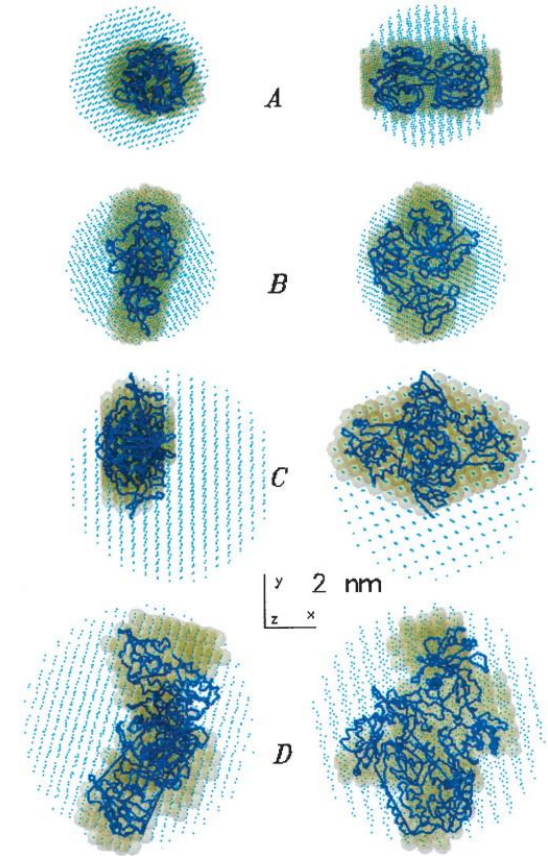
Skar-Gislinge *et al.* *J. Am. Chem. Soc.*

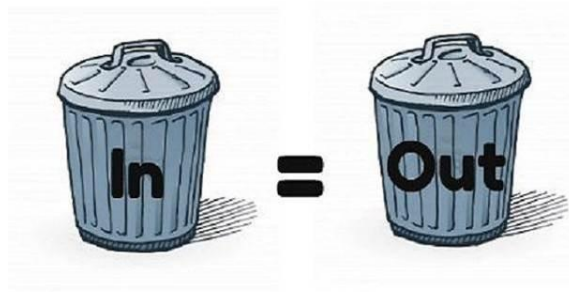


Chacón, P. *et al.* (1998)
Biophys. J. 74, 2760-2775.
 → minimization using genetic algorithm

Bead models

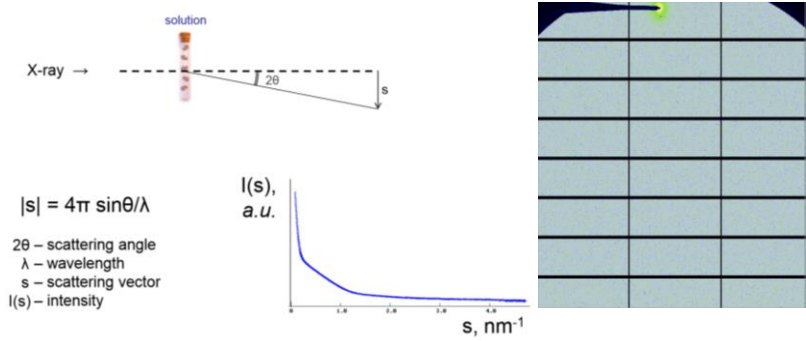
Svergun, D.I. (1999)
Biophys. J. 76, 2879-2886
 → minimisation using simulated annealing



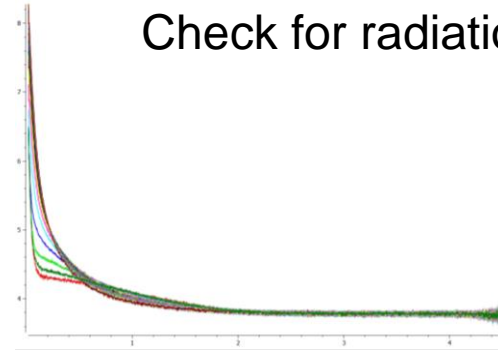


- Ab initio modelling (contrary to many other modelling approach) will always give a nice looking model that fit the data , even if the data are completely wrong.
- Make sure that the SAXS curves used for ab initio correspond to the form factor of the solutes you are trying to measure.

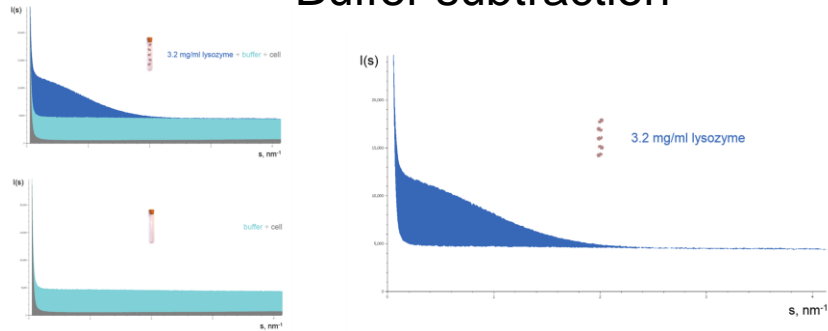
Data reduction



Check for radiation damage

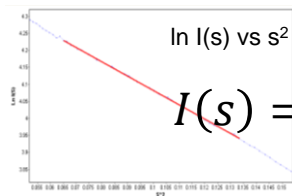


Buffer subtraction



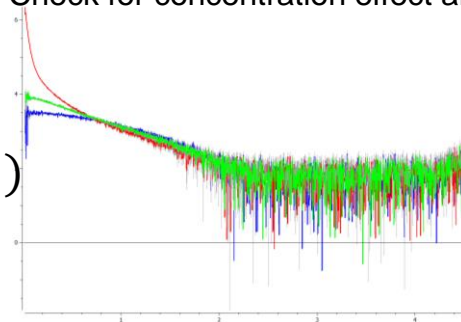
Check overall parameters, before ab initio modelling

Radius of gyration (Guinier)



$$I(s) = I_0 \exp\left(-\frac{1}{3} s^2 R_g^2\right)$$

Check for concentration effect and aggregation



Estimation of molecular weight by forward scattering

$$MW = \frac{I(0)}{c} \cdot \frac{(c_{st} \cdot MW_{st})}{I(0)_{st}}$$

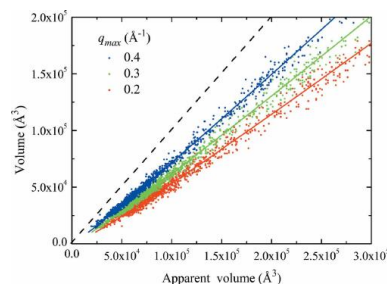
Porod Volume

$$V_{Porod} = 2 \cdot \pi^2 \frac{I(0)}{Q}$$

Volume of correlation

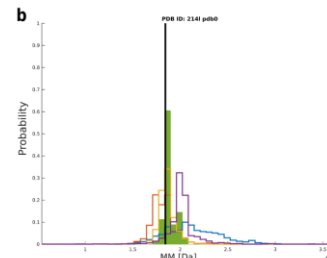
$$V_C = \frac{I(0)}{\int q \cdot I(q) dq}$$

SAXS mow



Fischer, H. et al. J. Appl. Cryst. 2010

Bayesian MW

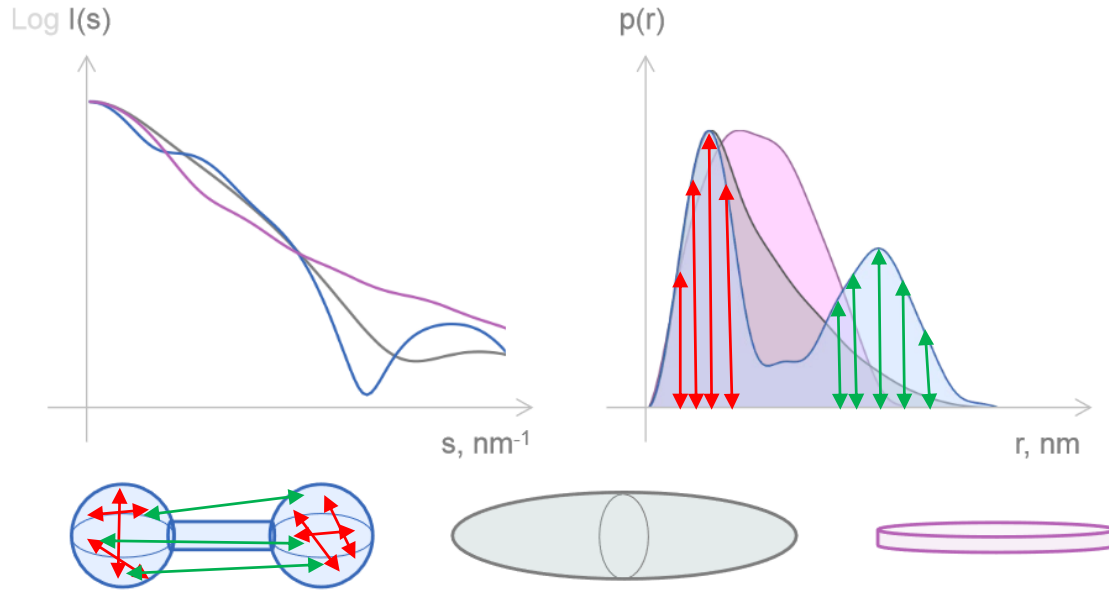


Rambo RP, Tainer JA. *Nature*. 2013

Hajizadeh NR, et al. *Sci. Rep.* 8:7288 (2018)



Distance distribution function

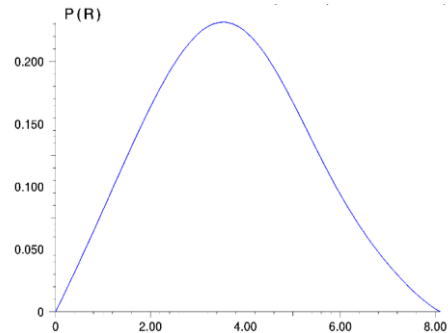
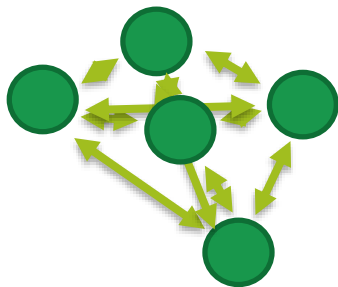


$$p(r) = \rho^2 \gamma_0(r) V r^2$$

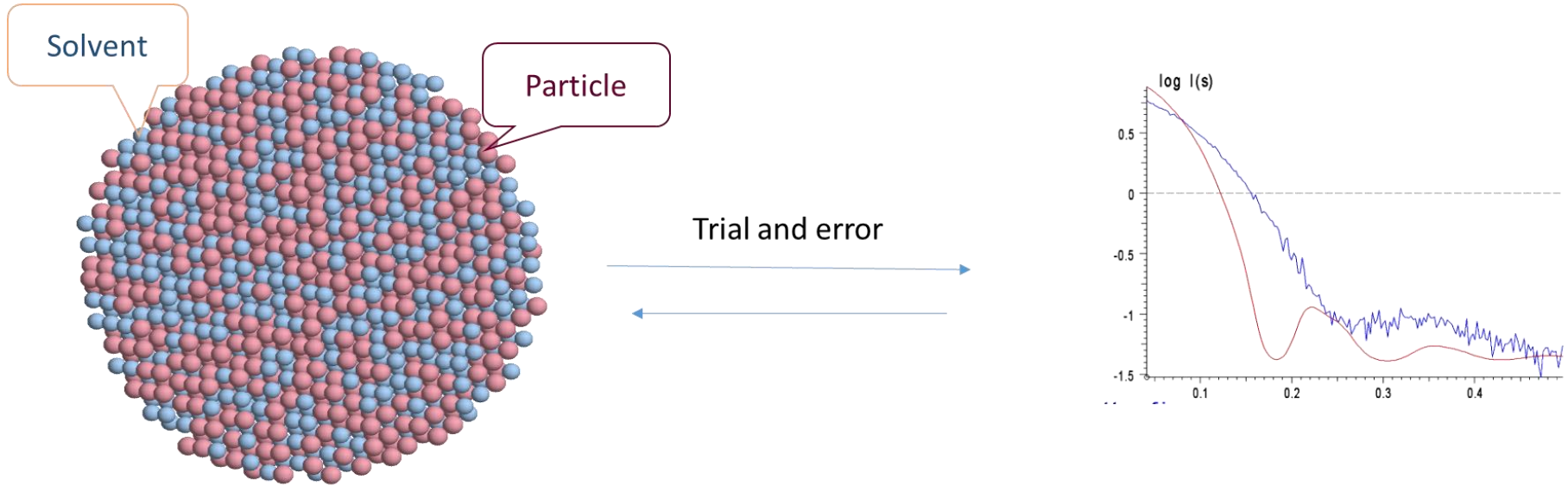
Where $\gamma_0(r)$ is the probability of finding a point within the particle at a distance r from a given point.

Ab initio bead modelling: Basic idea

- Find an ensemble of beads with the inter-bead distances are consistent with the $p(r)$



Beads on a grid

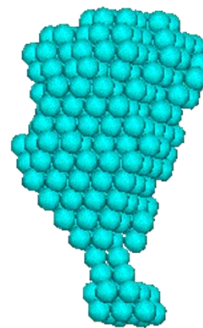
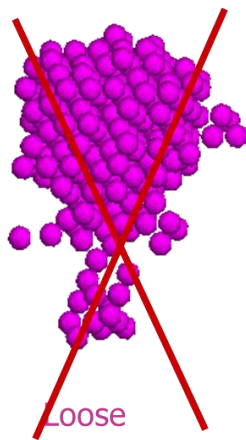
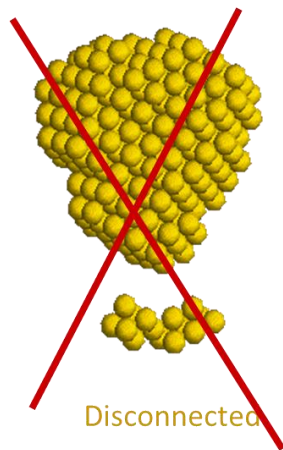


Computation of the theoretical SAXS curve from the bead ensemble and fit to the experimental SAXS data.

$$\chi^2 = \frac{1}{N} \sum_{j=1}^N \left(\frac{I_j - \langle I \rangle}{\sigma_j} \right)^2$$

Penalty terms

- Bead configuration should not only fit the data but also provide a compact model. This can be reinforced by the use of penalty terms.
- The looseness penalty term is computed from the bead configuration and is small when the bead ensemble has a compact configuration

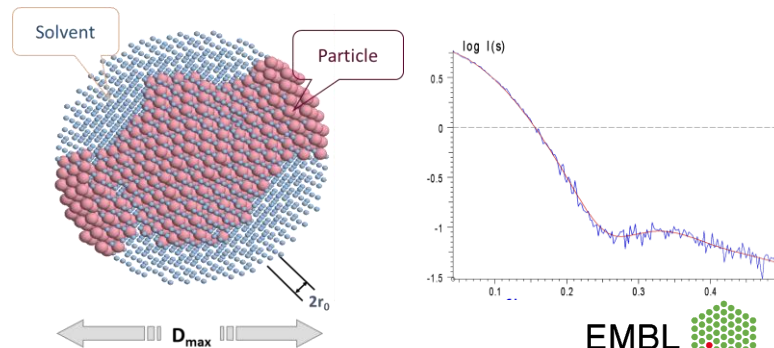
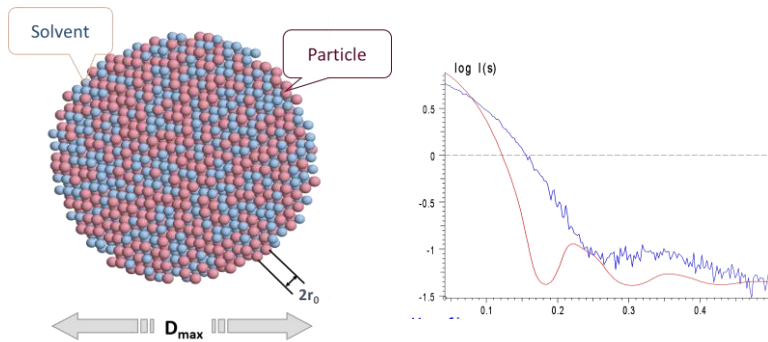


Finding good bead ensemble

Find the bead ensemble that minimized the target function:

$$f(X) = \chi^2 + \alpha \cdot P(X)$$

$$\chi^2 = \frac{1}{N} \sum_{i=1}^N \left(\frac{I_i - \langle I \rangle}{\sigma_i} \right)^2$$



Minimization of the target function

Parameterization:

a binary vector,
0 if solvent, 1 if particle

Solvent

Particle

$$f(X) = \chi^2 + \alpha \cdot P(X)$$

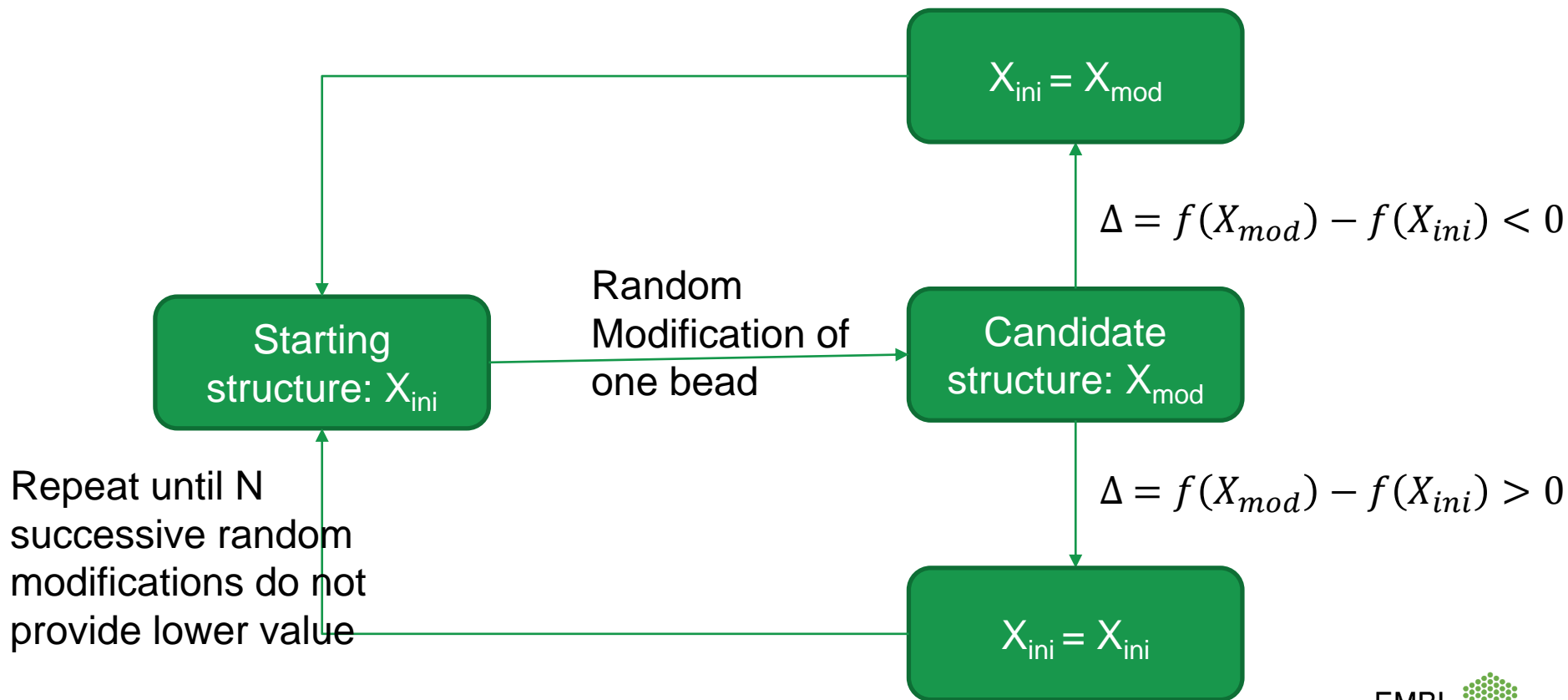
Iterative approach:

- a bead can be changed
- the effect of this change is evaluated: is the target function smaller after this change?
 - If yes, the changed structure is the new starting configuration for the next iteration.
 - If not, the unchanged structure is used.

$2r_0$

D_{\max}

Pure Monte Carlo



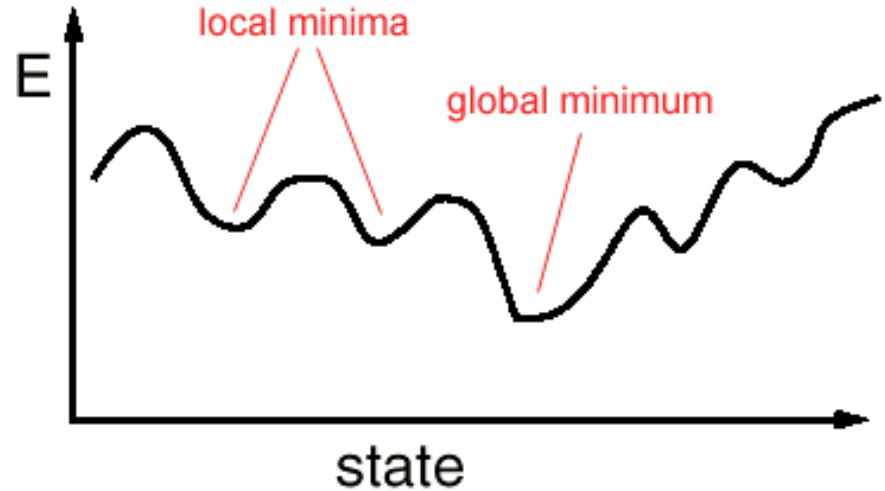
Local Minima vs global minimum

Local search can be trapped in a local minimum.

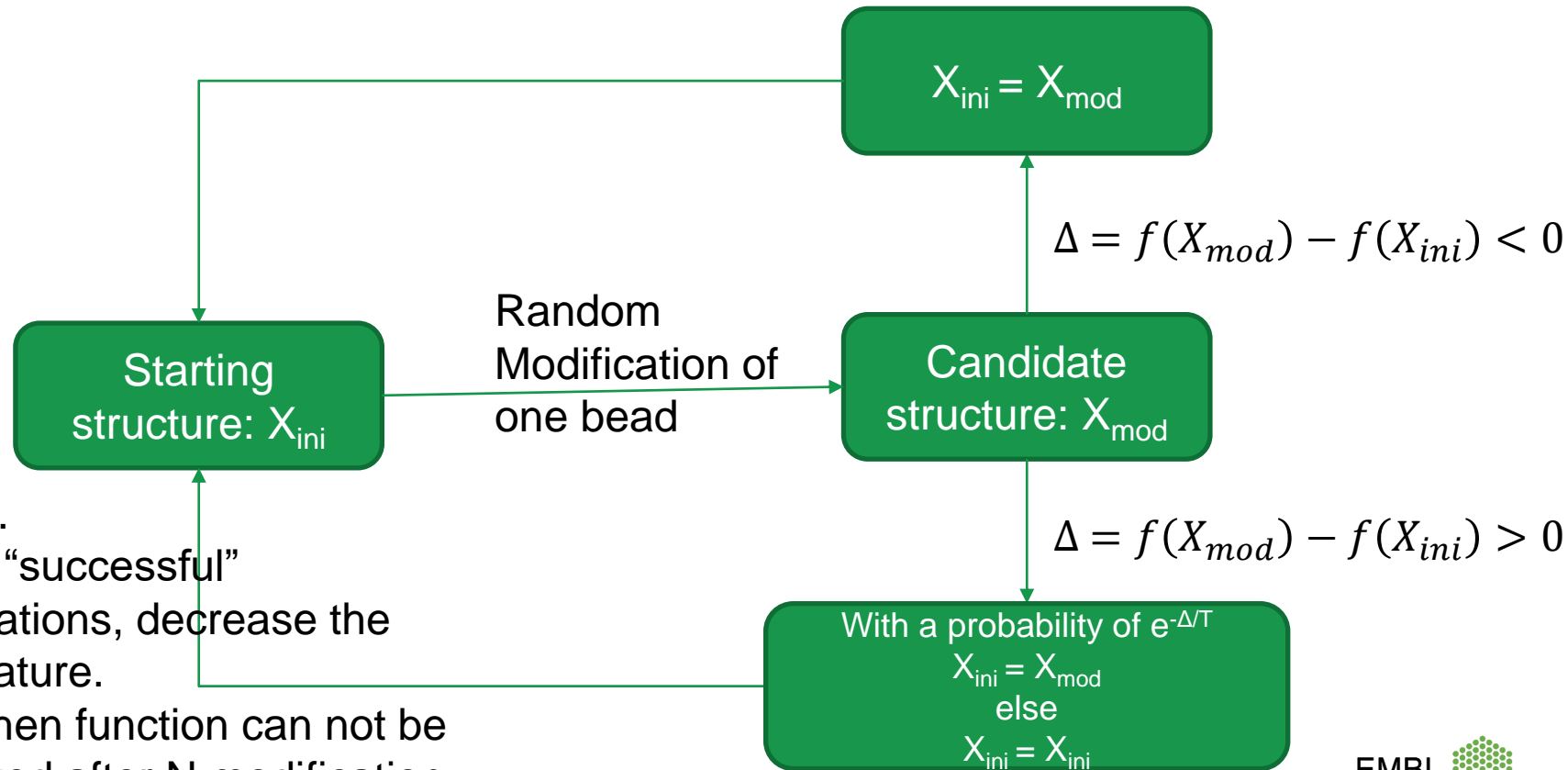
Pure Monte-Carlo search always goes to the closest local minimum (nature: rapid quenching and vitreous ice formation)

To get out of local minima, global search must be able to (sometimes) go to a worse point.

Slower annealing allows to search for a global minimum (nature: normal, e.g. slow freezing of water and ice formation)

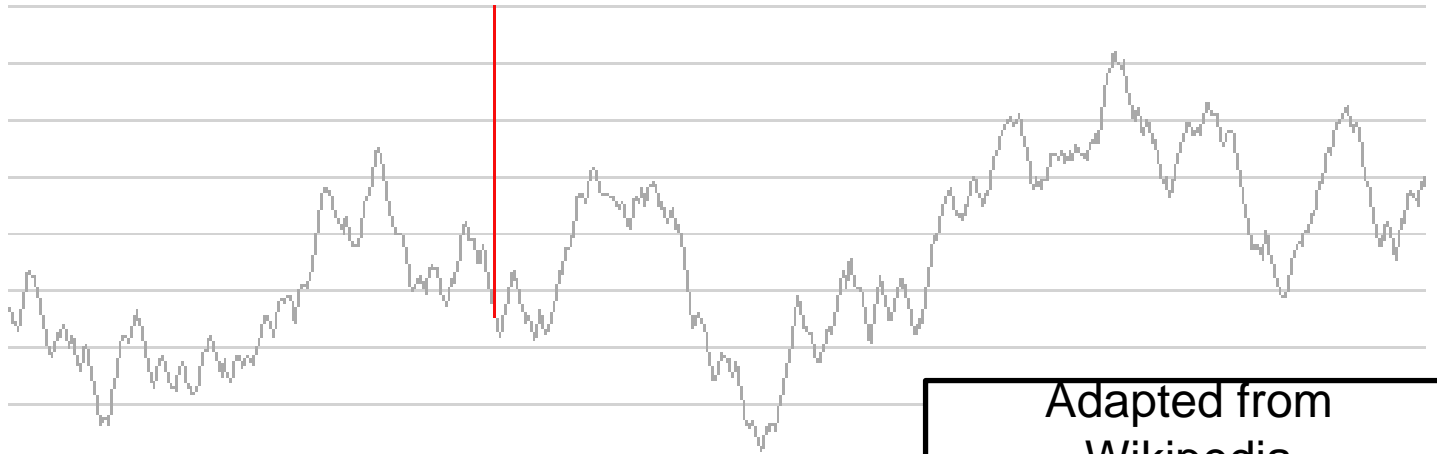


Simulated annealing



Repeat.
after M “successful”
modifications, decrease the
temperature.
Stop when function can not be
minimized after N modification.

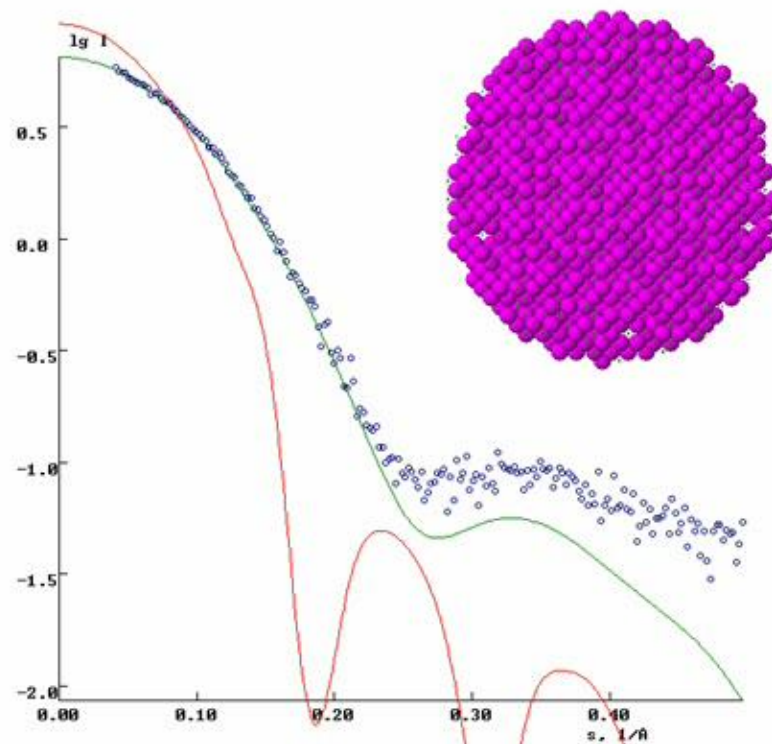
Simulated annealing



Adapted from
Wikipedia

Ab initio program

T = 0.100E-02 Rf = 0.50731 Los: 0.0966 DisCog: 0.0024 Scale = 0.910E-08
Ab initio shape reconstruction of lysozyme



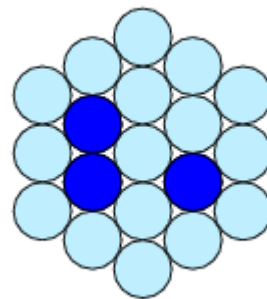
Gnom file : gnomyz.out

Log file : D:\DUnain\Main-05\Dammin\lyzdan.log

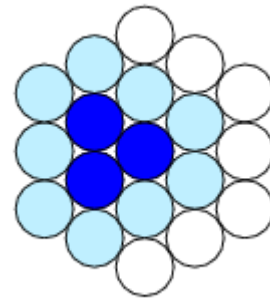
18-Aug-2005 10:09:28

DAMMIF

- Reimplementation of DAMMIN written in object oriented code
- About 25 to 40 times faster (about 1-2 min for fast run on a PC)
- Make use of multiple CPU
- Use adaptive search volume



DAMMIN

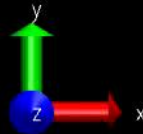
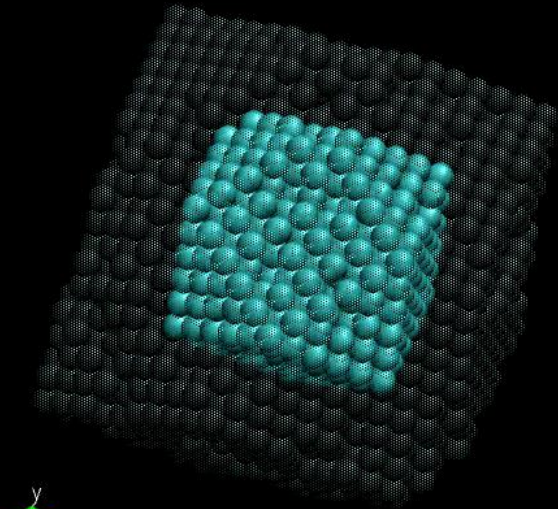


DAMMIF

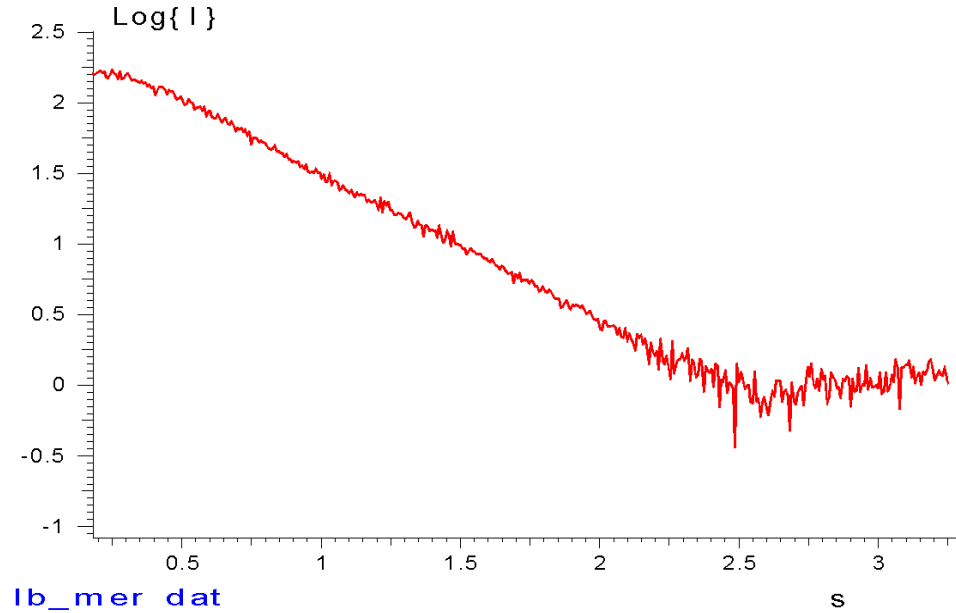
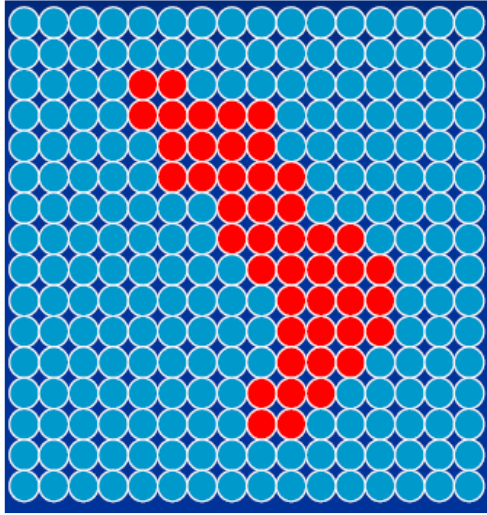
At the current iteration:

- dark blue particle, might become solvent
- light blue solvent, might become particle
- white solvent, won't change

DAMMIF in action

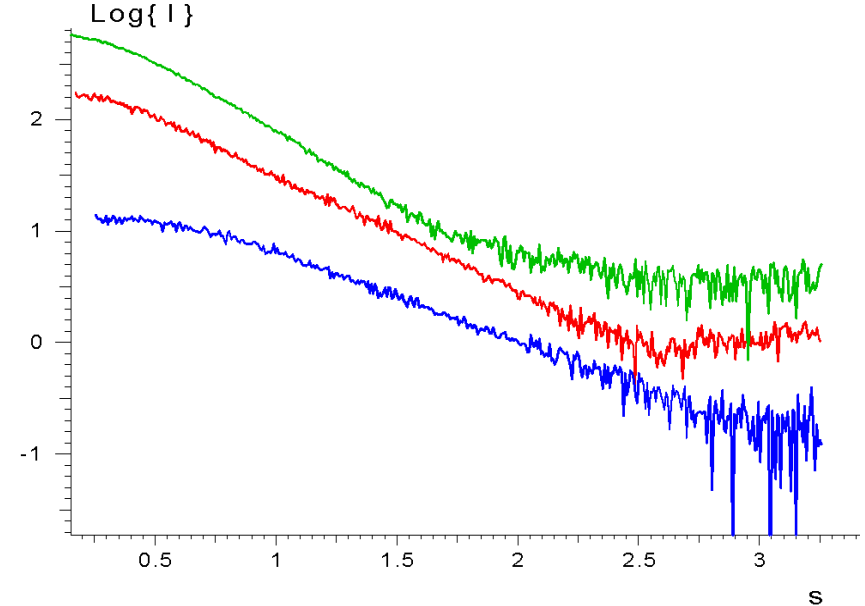
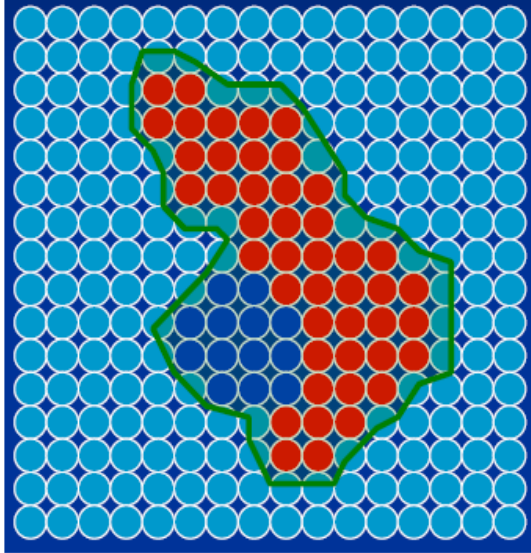


Shape analysis for multi-component systems: principle



One component, one scattering pattern:
“normal” shape determination

Shape analysis for multi-component systems: principle



Many components, many scattering patterns: shape and internal structure

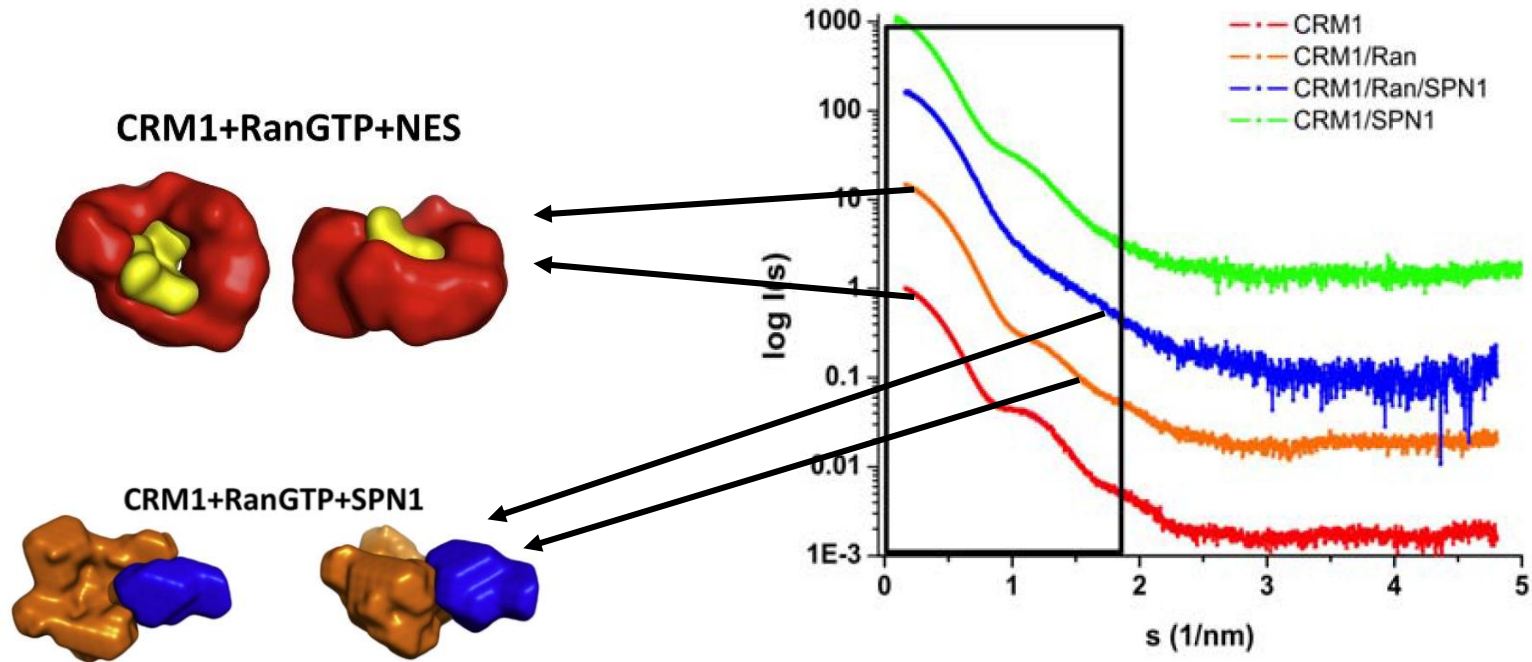


Svergun, D.I. (1999) Biophys. J. **76**, 2879-2886

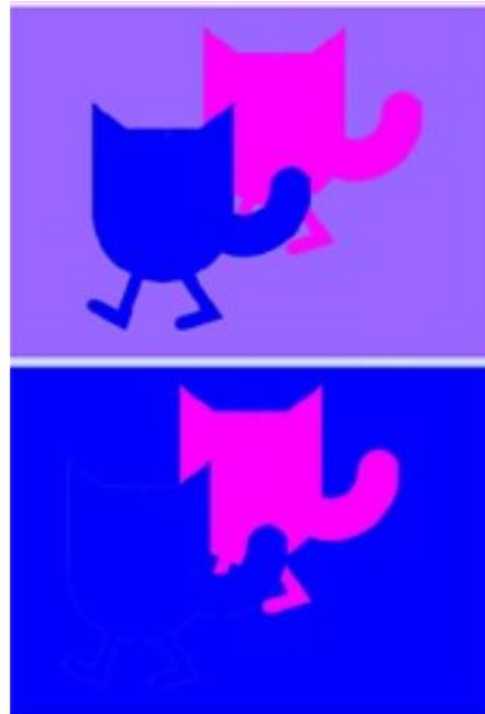
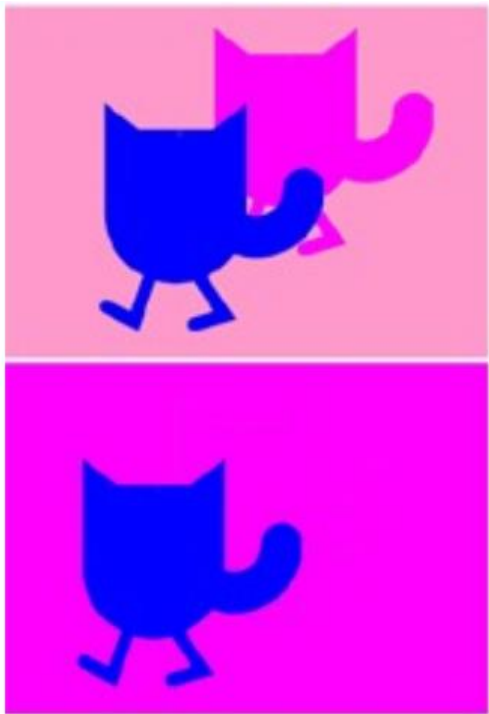
Svergun, D.I. & Nierhaus, K.H. (2000) J.



Example multi-component system



This approach is very useful for contrast matched data.



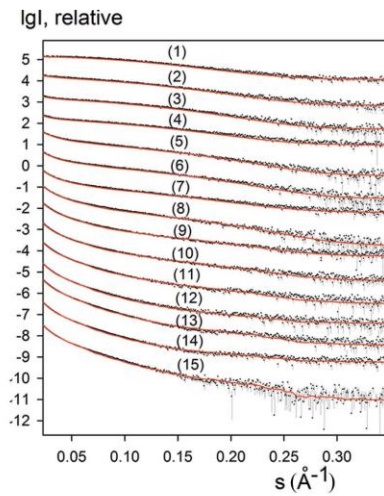
- Dummy atom modelling on mixture with known volume fraction

$$I_k(s) = v_{mk}I_m(s) + v_{ak}I_a(s) + v_{ik}I_i(s), \quad (1)$$

where v_{mk} , v_{ak} , and v_{ik} are the volume fractions of the components, $v_{mk} + v_{ak} + v_{ik} = 1$.

$$F(X) = \chi^2(X) + P(X),$$

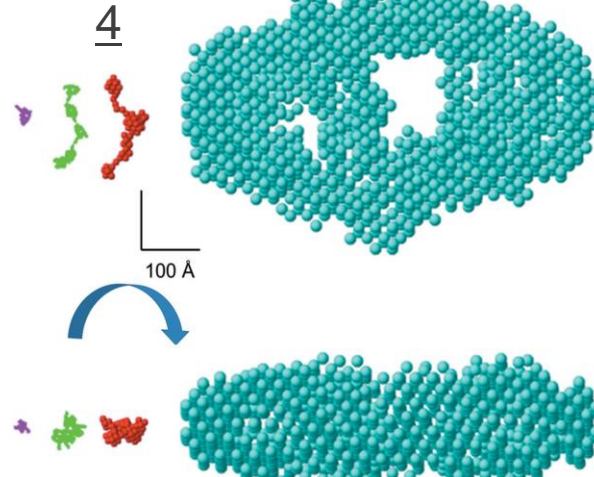
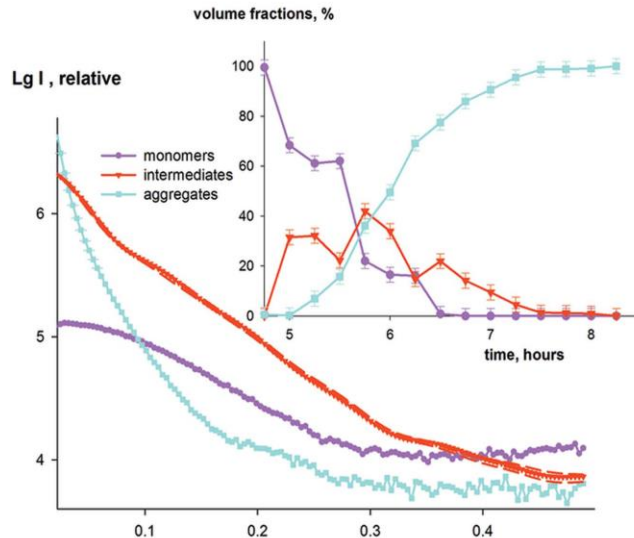
$$F(X) = \sum_{k=1}^K \chi_k^2(X) + \sum_j W_j \times P_j(X).$$

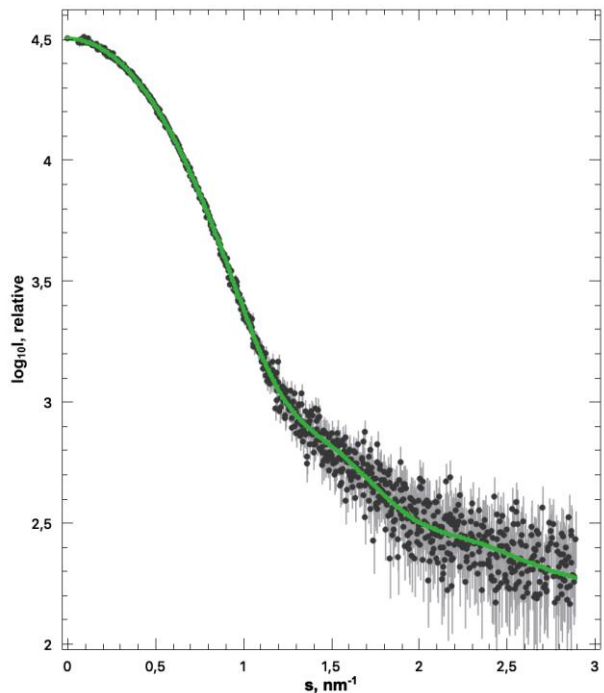
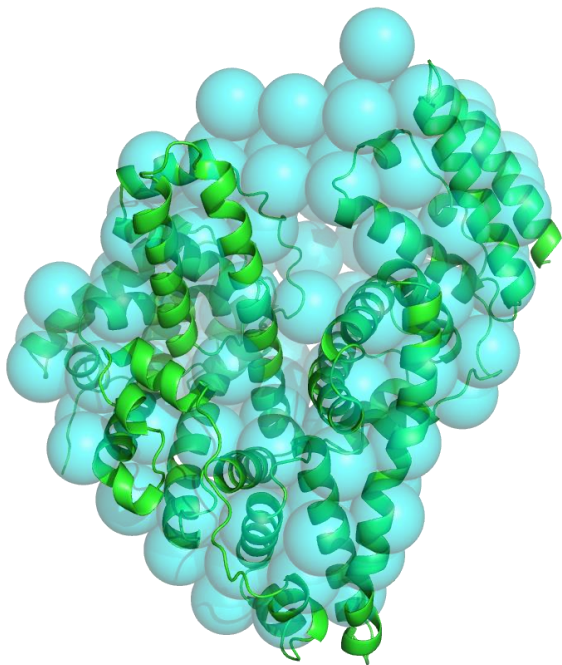


Konarev, P. V. & Svergun, D. I. (2018). IUC

Vestergaard B, Groenning M, Roessle M, Kastrup JS, de Weert Mv, et al. (2007) A Helical Structural Nucleus Is the Primary Elongating Unit of Insulin Amyloid Fibrils . PLOS Biology 5(5): e134.

<https://doi.org/10.1371/journal.pbio.0050134>

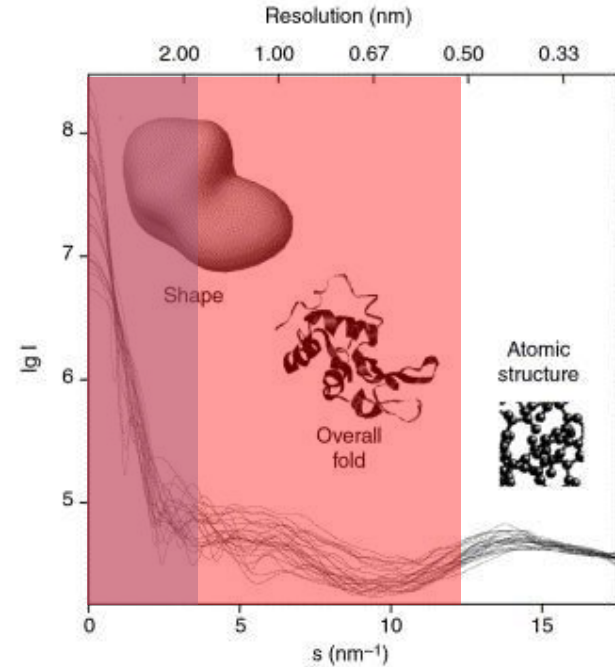
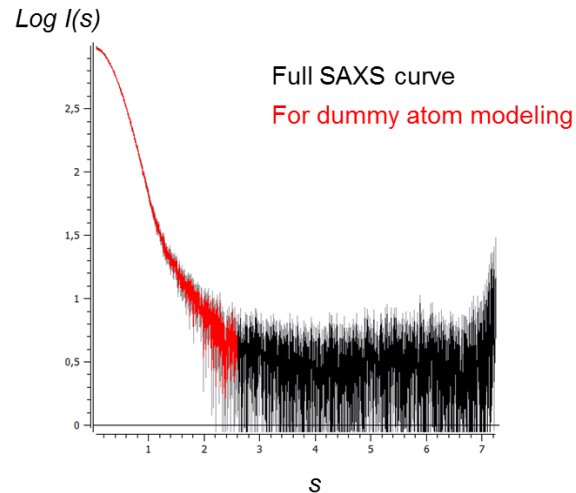




- Fit experimental data directly
 - Debye formula
 - Only penalty: minimize surface area
 - Runtime: minutes
-
- Example:
 - BSA monomer from SEC-SAXS
 - 967 experimental data points
 - Model superposition to monomer of 4F5S
 - Fit: red. $\chi^2=0.9$
 - MW: ~70 kDa
 - D_{\max} : ~9.0 nm

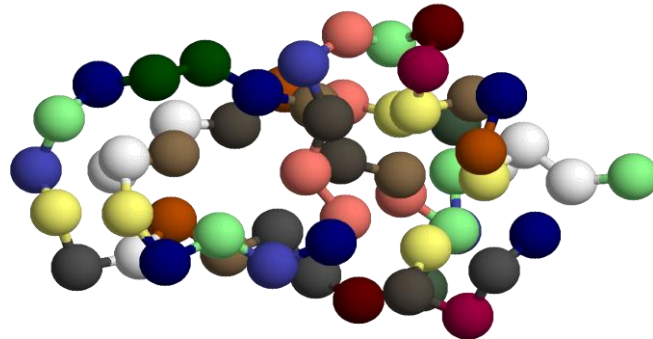
Resolution limit of dummy atom model

- For dummy atom models, the electron density within the protein is considered as homogeneous



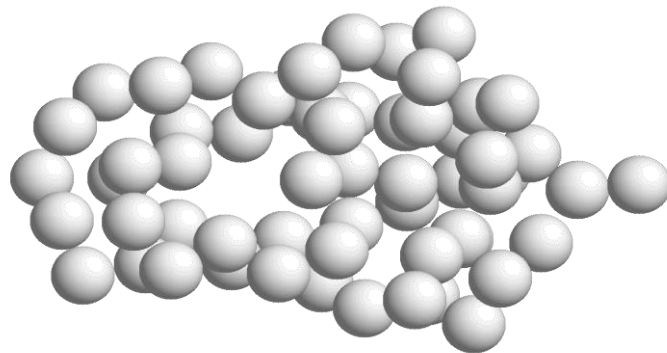
Dummy residue models

- Proteins typically consist of folded polypeptide chains composed of amino acid residues
- At a resolution of 0.5 nm each amino acid can be represented as one entity (dummy residue)



Dummy residue models

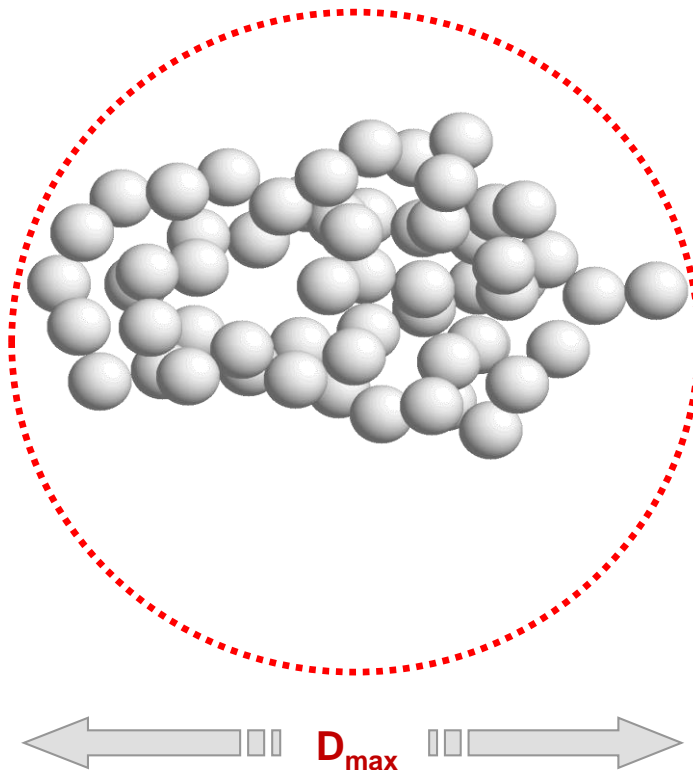
- Proteins typically consist of folded polypeptide chains composed of amino acid residues
- At a resolution of 0.5 nm each amino acid can be represented as one entity (dummy residue)
- In GASBOR a protein is represented by an ensemble of K dummy residues that are
 - Identical
 - Have no ordinal number
 - For simplicity are centered at the $C\alpha$ positions



Dummy residue models

GASBOR

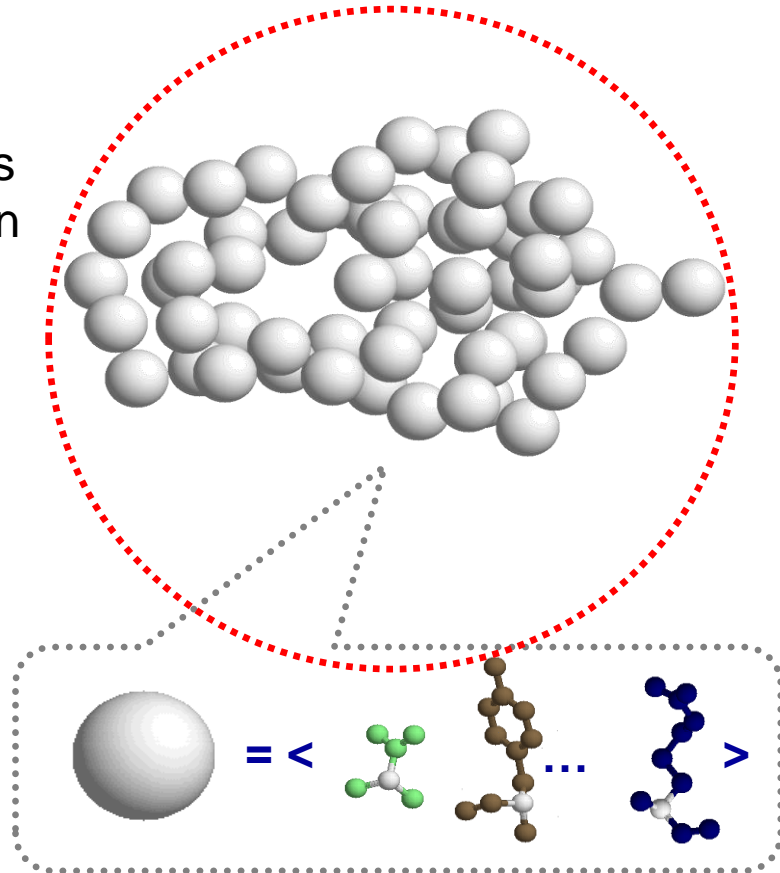
- GASBOR finds coordinates of K dummy residues within its search volume (red)



Dummy residue models

GASBOR

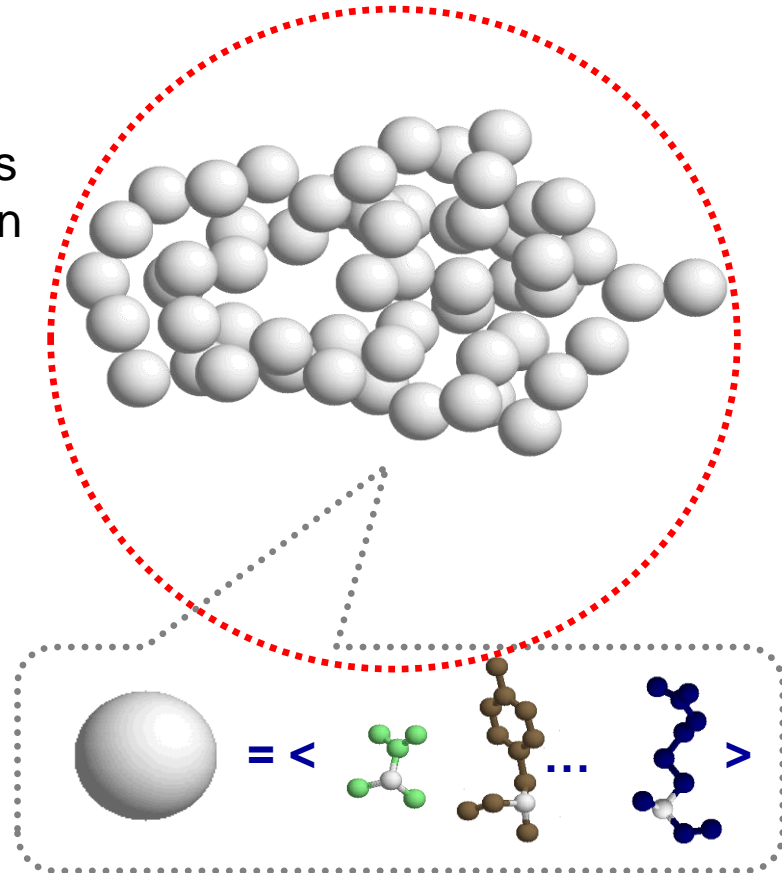
- GASBOR finds coordinates of K dummy residues within its search volume (red)
- Requires polypeptide chain-compatible arrangement of dummy residues



Dummy residue models

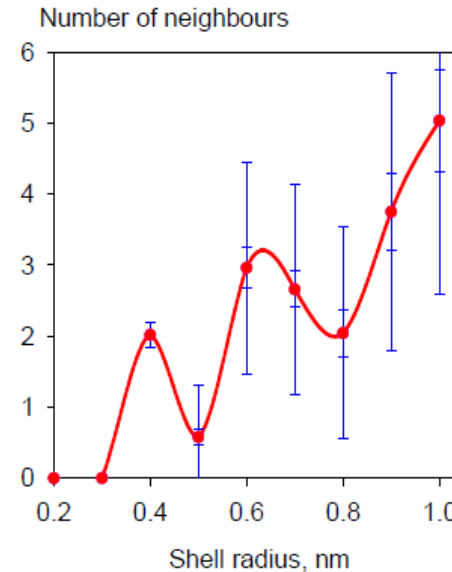
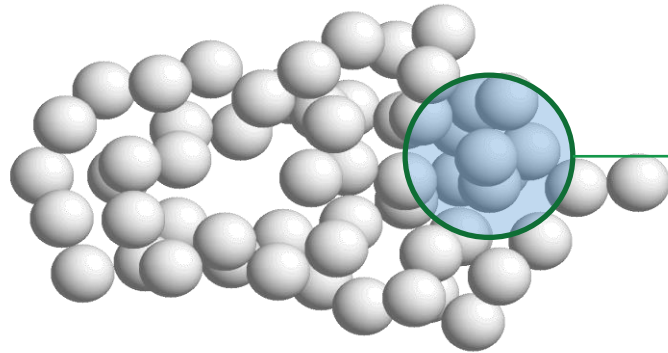
GASBOR

- GASBOR finds coordinates of K dummy residues within its search volume (red)
- Requires polypeptide chain-compatible arrangement of dummy residues
- Scattering is computed using the Debye (1915) formula

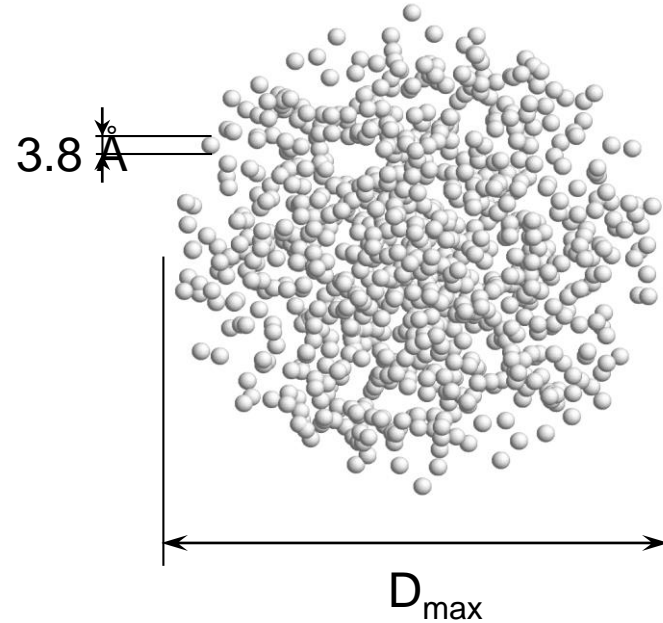


Distribution of neighbours

- Excluded volume effects and local interactions lead to a characteristic distribution of nearest neighbors around a given residue in a polypeptide chain

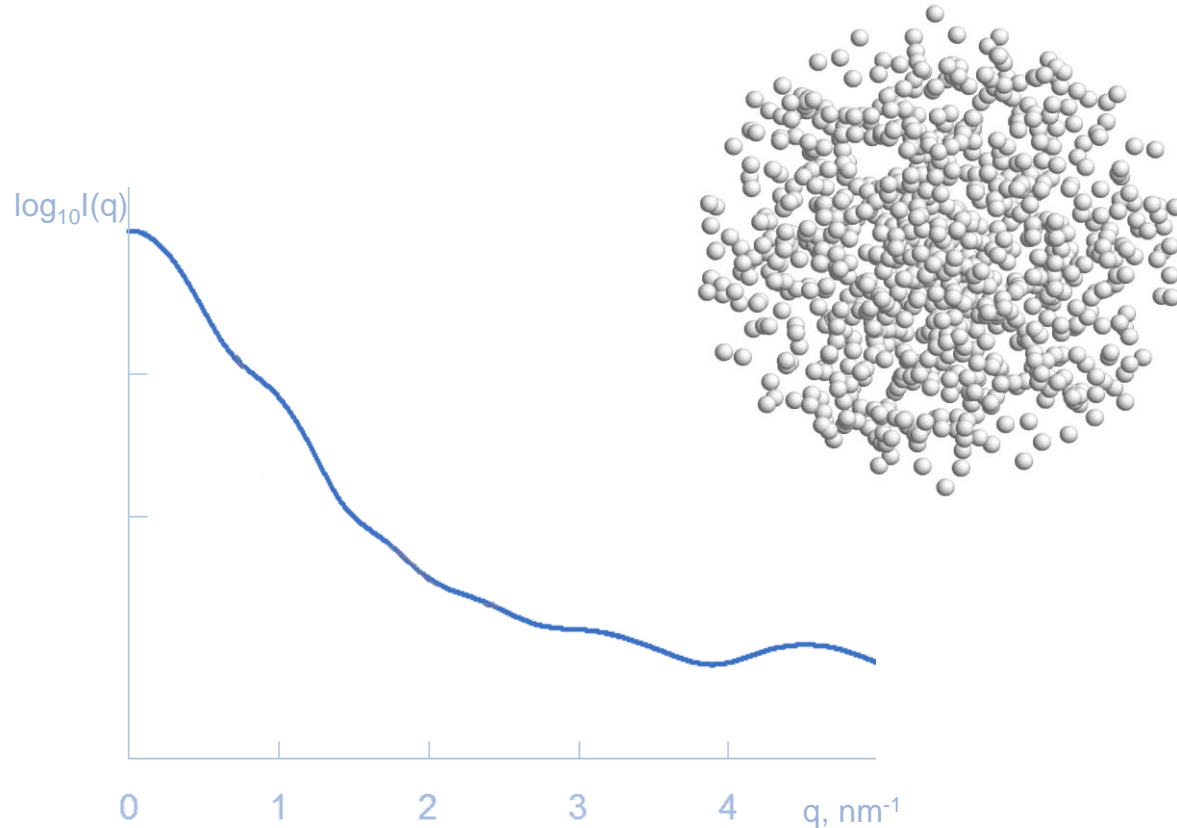


Ab initio reconstruction: dummy residue modelling

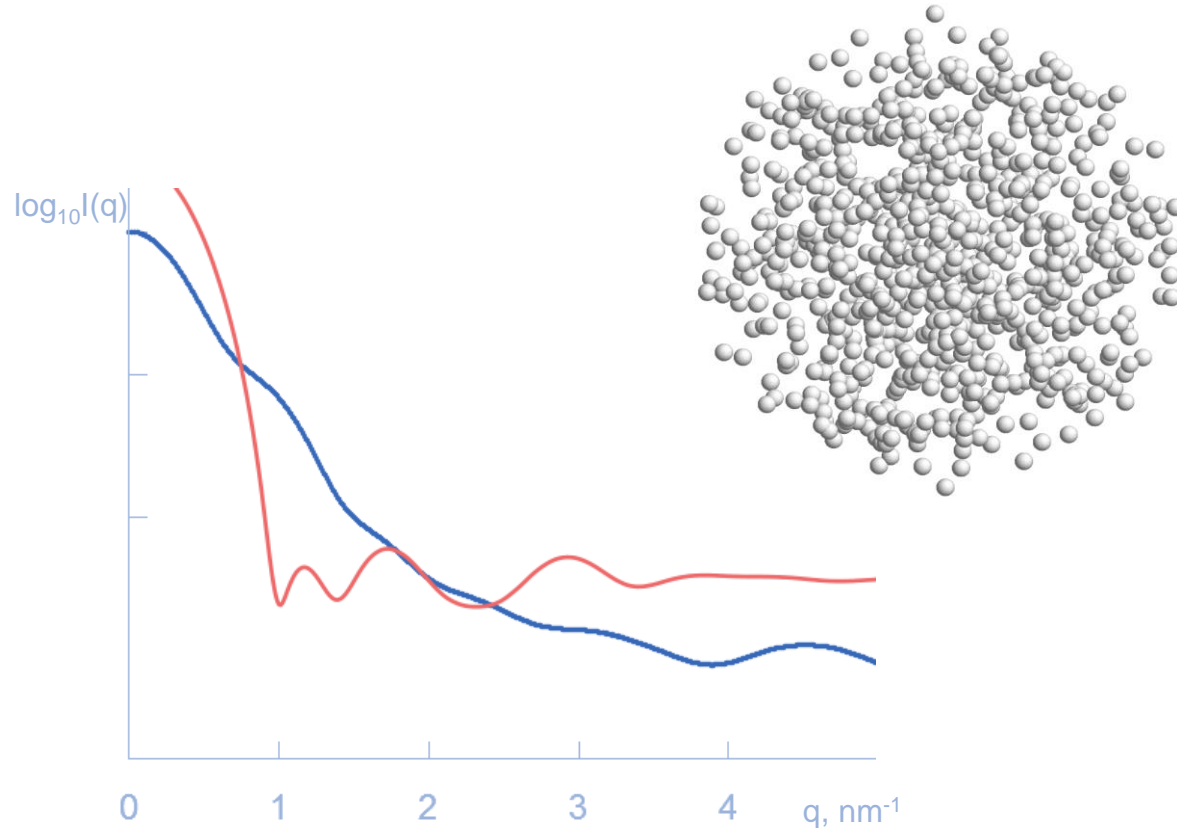


Svergun, D.I., Petoukhov, M.V, Koch, M.H.J. (2001)
Biophys J 80, 2946–2953.

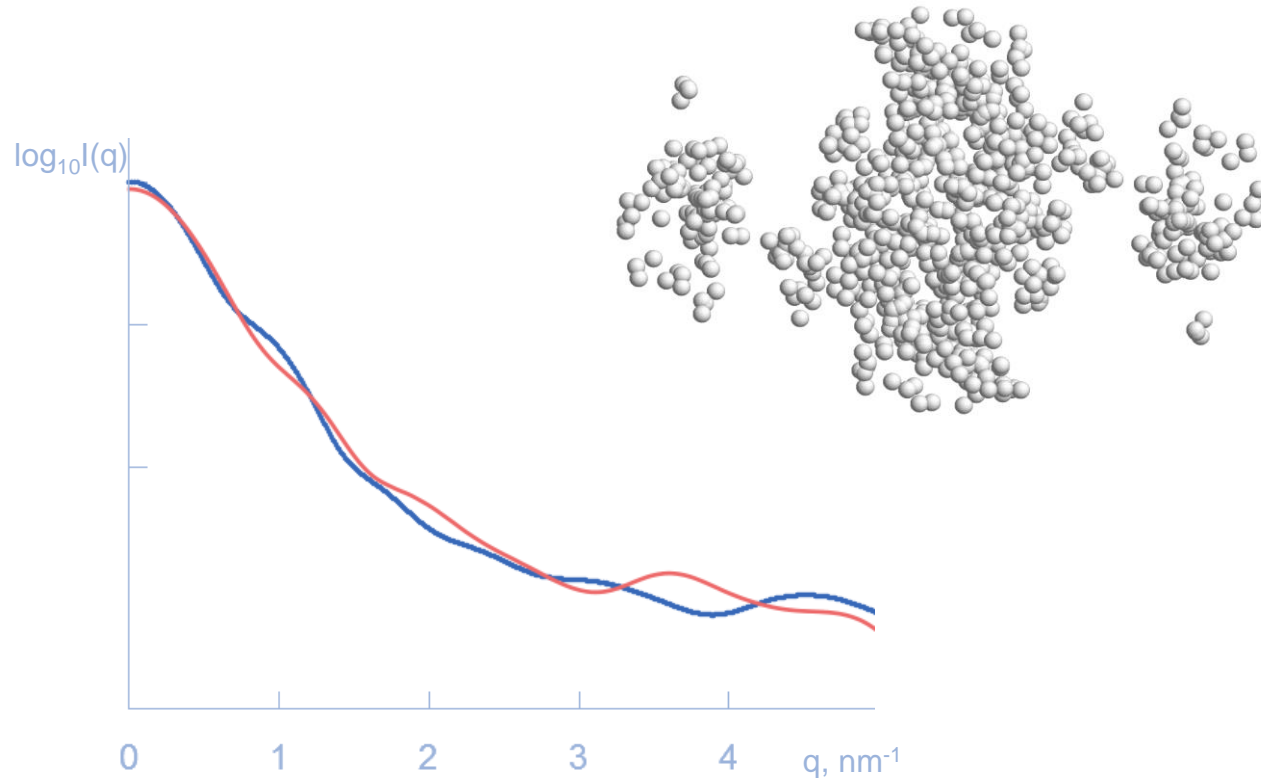
Ab initio reconstruction: dummy residue modelling



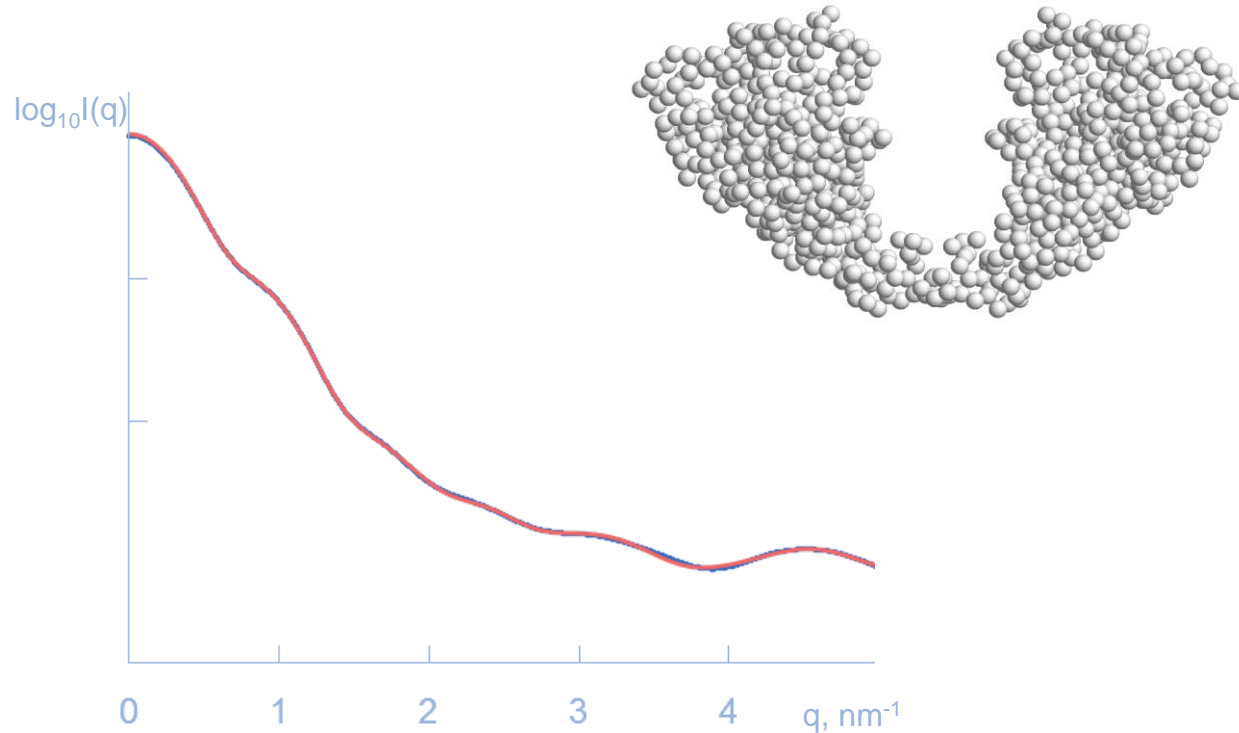
Ab initio reconstruction: dummy residue modelling



Ab initio reconstruction: dummy residue modelling



Ab initio reconstruction: dummy residue modelling



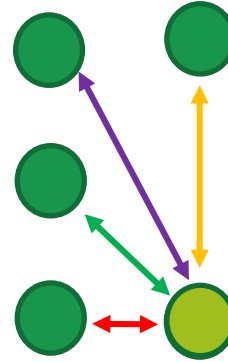
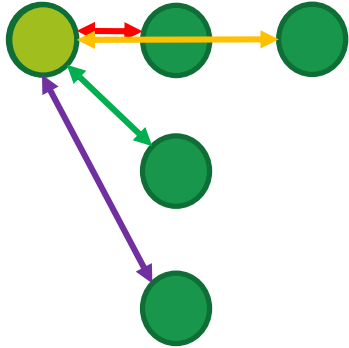


Svergun, D.I., Petoukhov, M.V, Koch, M.H.J. (2001)
Biophys J 80, 2946–2953

- Use dummy residues with average density (**fixed radius of 1.9 Å**)
- Number(dummy residues) = Number(AA) = K (**fixed number**)
- Distances to neighbor “residues” like for proteins
- Fixed search space
- Scattering is computed using Debye formula
- Use **higher angles** (up to 12 nm^{-1})
- Only for proteins **smaller than 660 kDa**

Words of caution

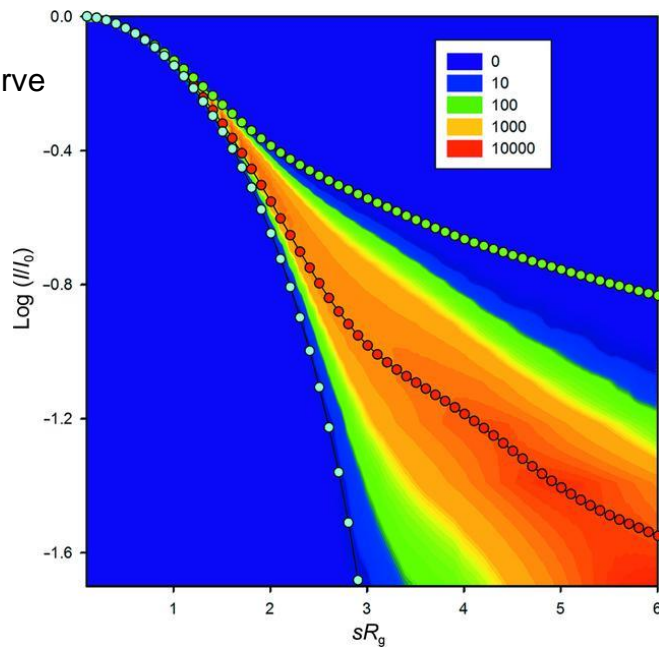
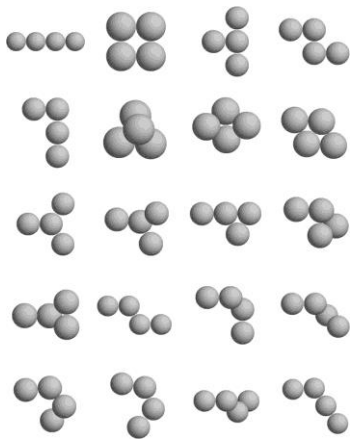
Ambiguity in SAXS: C-T



Measure of the ambiguity



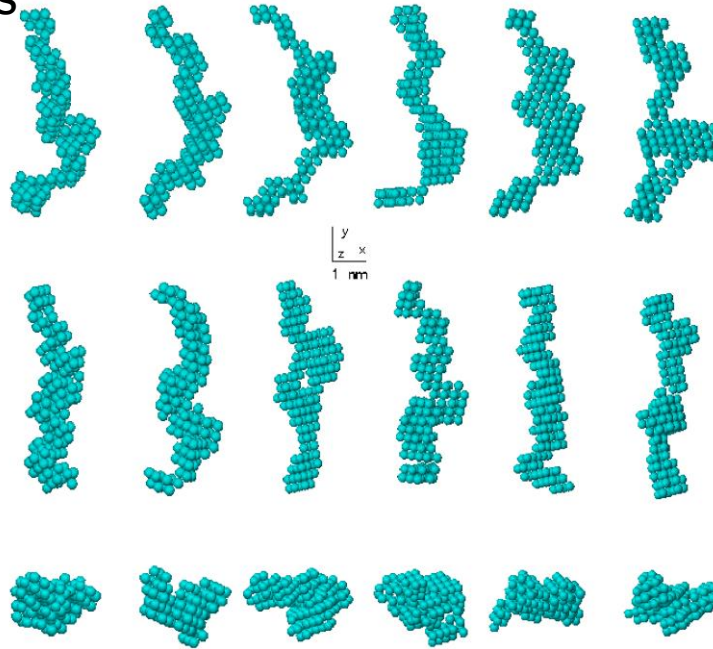
Quantitative measure of the ambiguity from the SAXS curve



- 14000 shape topologies generated (up to seven beads closely packed on hexagonal grid).
- Scattering curves computed and rescaled to keep only shape topology information.
- Scattering map computed from these curves.
- By plotting the experimental SAXS curves on the map, ambiguity intrinsic to the curve can be estimated .

Ab initio model validity

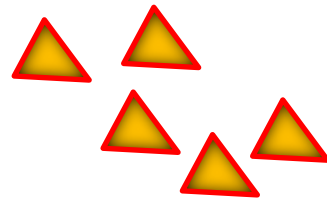
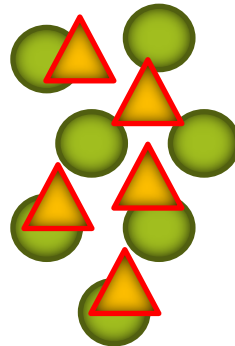
Shape determination of 5S RNA: six DAMMIN models yielding identical fits



Ab initio model validity

SUPCOMB

- Superimpose models by minimizing the Normalized Spatial Discrepancy (NSD)
- Steps
 - Principle axes alignment
 - Gradient minimization
 - Local grid search



SUPALM

- Aligns models in Fourier space using spherical harmonics representation
- For MDa size particles – about 10 times faster than SUPCOMB

Reduce ambiguity of ab-initio model

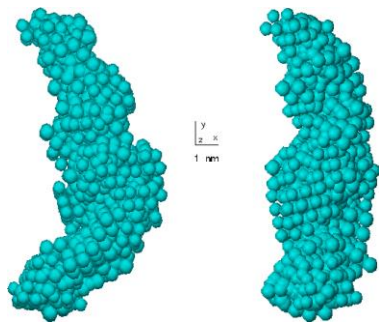
- To reduce ambiguity, several models are built, averaged and compared



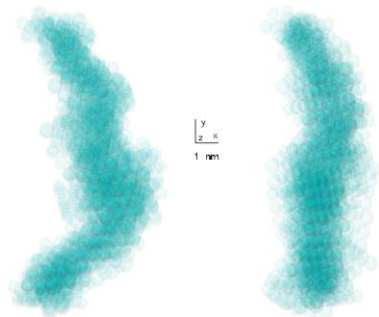
- $NSD_i = \langle NSD_{ij} \rangle_j$
- $\text{MIN}(NSD_i) \Rightarrow \text{typical (most prob)}$
- $\langle NSD \rangle + 2 \sigma (NSD) \Rightarrow \text{threshold}$

| NSD | 1 | 2 | ... | ... | i | ... | j | ... | N |
|-----|---|---|-----|-----|-------------------|-----|-------------------|-----|---|
| 1 | | | | | | | | | |
| 2 | | | | | | | | | |
| ... | | | | | | | | | |
| ... | | | | | | | | | |
| i | | | | | | | NSD _{ij} | | |
| ... | | | | | | | | | |
| j | | | | | NSD _{ji} | | | | |
| ... | | | | | | | | | |
| N | | | | | | | | | |

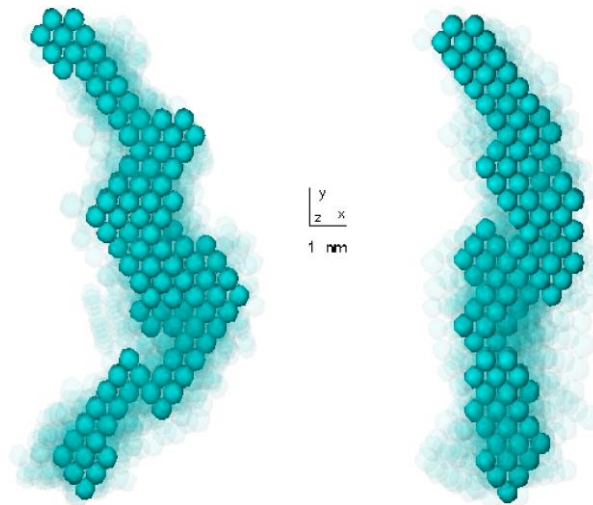
Model validity



5S RNA – Solution spread region



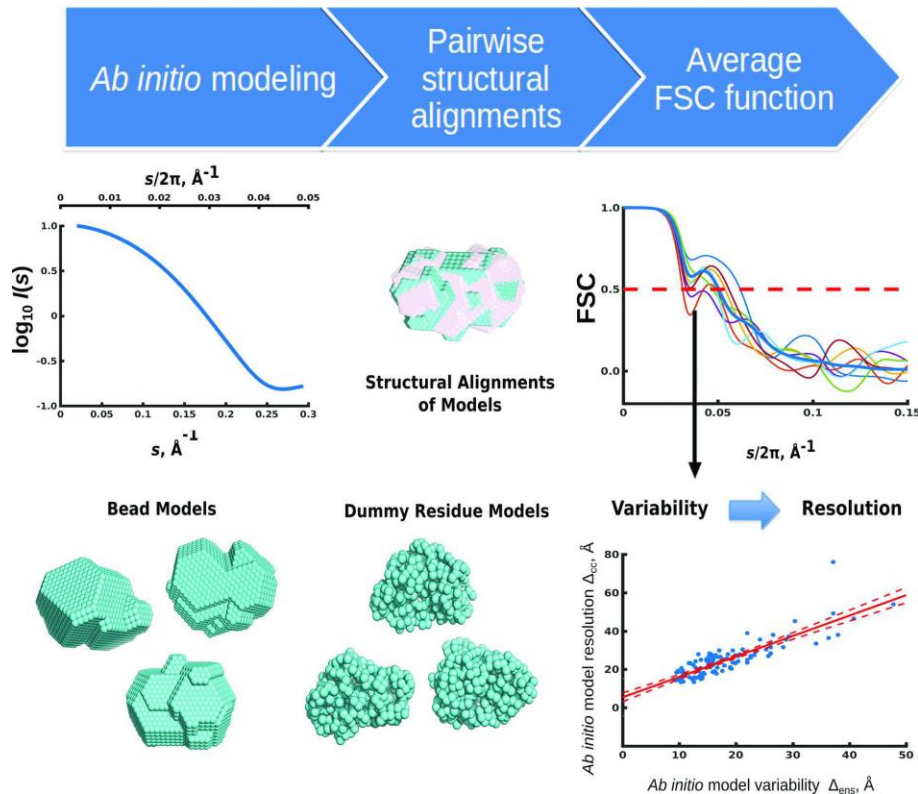
5S RNA – Most Populated Volume



5S RNA – Final Solution
within the Spread Region



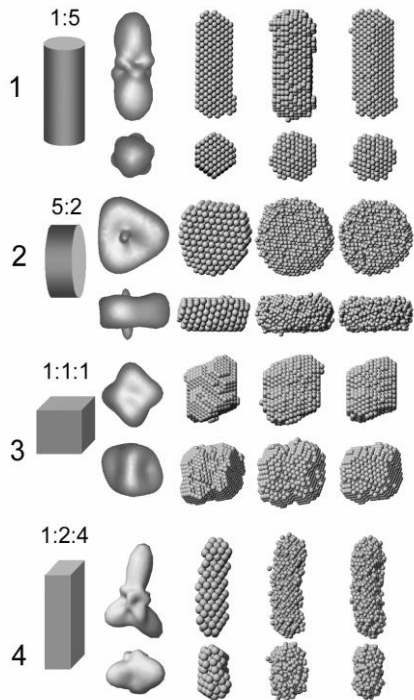
Resolution of ab initio models



- “Measure ambiguity to estimate resolution”
- Resolution estimated from a set of (10-20) bead model.
- Model compared and aligned.
- Measure of the variability gives an estimation of the resolution

Can all shapes be reconstructed by ab initio modelling?

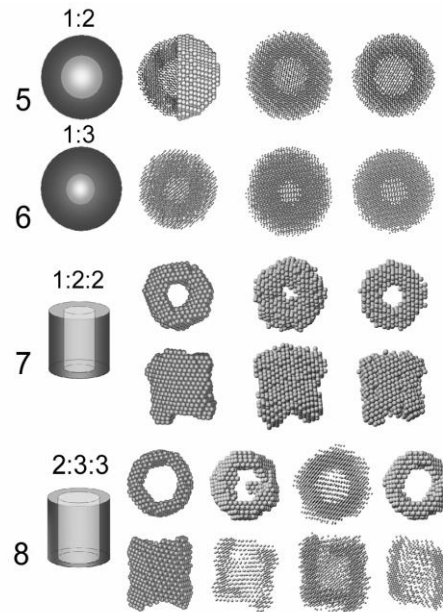
Volkov, V. V. & Svergun, D. I. (2003). J. Appl. Cryst. 36, 860-864.



Globular solid particles

solid bodies with moderate anisotropy
(elongated particles up to 1:5
and flattened up to 5:2) can be
reliably reconstructed from the
SAXS data.
Mean value NSD : 0.4-0.7

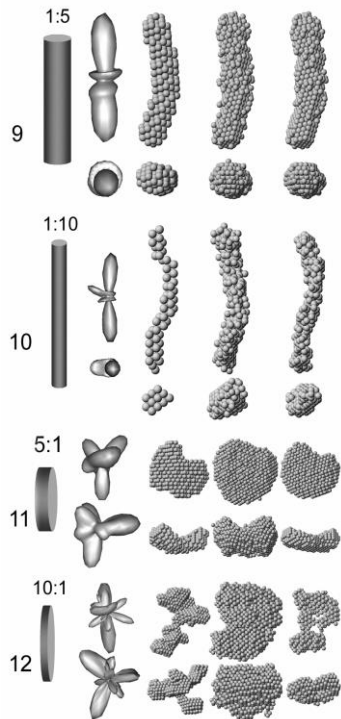
Hollow globular models
can also be well
reconstructed



Hollow globular particles

Can all shapes be reconstructed by ab initio modelling?

Volkov, V. V. & Svergun, D. I. (2003). J. Appl. Cryst. 36, 860-864.

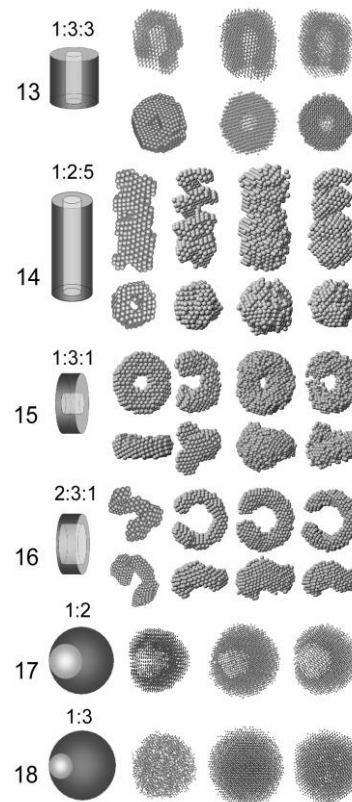


anisotropic solid particles

Shape reconstructions of anisometric particles are less stable and reliable.

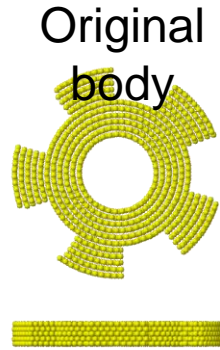
Elongated hollow body: the channels may appear closed from one or both sides
For hollow flattened the resulting shapes may show a helical turn instead of a hollow disk, even after the averaging.

Acentric voids in hollow spheres are only reconstructed if r/R is about 0.5



Hollow anisotropic and acentric particles

Use of symmetry.



Conclusion

- Ab initio methods are powerful tools to build model from SAXS data.
- Ab initio methods always provide good looking models that fit the data (even if they shouldn't → Beware of what data you put in)
- Different kind of models can be built (dummy atom model, dummy residue model, multiphase)
- The models built are of low resolution and have some ambiguity but methods now exist to estimate this ambiguity and resolution
- Further reduce ambiguity → add information

Questions?