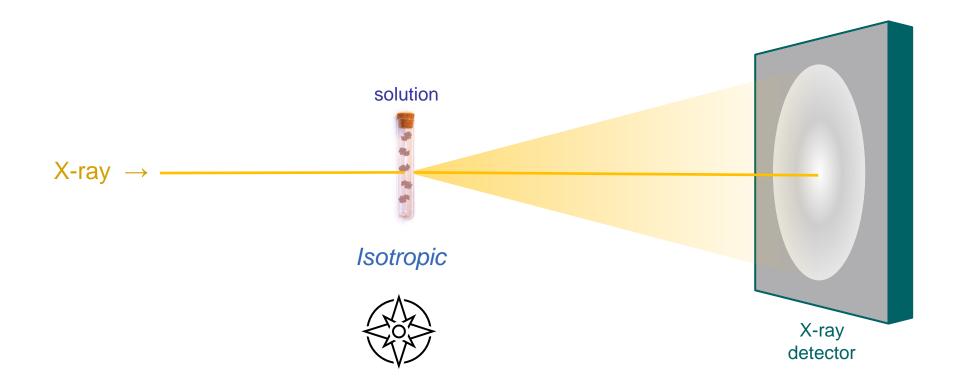
Primary data inspection and reduction with **PRIMUS**: guided tutorial

Stefano Da Vela, Melissa Gräwert, Al Kikhney, Cy Jeffries



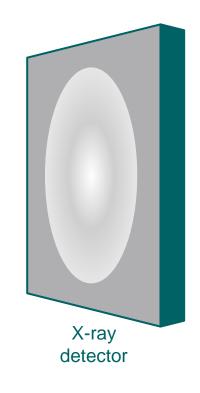
First, a brief summary...

SAXS data collection



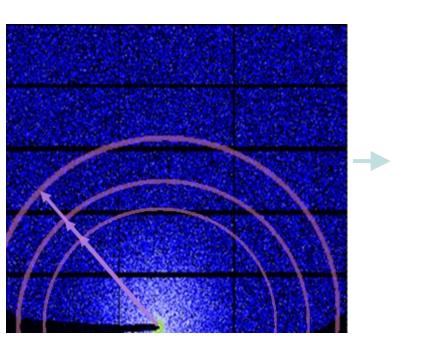
SANS: conceptually similar

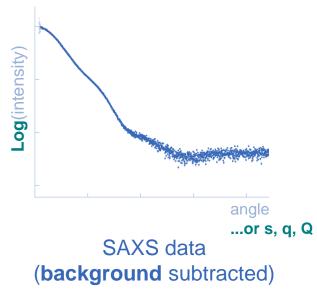
SAXS data collection



Also Isotropic

SAXS data



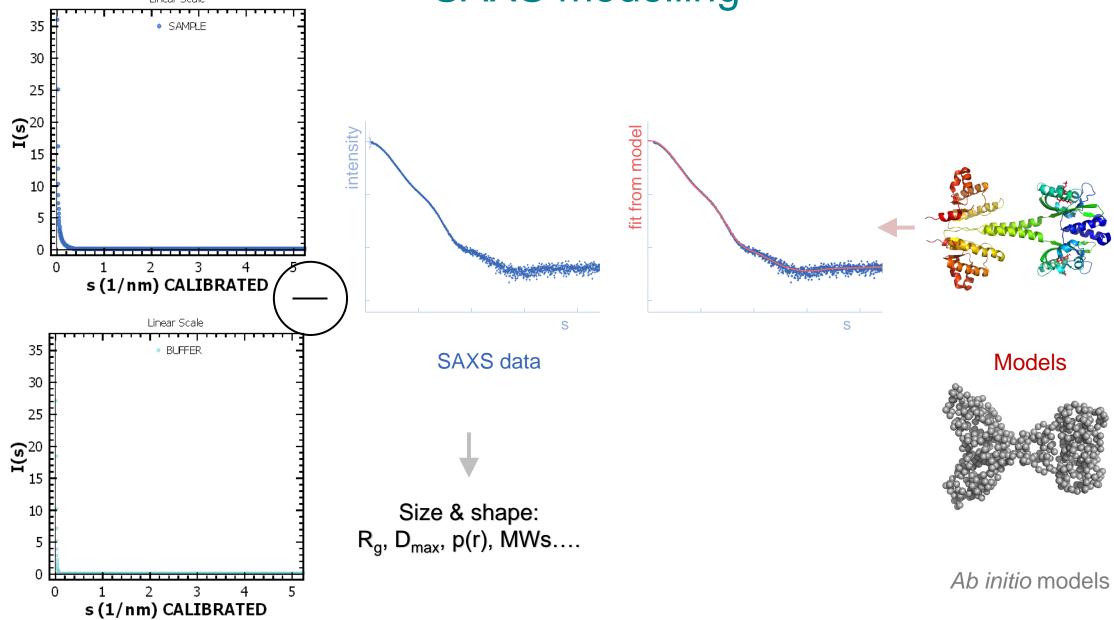


$$|s| = \frac{4\pi \sin\theta}{\lambda}$$

Units: reciprocal of a length (commonly nm⁻¹ or Å⁻¹=0.1*nm⁻¹)

nm scale information

SAXS modelling





- >100 programs:
 - GUI
 - command-line
 - PyMOL plugin
- ATSAS online



- Commercial licenses:
 - biosaxs.com/software
- Academic licenses: www.embl-hamburg.de/biosaxs/download.html

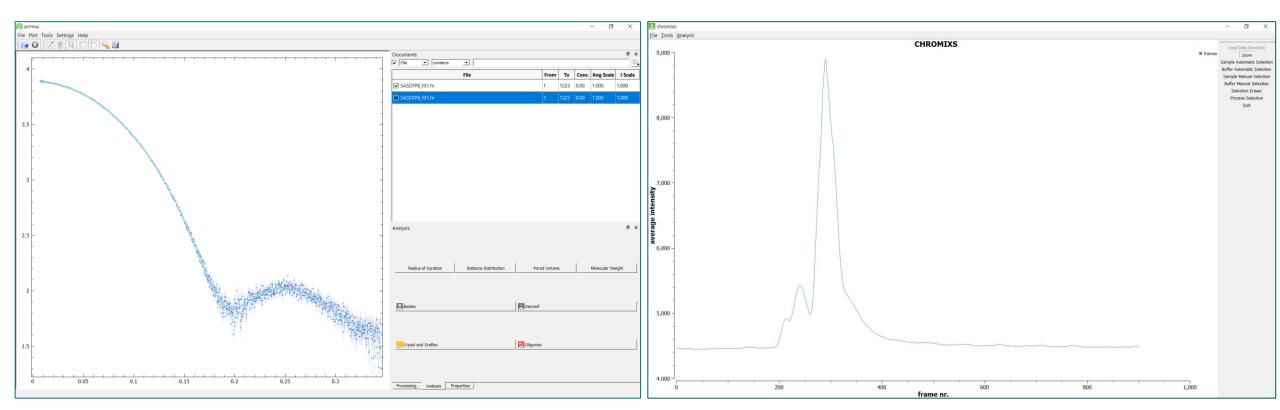
K. Manalastas-Cantos, P.V. Konarev, N.R. Hajizadeh, A.G. Kikhney, M.V. Petoukhov, D.S. Molodenskiy, A. Panjkovich, H.D.T. Mertens, A. Gruzinov, C. Borges, C.M. Jeffries, D.I. Svergun and D. Franke (2021) **ATSAS 3.0**: Expanded functionality and new tools for small-angle scattering data analysis *J. Appl. Cryst.* 54, 343-355

Focus: Primus, Chromixs



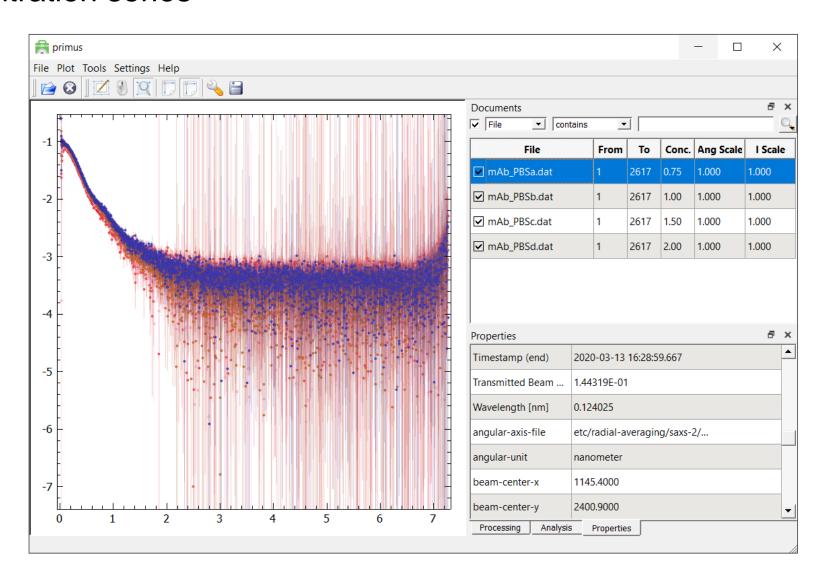


- PRIMUS: plotting SAXS curves and fits, elementary processing, overall parameters, modeling calling other programs of the ATSAS suite
- CHROMIXS: SEC-SAXS data reduction, inspection of chromatograms, overall parameters across SEC peak(s)

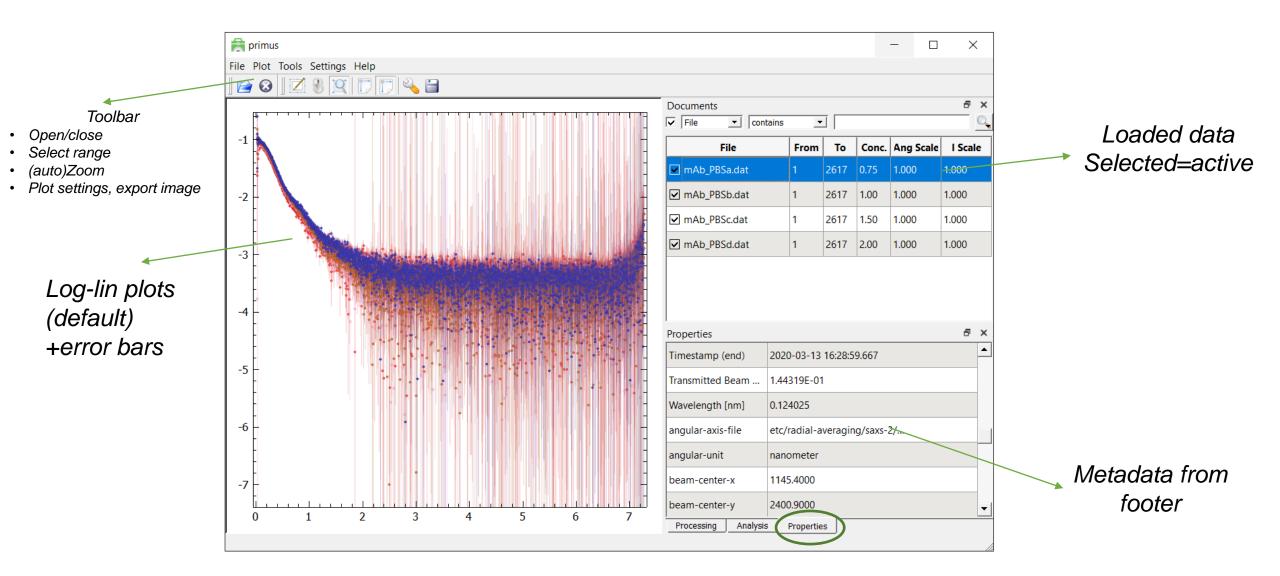


PRIMUS: basics

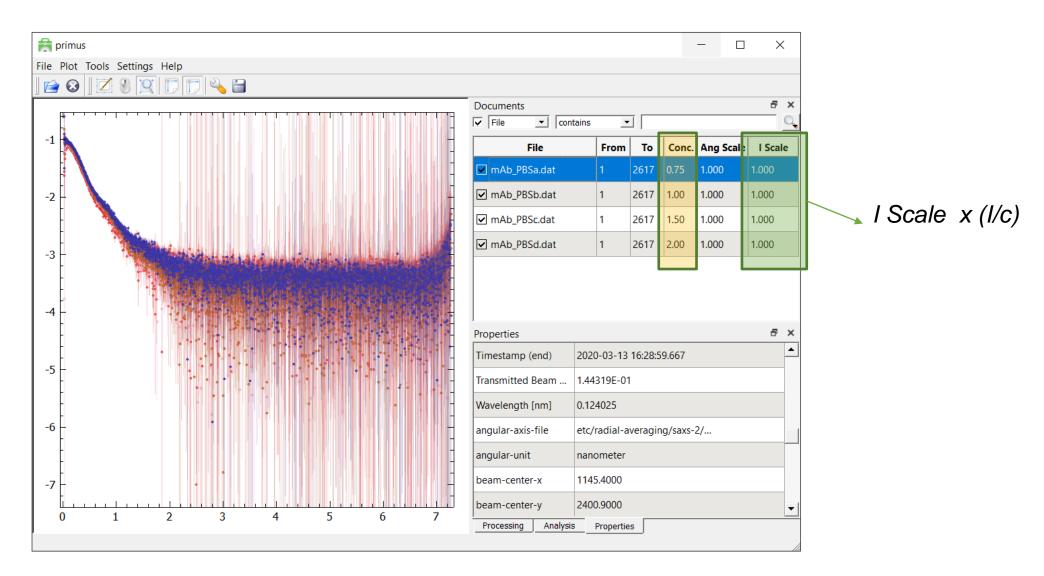
A concentration series



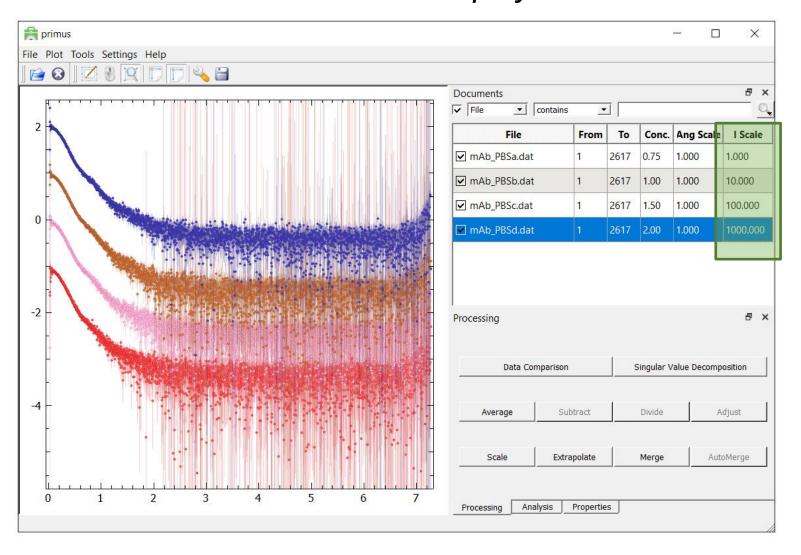
A concentration series



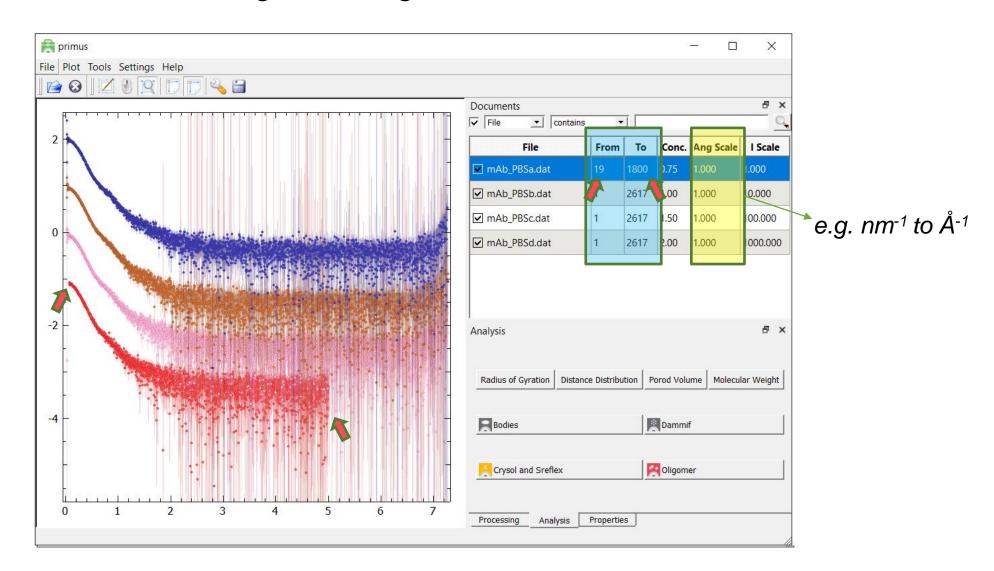
A concentration series: I/c vs s



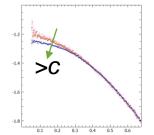
• A concentration series: I/c vs s shifted for display



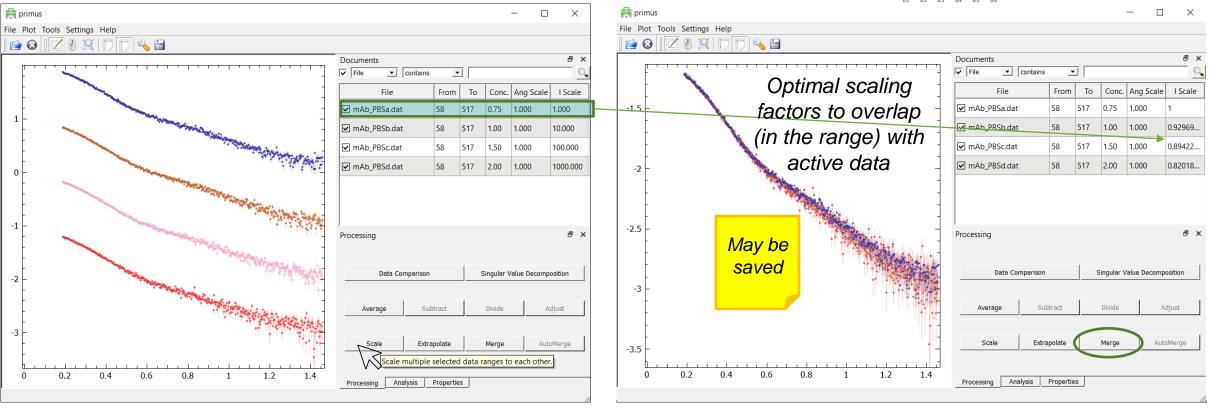
A concentration series: range and angular scale



A concentration series: scaling, comparing low angles

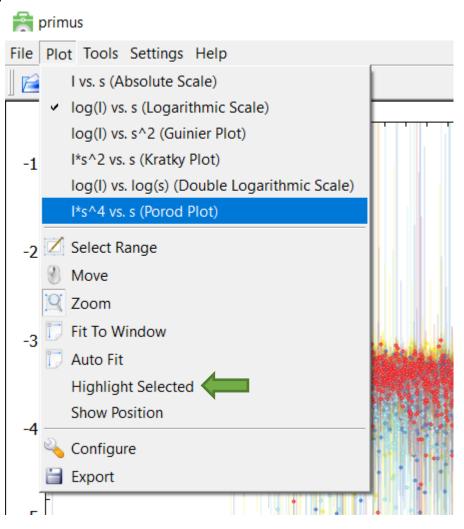


e.g. detection of structure factor or self-association

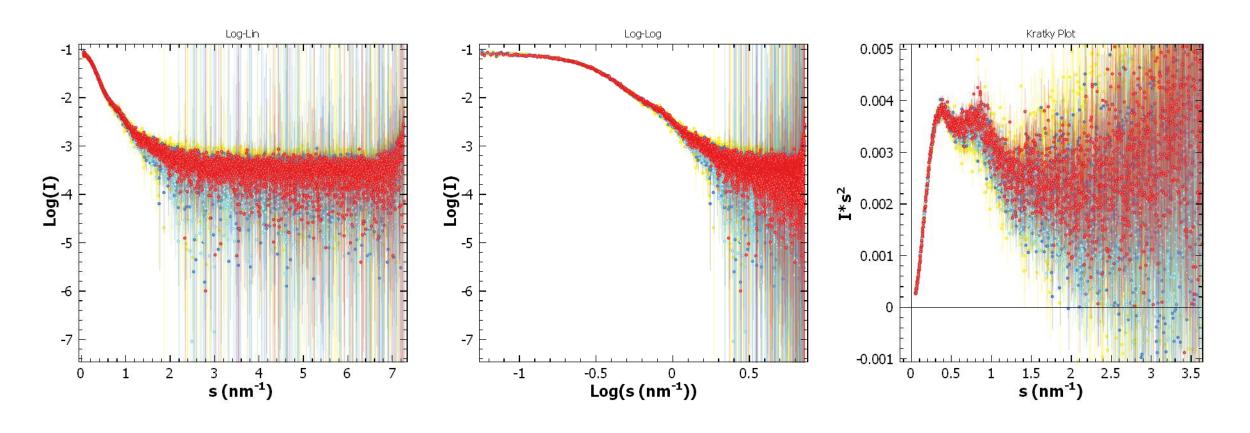


Rule-of-thumb: choose highest concentration curve (higher signal-to-noise) with overlap to all the lowest concentration ones for parameters evaluation and modeling. Or: merge/extrapolate.

Alternatives for plotting

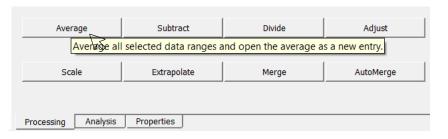


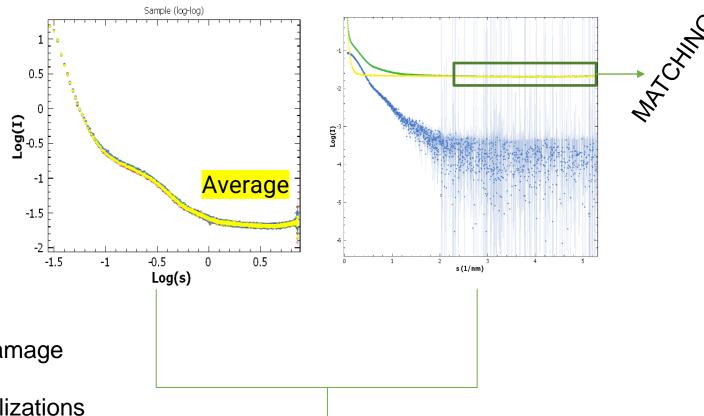
Alternatives for plotting most commonly used for Bio-SAXS



Same data in 3 plotting modes

More math with curves:



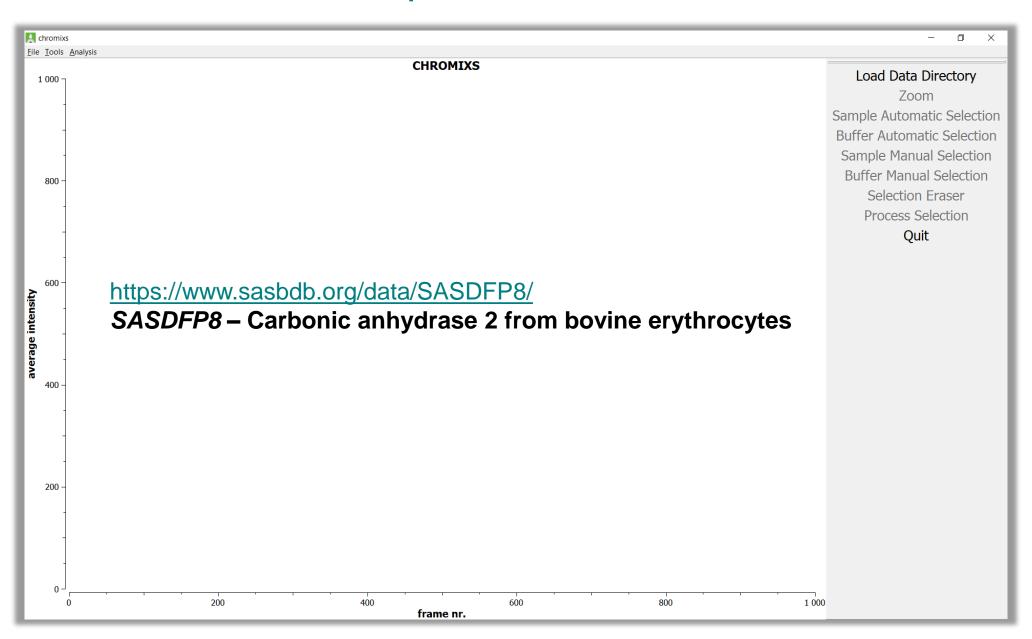


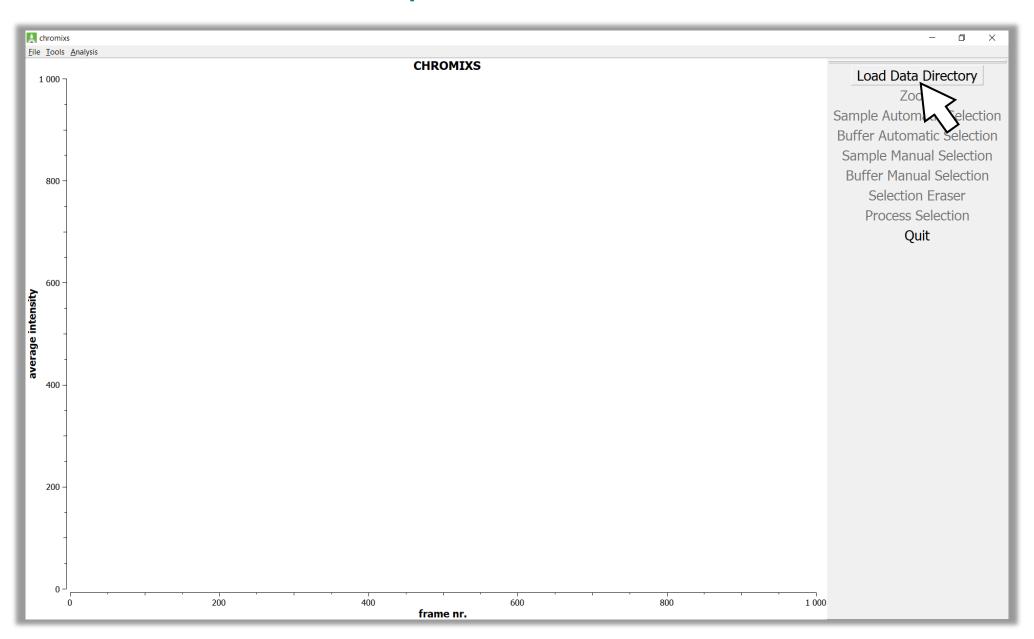
Mostly automated

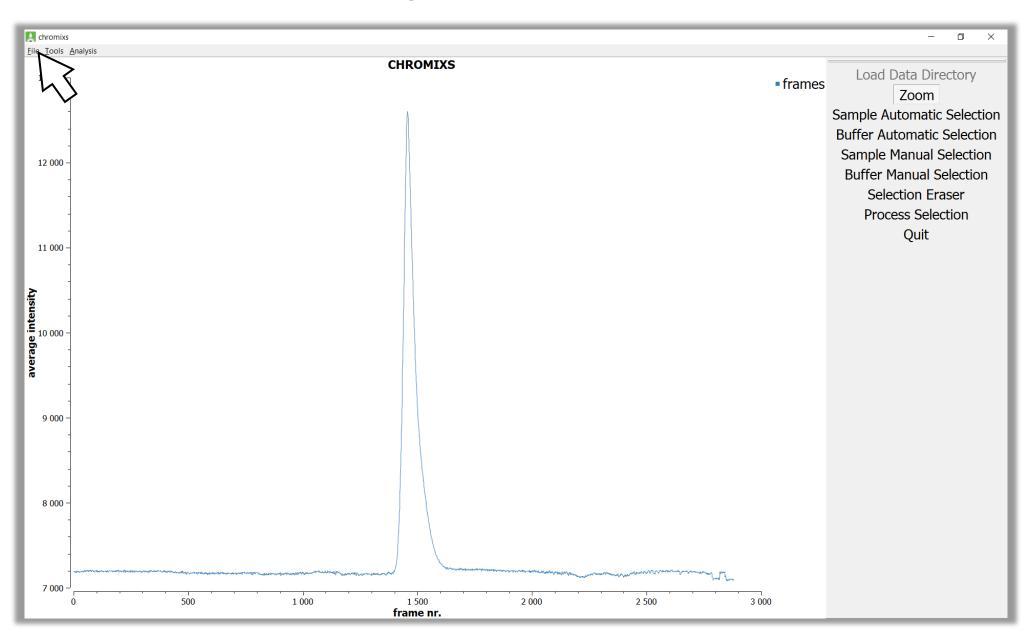
- Averaging exposures and checking for filling/damage
- Subtracting background manually
- Compensating inaccurate concentration normalizations
- Experimental structure factors

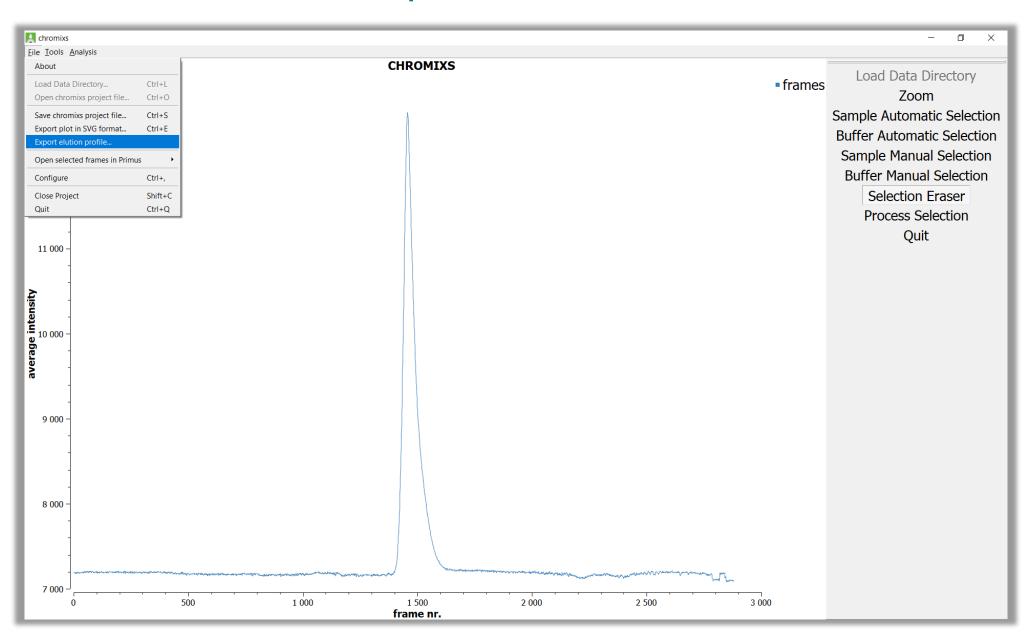
•

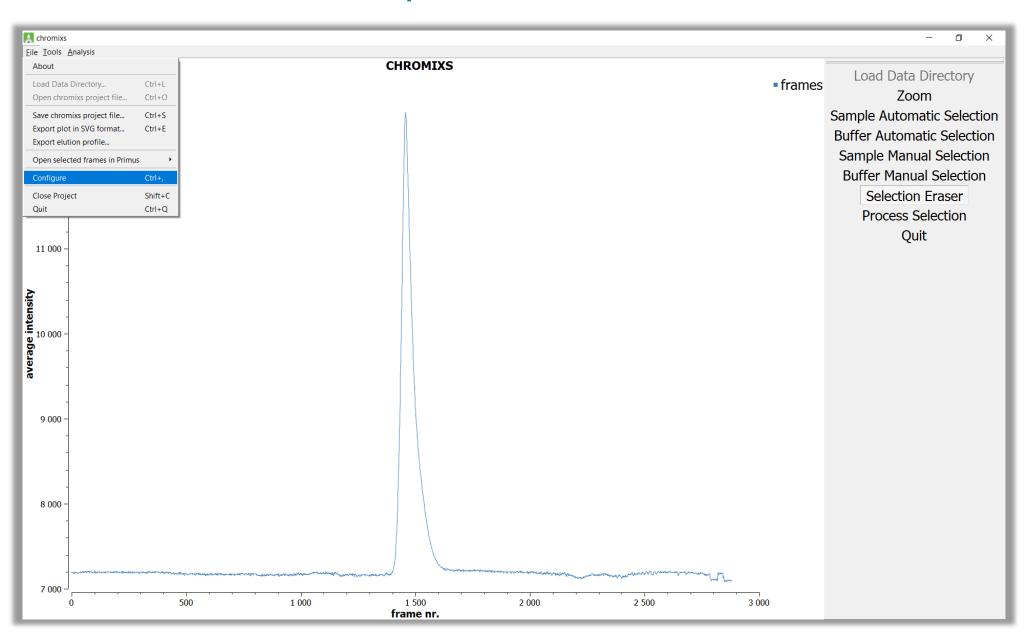
CHROMIXS: SAXS curves from SEC-SAXS

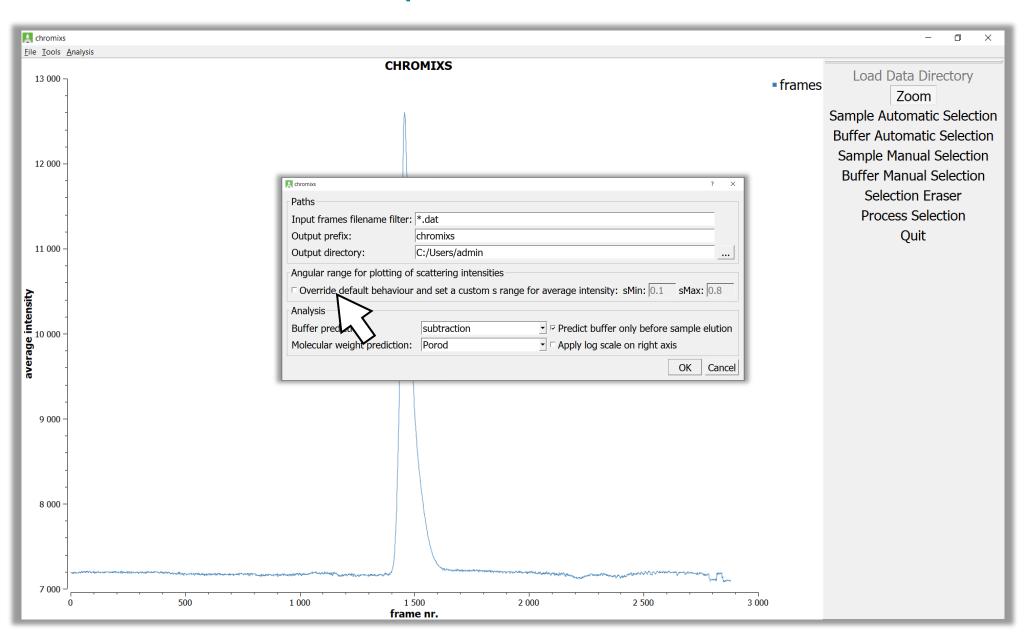


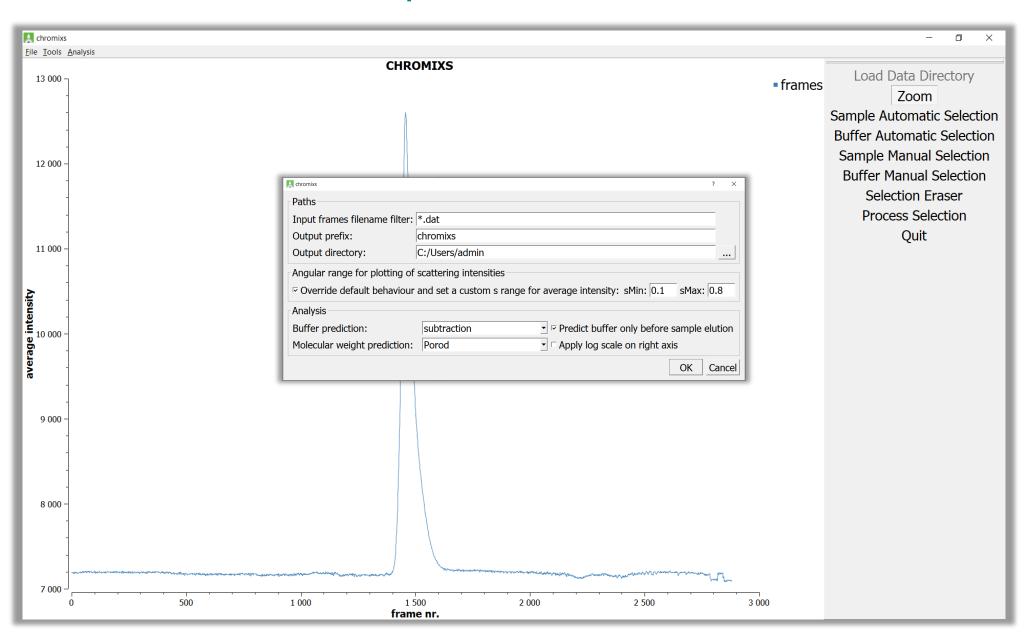


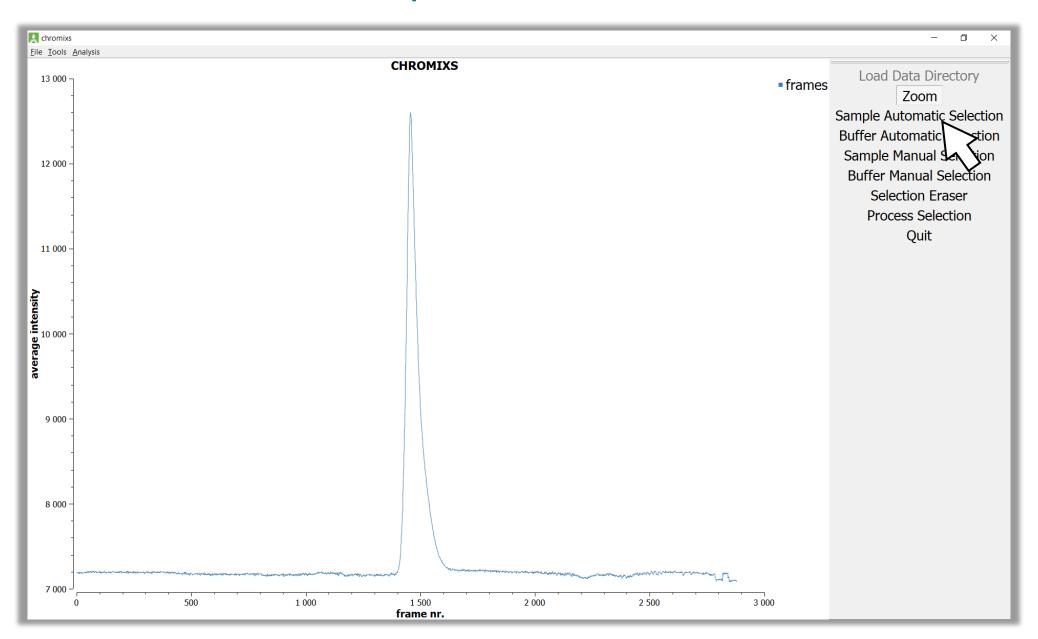


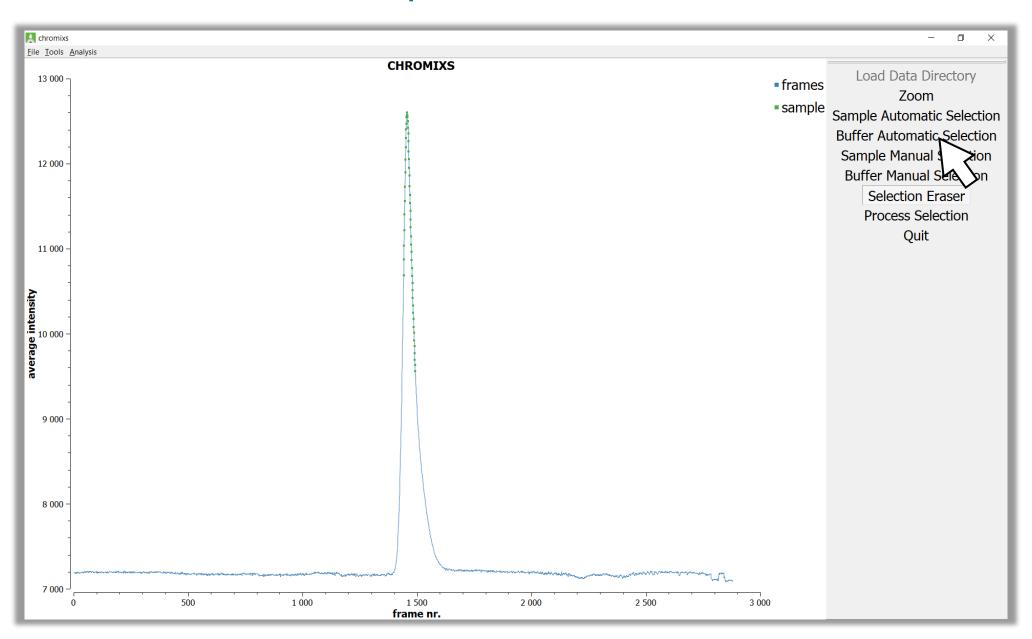


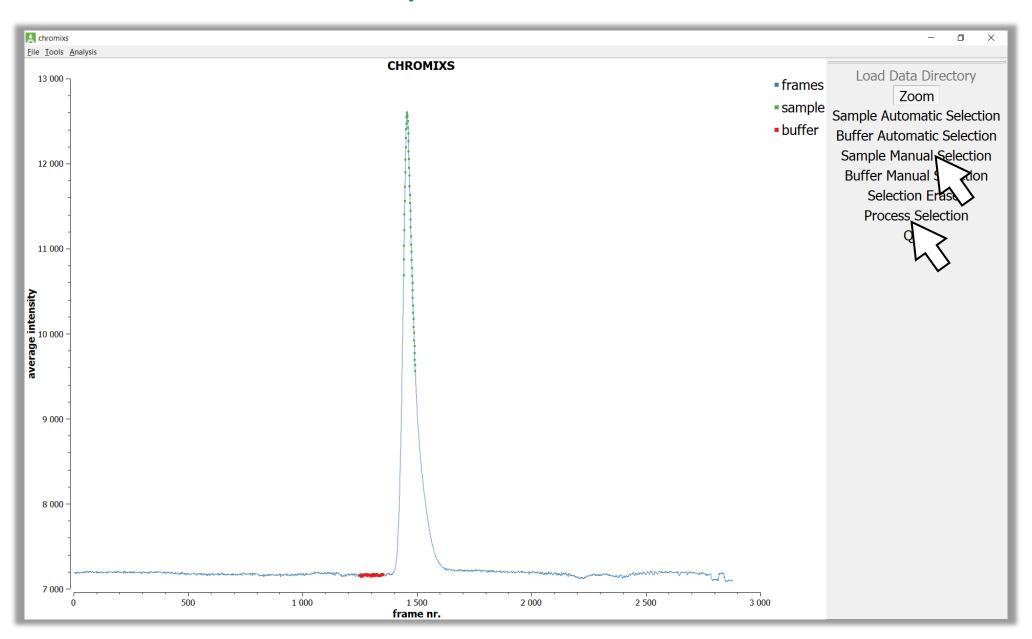




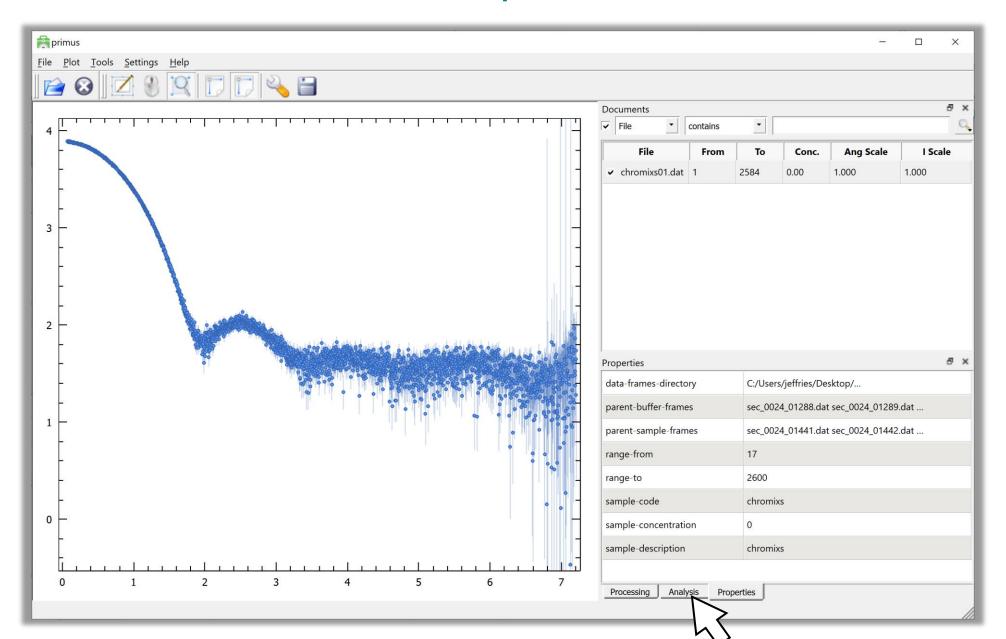






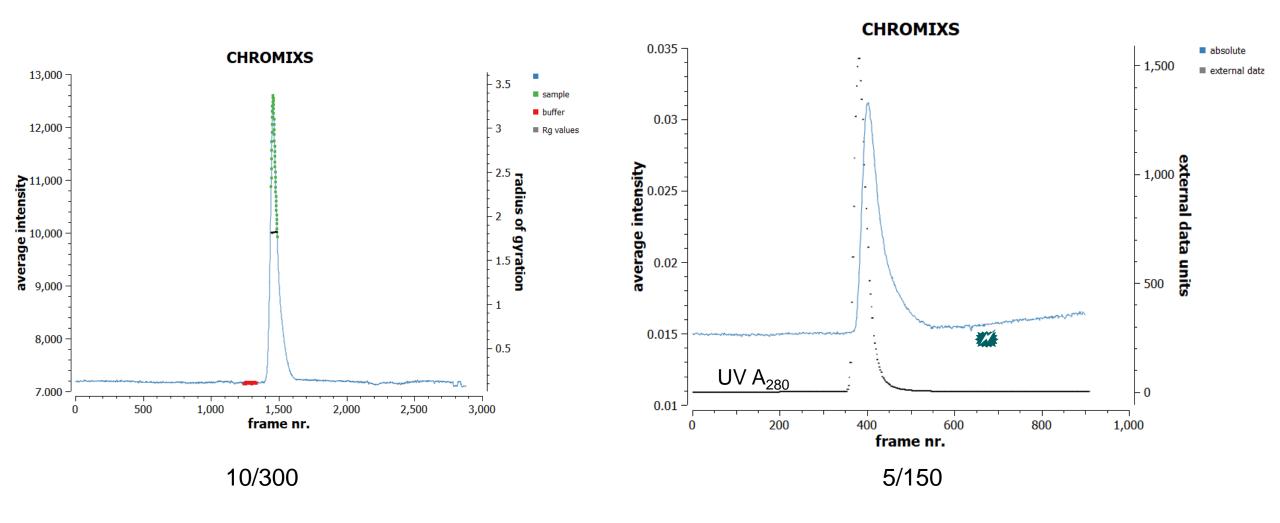


CHROMIXS opens PRIMUS



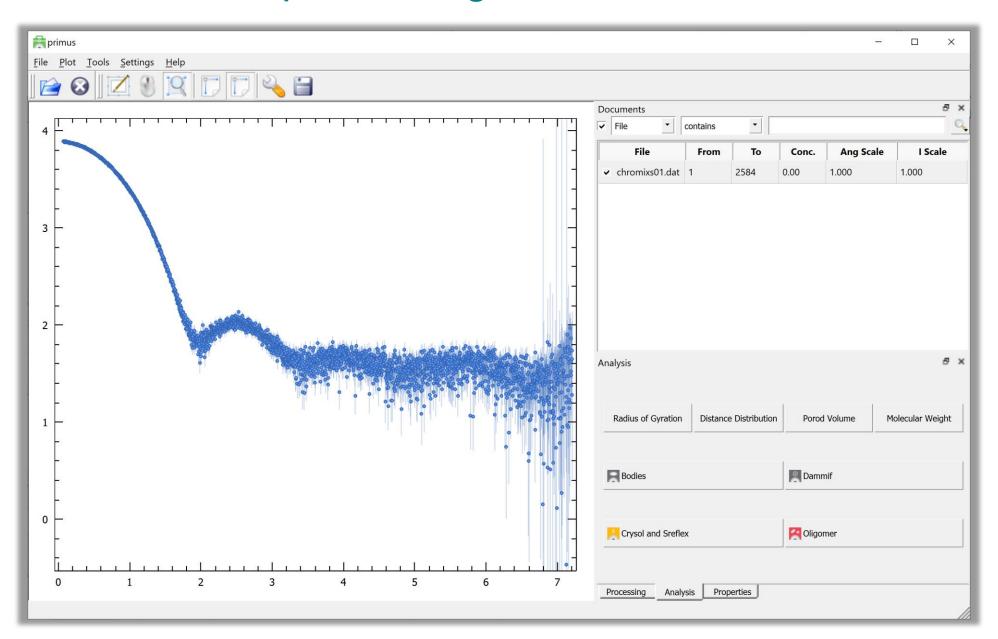
Stable size underneath the peak, comparison with UV

Correct background, monodispersion

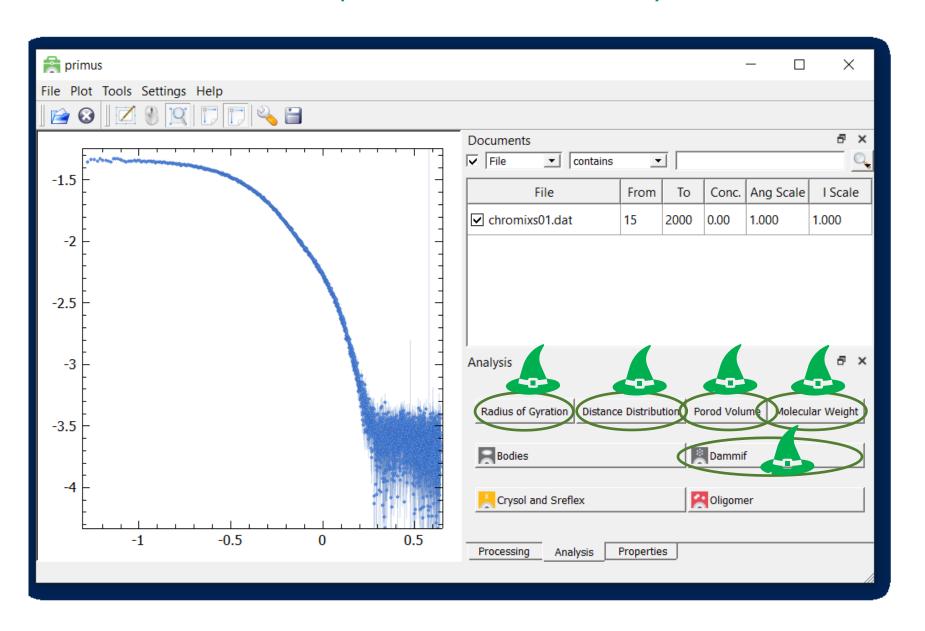


PRIMUS: parameters extraction and basic modeling

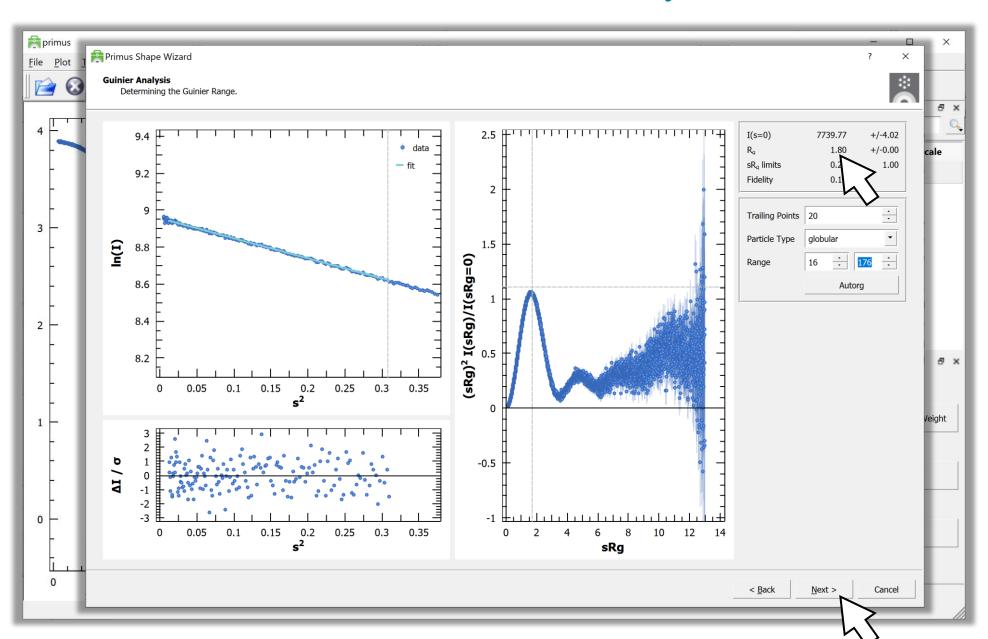
PRIMUS: processing our SEC-SAXS curve



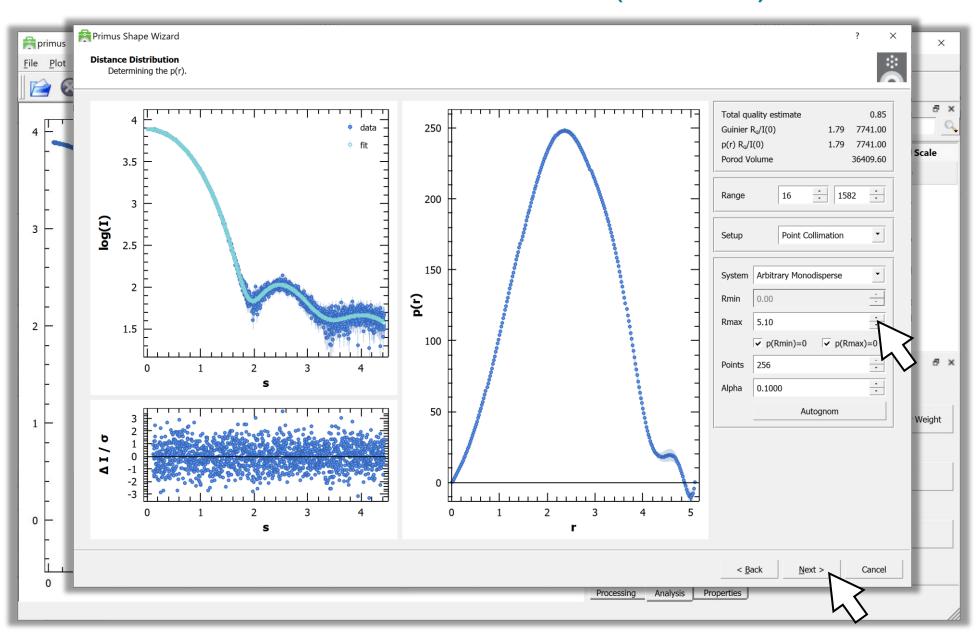
Most Used (and Most Useful) Wizards



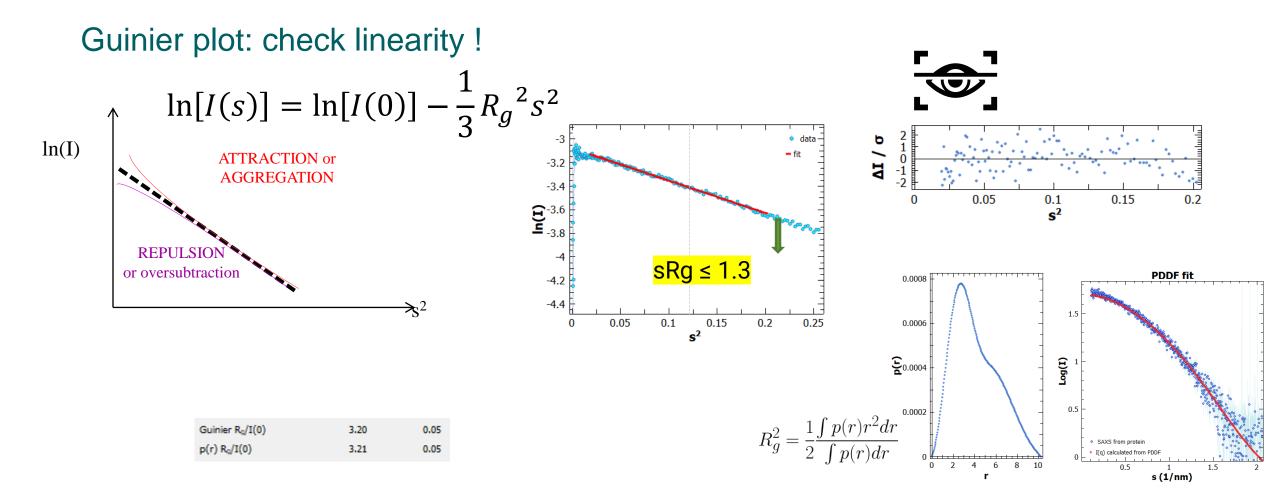
PRIMUS Guinier Analysis



Distance Distribution (GNOM)



Rg from Guinier Plot vs from P(r)

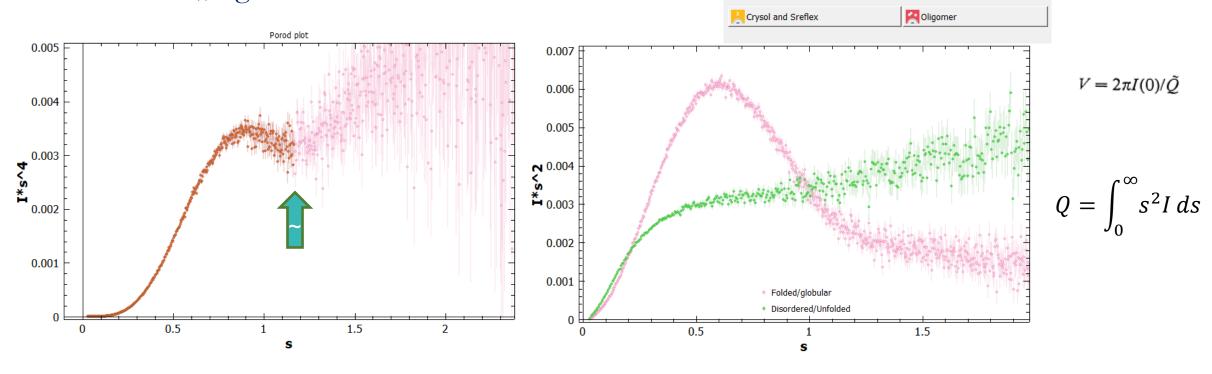


Compare Rg (Guinier) vs Rg (PDDF): if Rg Guinier blatantly > ... aggregates

Porod volume Vp

Distance Distribution | Porod Volume | Molecular Weight

- Depends on a good background subtraction!
- For proteins MW~0.625*Vp NAs MW~Vp (check consistency with theoretical one!)
- May "look smaller" for disodered molecules
- ➤ DATPOROD or GUI wizard
- > Evtl.from "regularized" curve from PDDF



Molecular mass (Mr, MW...)

- > MW from I(0) [I(s) or PDDF-regularization]: a physical measurement
 - calibration: H2O (T!), BSA
 - ~10% accuracy for ,,clean" globular protein data
 - Accuracy crucially dependent on CONCENTRATION I(0)/c
 - n/a if data were manually scaled (e.g. Bkg correction, c inaccurate)
 - Sensitive to aggregation/structure factor



- > SEC-SAXS MW from I(0): matching A280 (...) otherwise n/a
- > Other MW metrics e.g. MoW, s&s, Vc, Qp, Vp...
 - Bayesian MW: e.g.

Bayesian Inference

MW Estimate [Da] 62350

MW Probability [%] 70.38

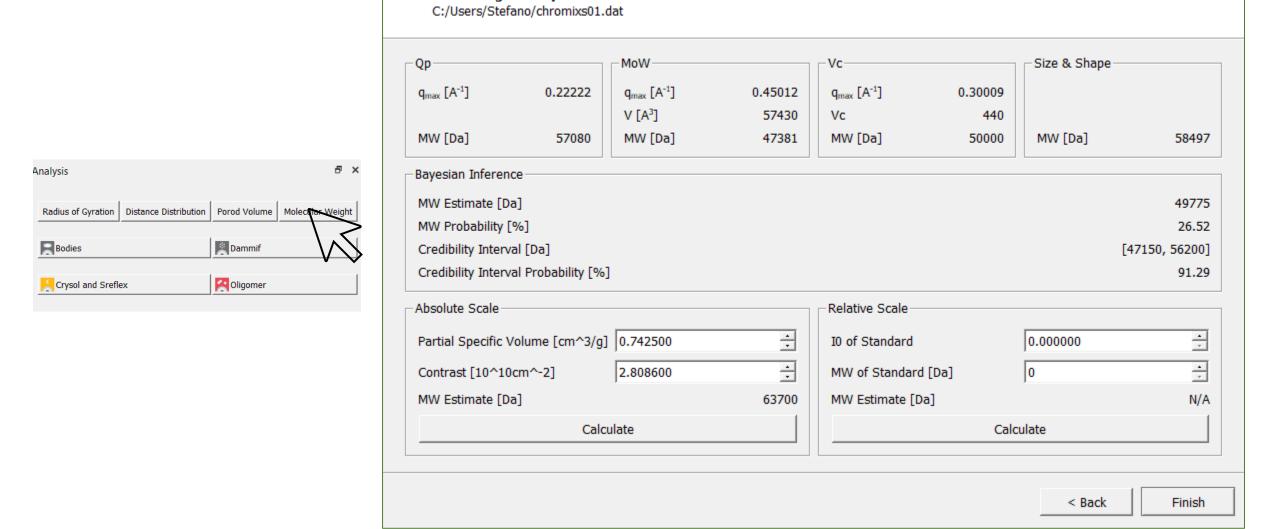
Credibility Interval [Da] [60200, 66250]

Credibility Interval Probability [%] 92.44

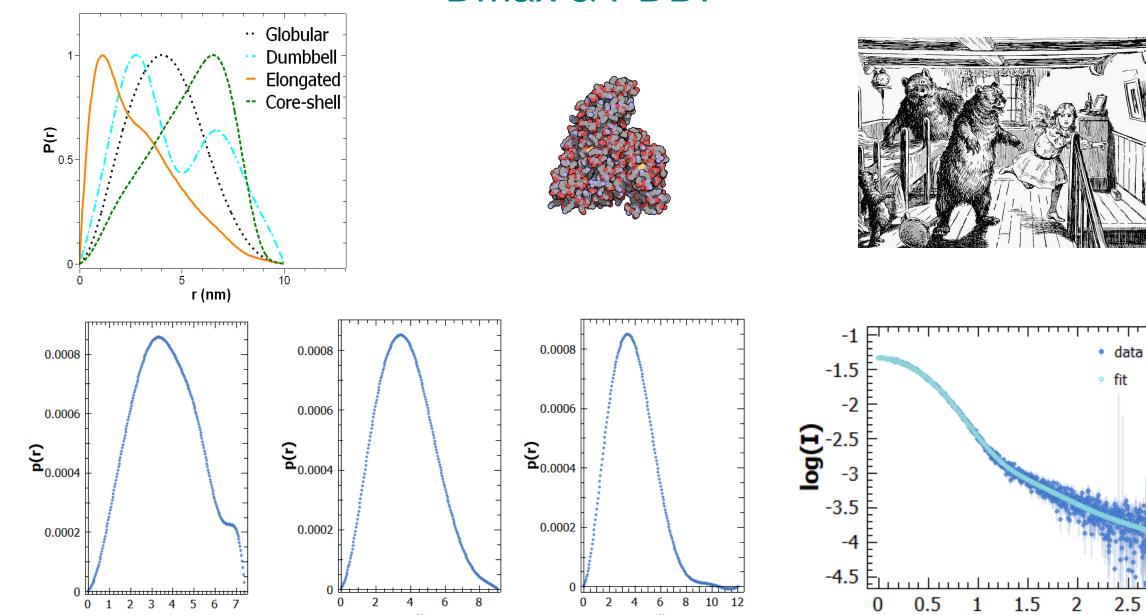
PRIMUS MW wizard

Primus Molecular Weight Wizard

Molecular Weight Analysis



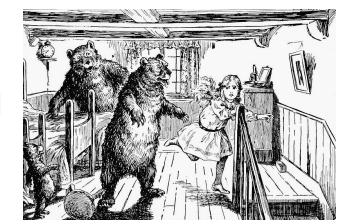
Dmax & PDDF

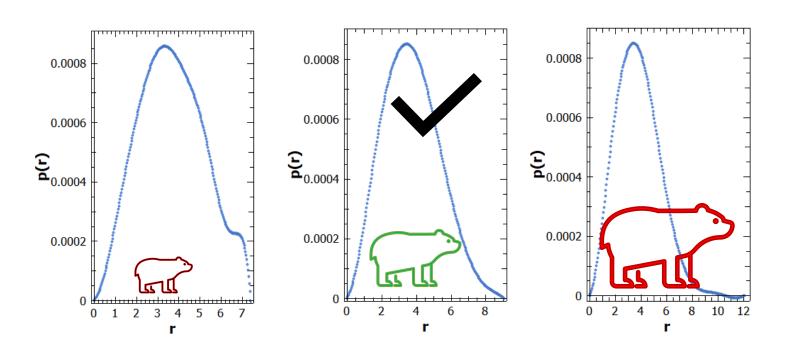


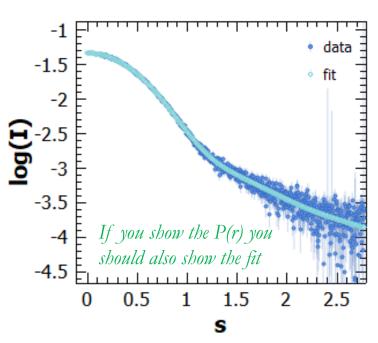
Dmax & PDDF

Warning: D_{max}*s_{min} greater than PI.

- P(r) for a compact folded globular protein
- Unless "extrapolated" Dmax $\leq \pi/s_{min}$
- "Perceptual criteria"
- Unlikely to be more precise than 0.5 nm
- Dmax and fit only as good as the PDDF

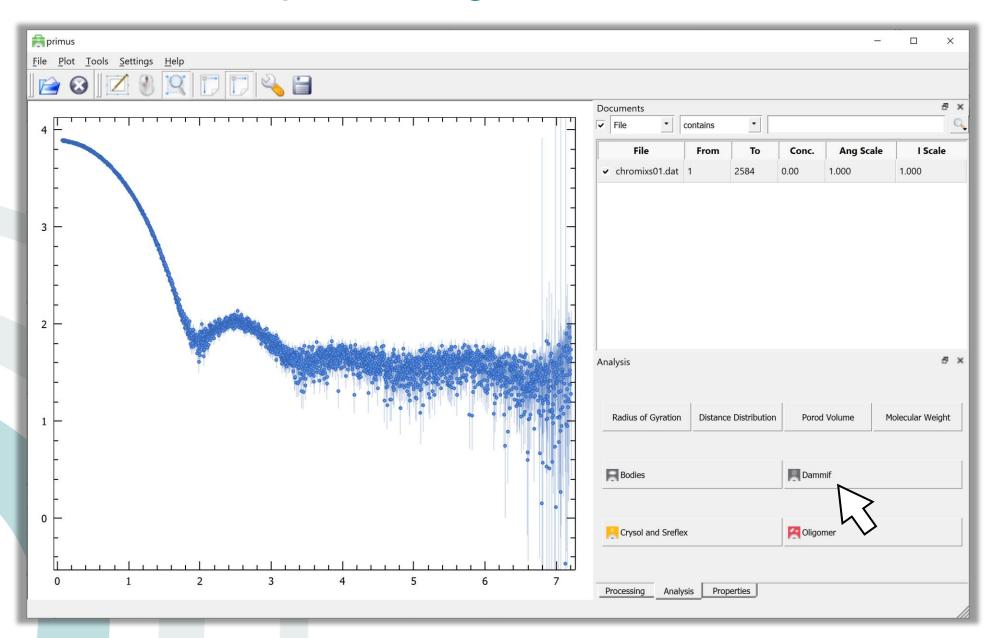




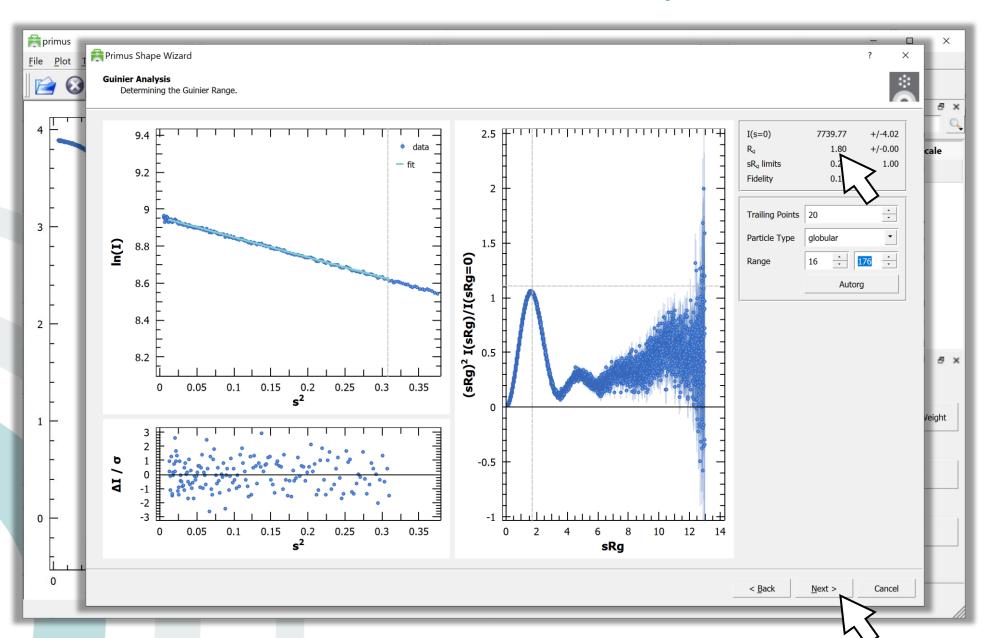


With a good P(r), we can start the modeling

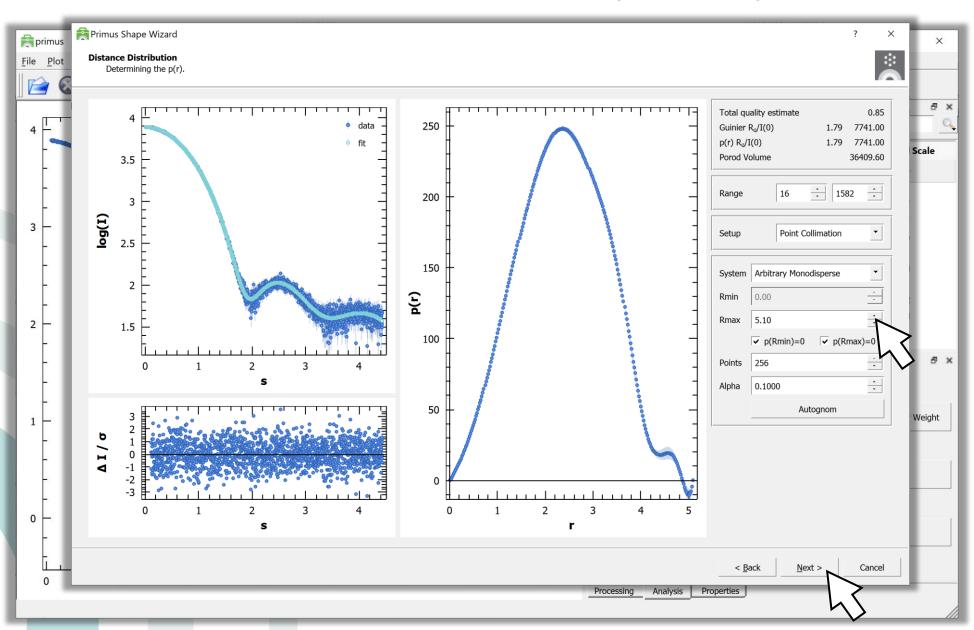
PRIMUS: processing our SEC-SAXS curve



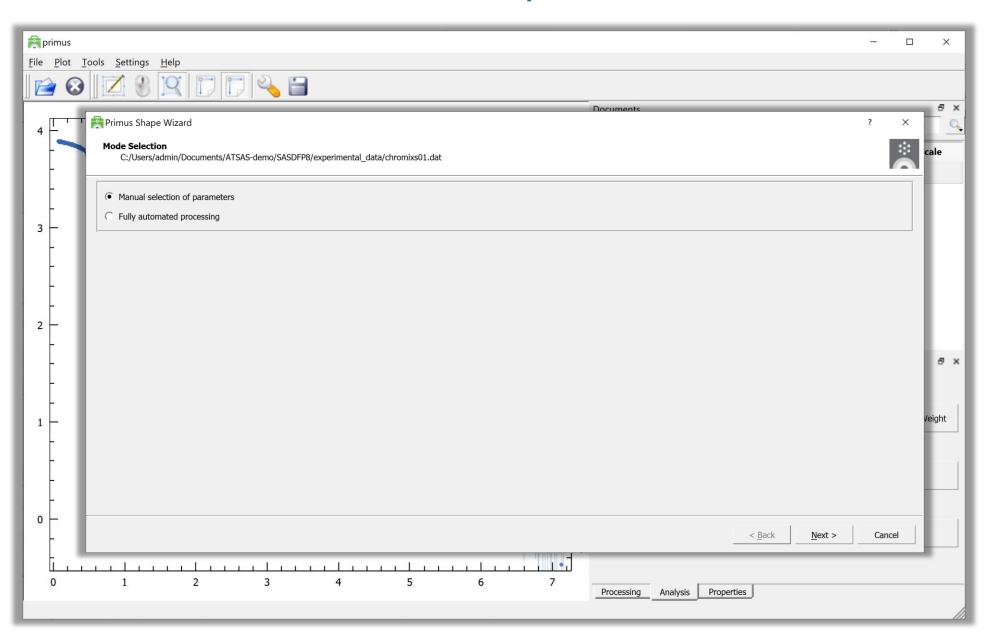
PRIMUS Guinier Analysis



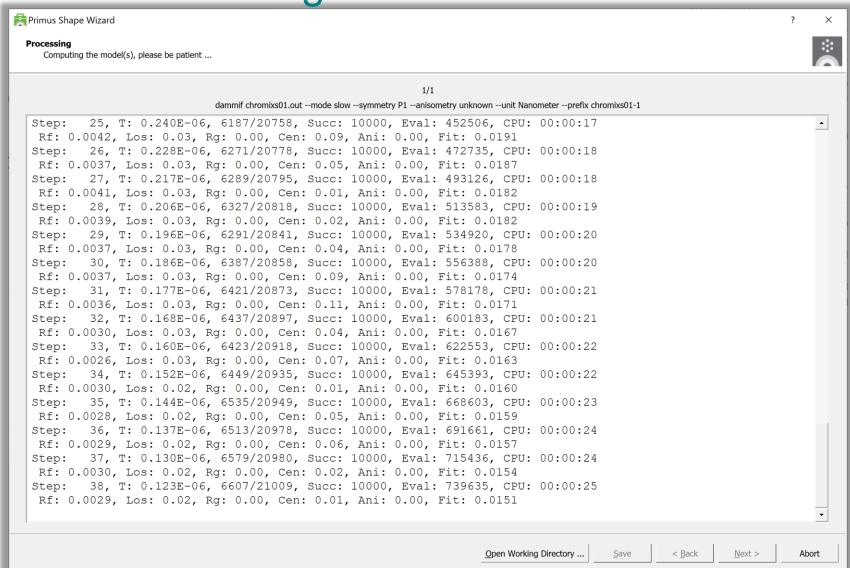
Distance Distribution (GNOM)



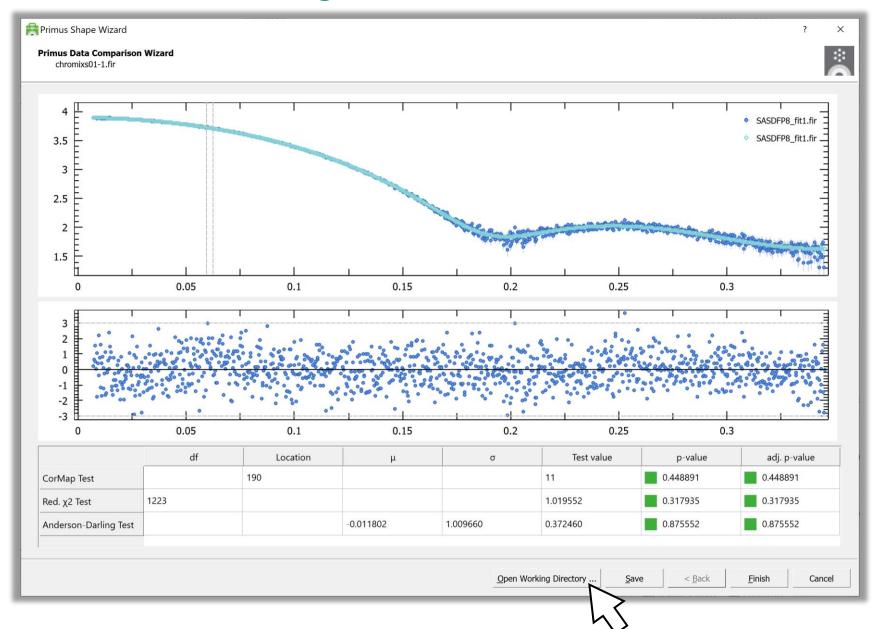
PRIMUS Shape Wizard



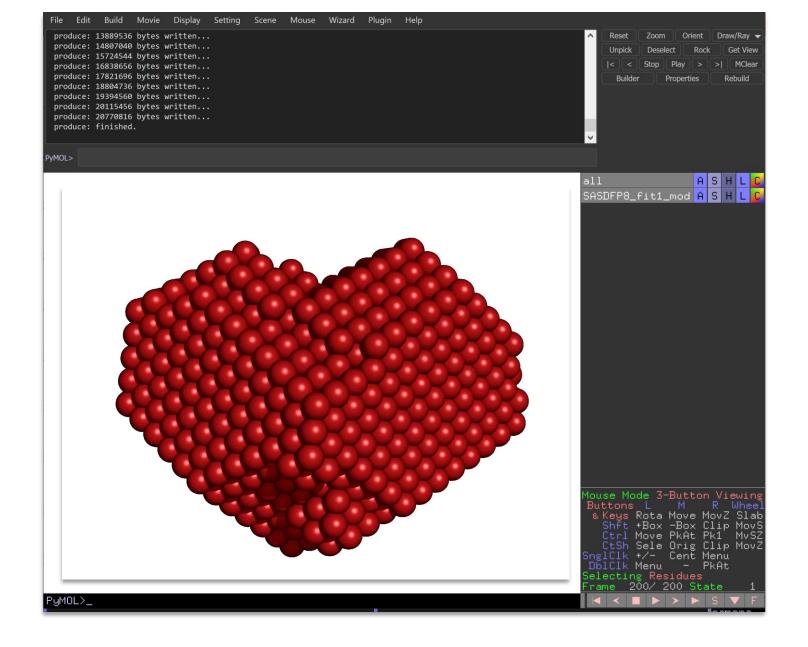
Running DAMMIF/DAMMIN



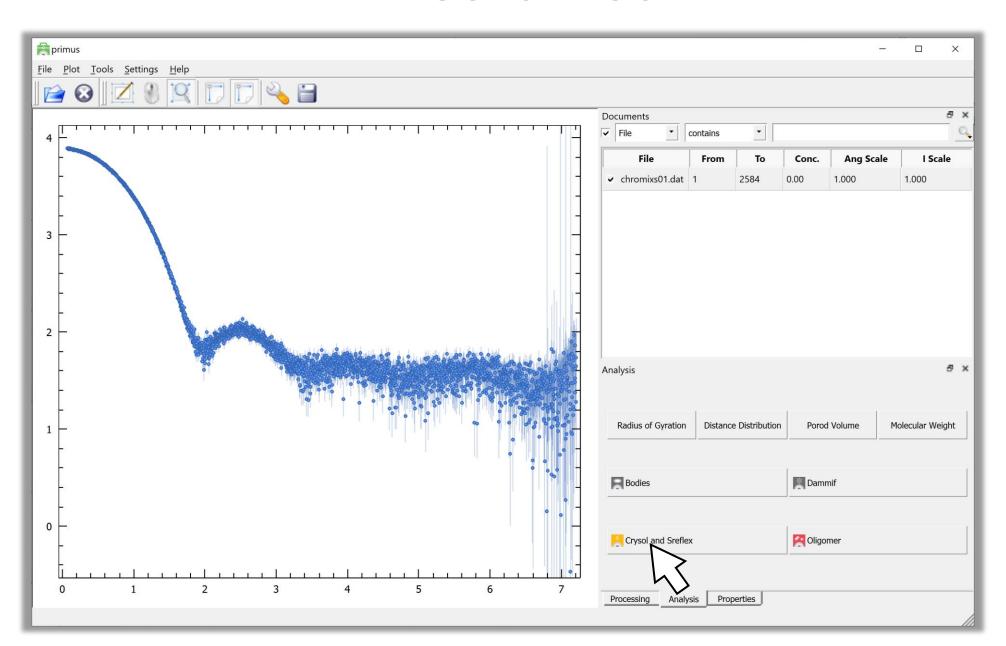
Running DAMMIF/DAMMIN



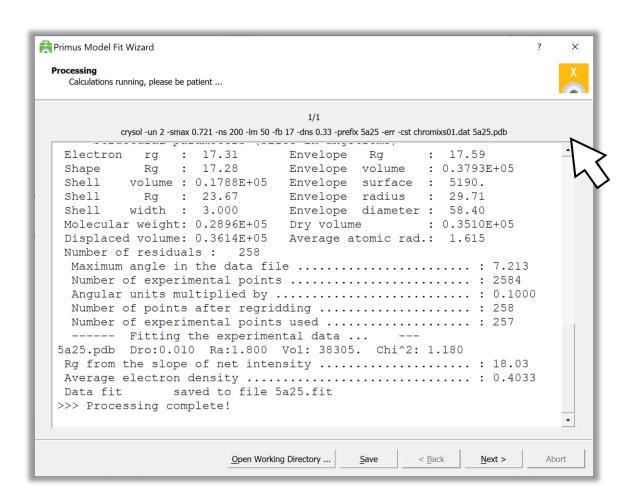
Let's now open one of the 10 models....



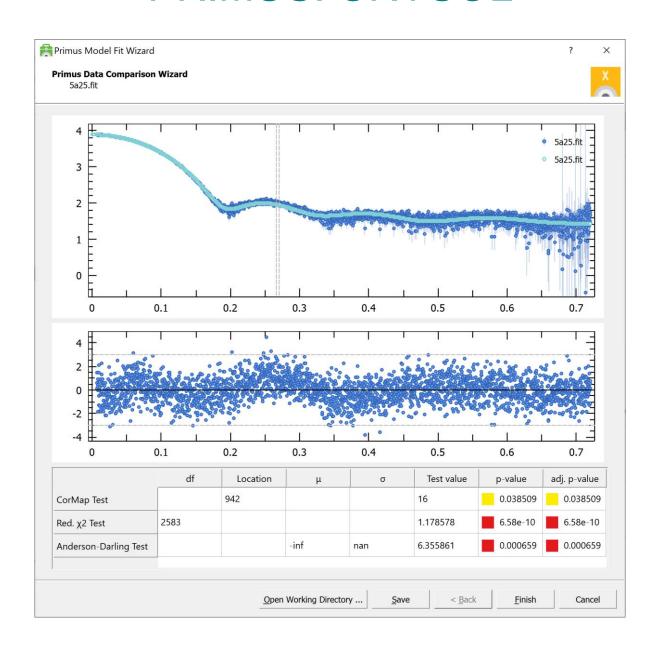
PRIMUS: CRYSOL

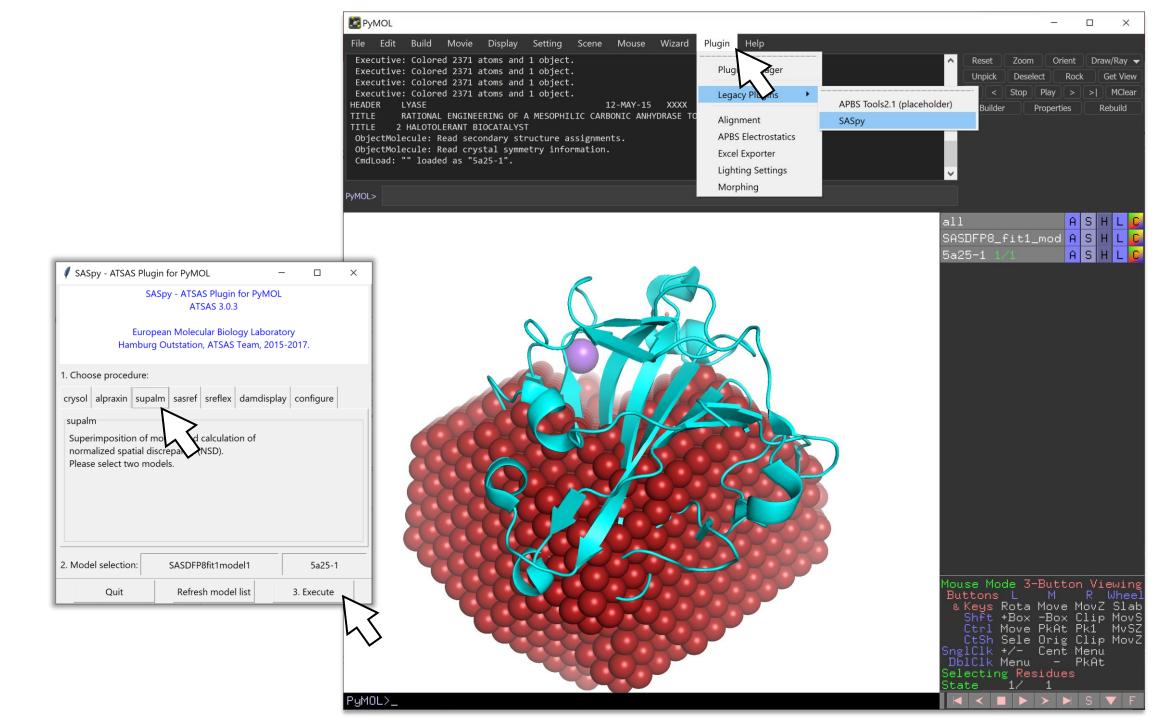


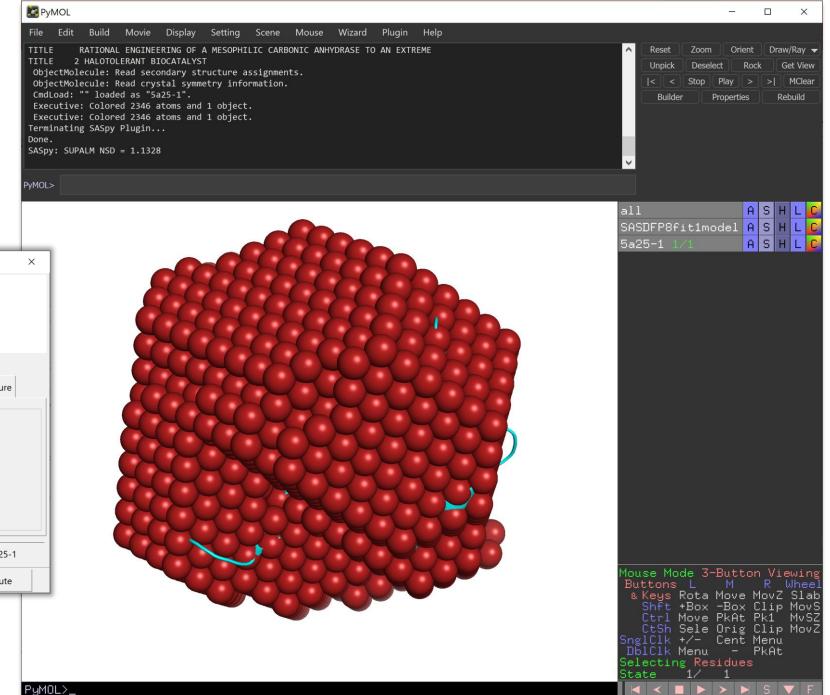
PRIMUS: CRYSOL

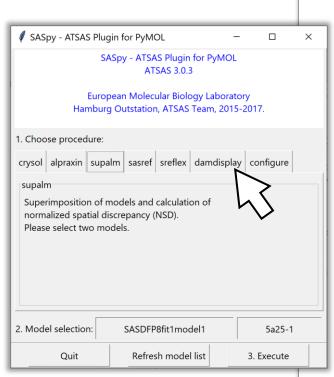


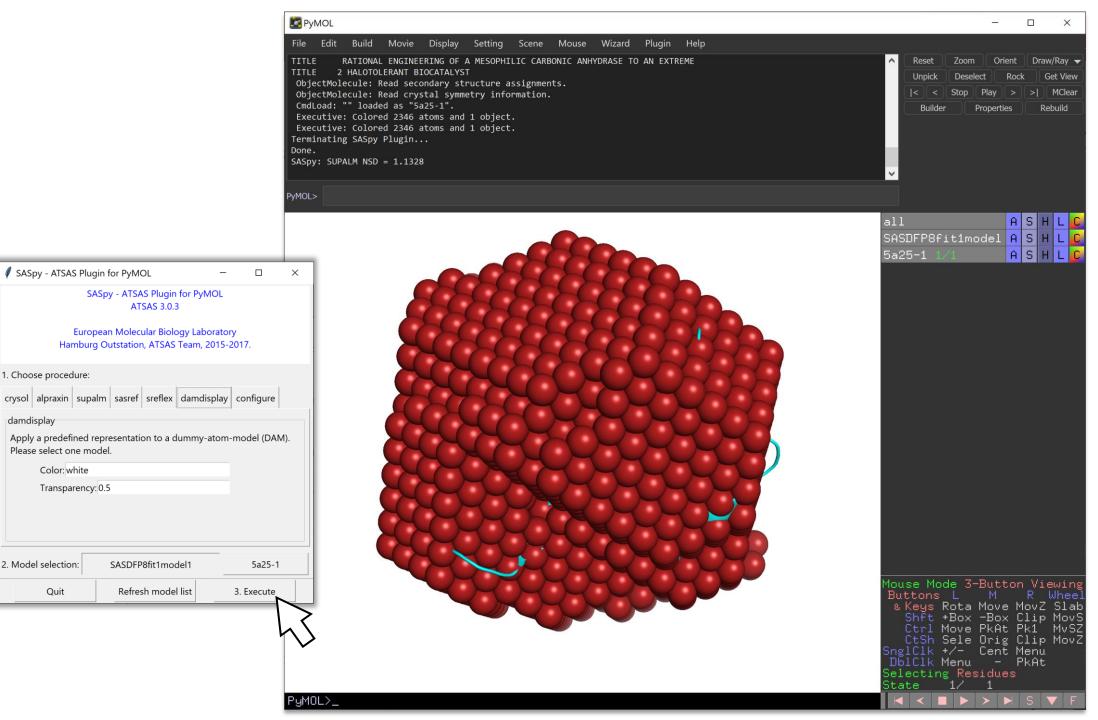
PRIMUS: CRYSOL











SASpy - ATSAS Plugin for PyMOL

1. Choose procedure:

Please select one model. Color: white Transparency: 0.5

damdisplay

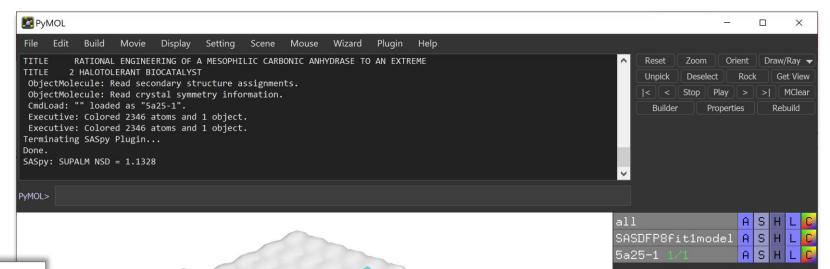
2. Model selection:

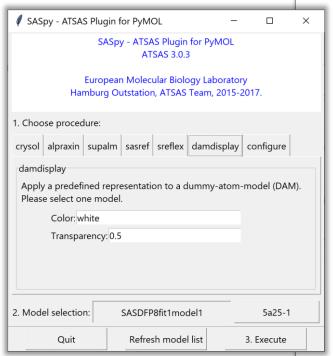
Quit

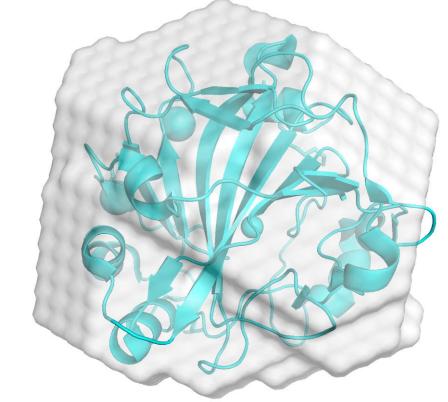
ATSAS 3.0.3

SASDFP8fit1model1

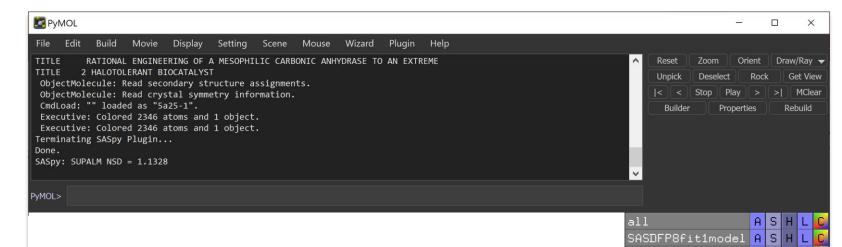
Refresh model list

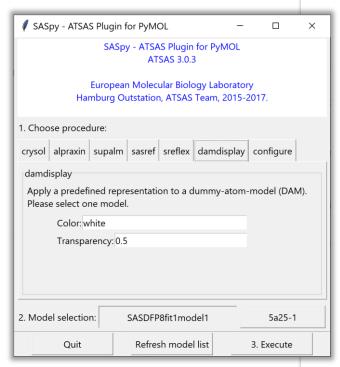


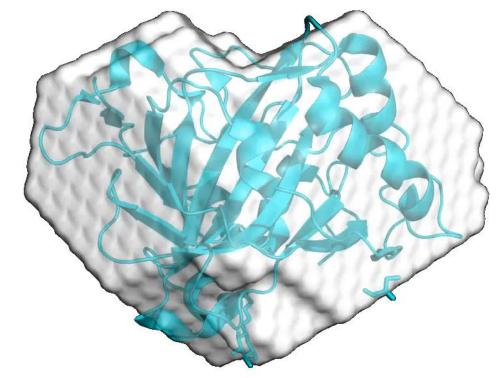














5a25-1

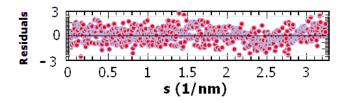
Metrics for fit quality

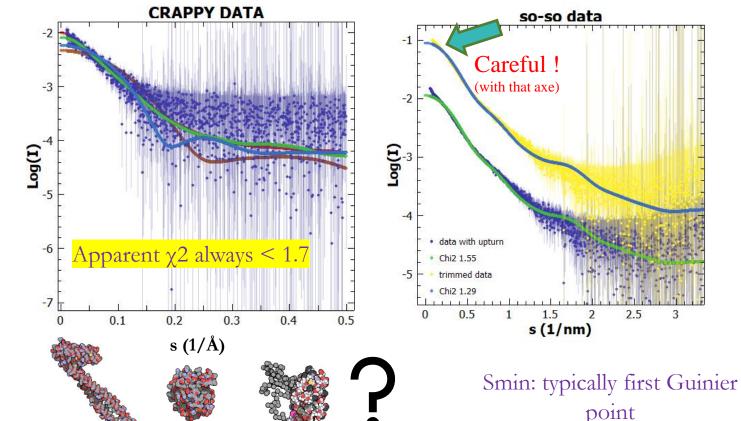
Most commonly used: $\chi 2$ (but...sensitive to I error estimate) LOOK to the data (noise level/point dispersion, regions with larger disagreement/residuals) Consider proper data range

$$\chi^{2} = \frac{1}{n-1} \sum_{i=1}^{n} \left(\frac{I_{exp}(s_{i}) - I_{calc}(s_{i})}{\sigma_{i}} \right)^{2}$$

Cormap - p

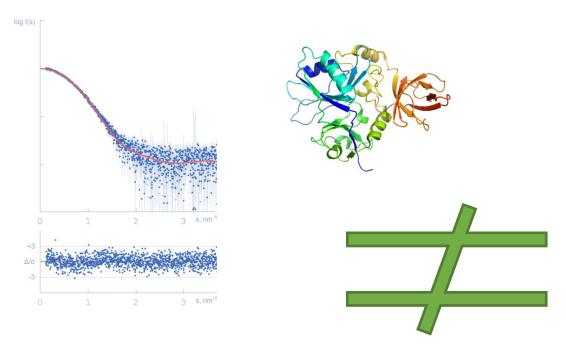
$$Vr, \chi^2_{free}$$



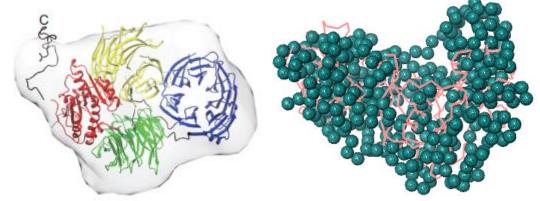


Terminology: ab initio models, fitting

- Fit to the data
 - Agreement or discrepancy
 Between ab initio or molecular model
 and the SAXS data



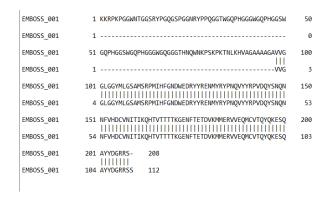
- Overlap to the ab initio reconstruction
 - Visual comparison between models
 maximizing the overlap (not a real fit)

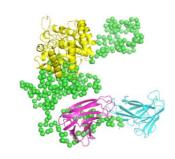


Not fitting?

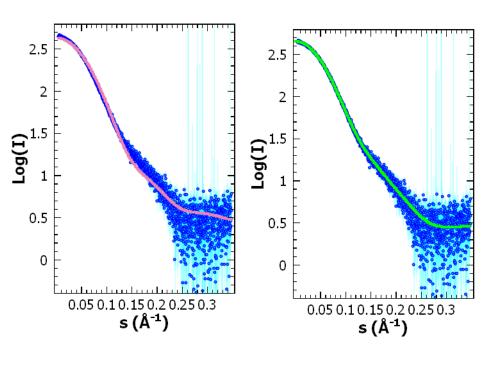
That's good information too!

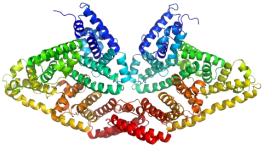
- Consider data (range, structure factor, subtraction)
- Consider structure completeness
- Consider association state(s)







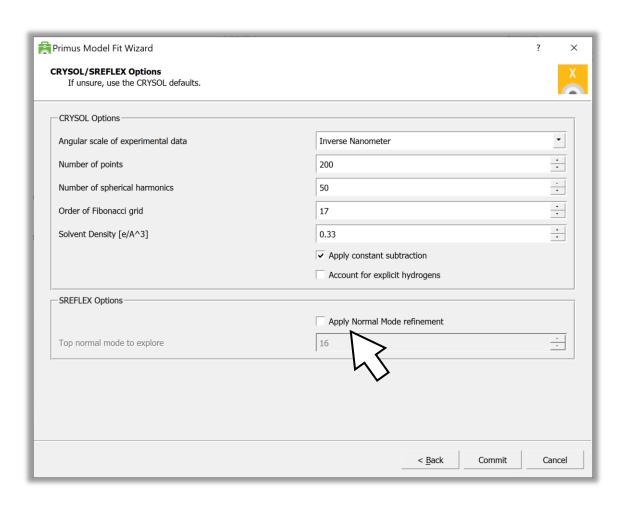




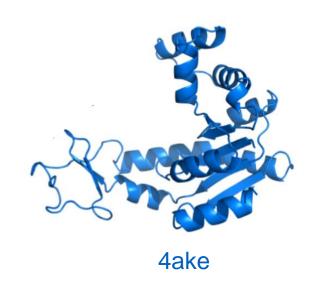
Not fitting and suspecting conformational change?

- Crystal contacts
- Structure only in complex with interaction partner
- Etc.

SREFLEX: flexible refinement

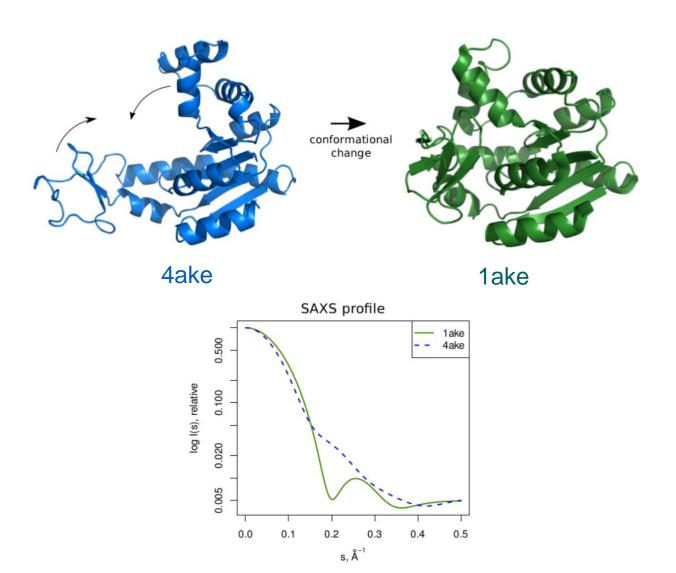


SREFLEX: refinement through flexibility



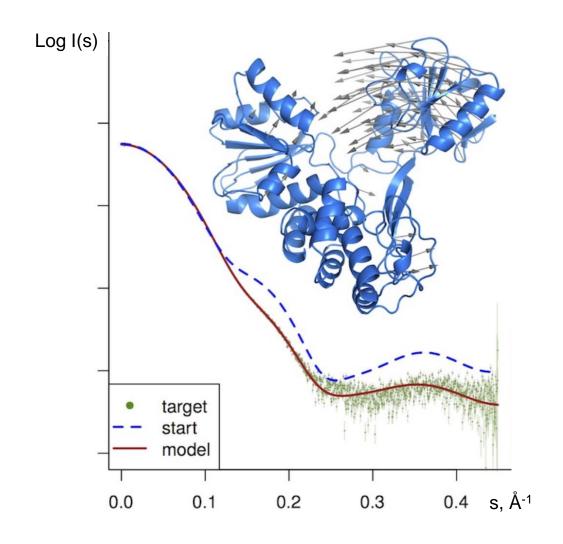
1ake

SREFLEX: refinement through flexibility





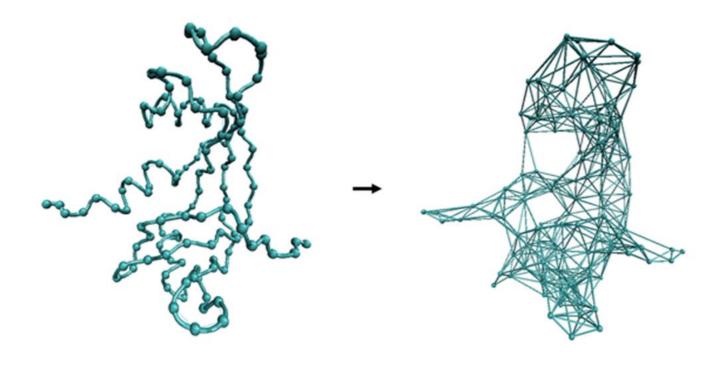
SAS REfinement through FLEXibility based on normal mode analysis



SREFLEX: Panjkovich A. and Svergun D.I. (2016) Phys. Chem. Chem. Phys. 18, 5707-5719



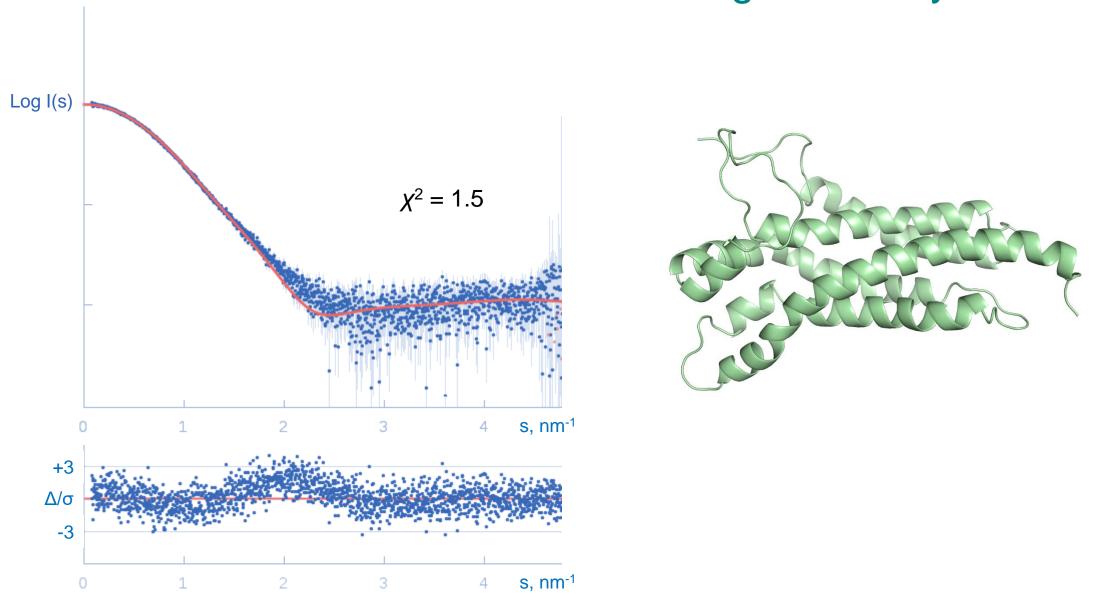
Estimating protein flexibility: normal mode analysis (NMA)



 $C\alpha$ trace

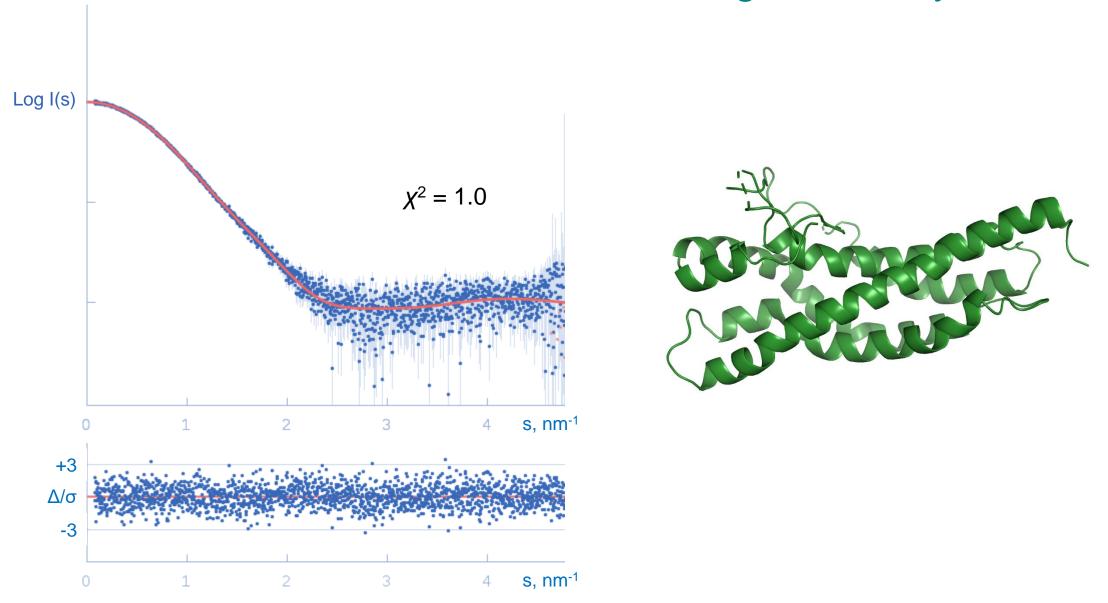
elastic network model

SREFLEX: refinement through flexibility



SASDC36 – Structural and functional dissection of the DH and PH domains of oncogenic Bcr-Abl tyrosine kinase

SREFLEX: refinement through flexibility



SASDC36 – Structural and functional dissection of the DH and PH domains of oncogenic Bcr-Abl tyrosine kinase

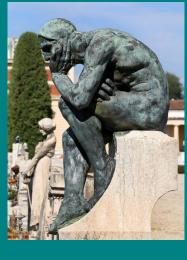
> HINTS:

Click your way around by trial and error

Look inside the files (open as text files .dat, .fir, .out etc.)!

Learn to use CLI

Look at fits visually and think (larger? smaller? etc.)



BI((O))SAXS

The original solution

Thanks for your attention: keep exploring the programs....



sdavela@embl-hamburg.de

